

Faculty-student matchmaking session: summer research in the biological sciences

**January 14, 2019
SSS 114**

**Sandy Chang, MD/PhD
Associate Dean of STEM Education and Undergraduate Research**

Performing cutting edge research is an integral component of Yale's undergraduate education



Yale Summer Fellowships

Yale College First Year Summer Research Fellowships

- Supports 10-12 weeks of natural science research in Yale labs. Fellowships pay \$430/week., max 10 weeks funded.
- Last year, 83% first years received funding.
- As part of their research experience, students participate in a weekly class called “Entering Research”
- Fellowships due March 7.

STARS Summer Research fellowship

- Supports 30 rising sophomores doing 10 weeks of research in Yale labs. All expenses paid, plus \$2,500 stipend.
- Last year 20% of students who applied received this fellowship.
- Students must first identify a mentor (Jan 28th). Selected students have to submit a proposal (Feb 27).

Yale College Dean's and Rosenfeld Research Fellowships

- Supports sophomores and juniors doing research in Yale labs. Fellowships pay \$430-450/week, max 10 weeks funded.
- Last year, 60% sophomores and juniors received funding.
- Fellowships due Feb. 21.

Student-Faculty Matchmaking

- Students pick two potential mentors for further interactions.
- Faculties choose the students they want to mentor, and work with them to craft a good research proposal.
- Students who do not match with mentors, and faculties who do not get students should contact me for an additional round of matching (electronically).
- For students on financial aid, you can use your domestic summer award (DSA) to help fund your summer research. Check “yes” on the DSA box in the fellowship application.
- Much more info at <https://science.yalecollege.yale.edu/>

Faculty sciences

Bergwitz Lab information

Lab Head (top photo third from left):

Clemens Bergwitz, M.D.

website:

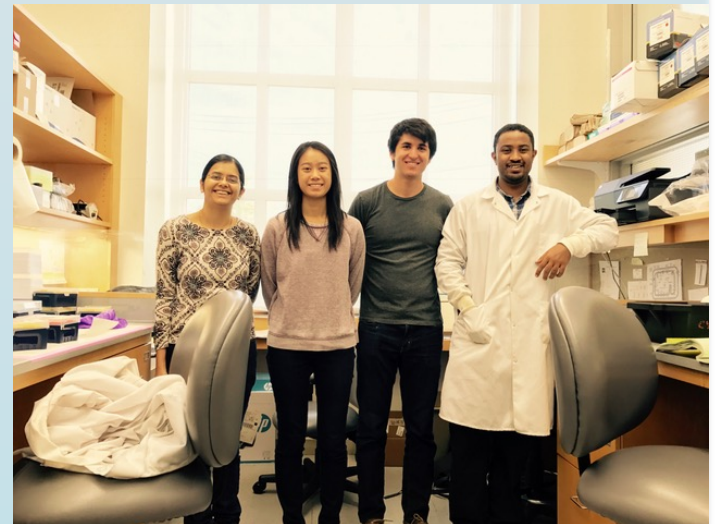
<http://medicine.yale.edu/lab/bergwitz/>

Associate Professor of Medicine
Yale University School of Medicine
Section Endocrinology and Metabolism
Anlyan Center (TAC), Office S117, Lab S110
1 Gilbert Street
New Haven, CT06519

Work Phone: 203-737-5450

Work Fax: 203-785-5535

email: clemens.bergwitz@yale.edu



Summer projects 2019

Our research focuses on inborn errors of phosphate metabolism and the endocrine regulation of phosphate homeostasis with emphasis on the metabolic and homeostatic effects of phosphate.

Students will independently conduct one the following projects (2-3 slots available):

- Determine bioavailability of dietary phosphate in mice (RNA extraction from mouse gut&kidneys, qRT-PCR based gene expression analysis)
- Determine ATP content and mitochondrial function in skeletal muscle lacking phosphate-transporters (cell culture, seahorse assays, luciferase-based ATP/ADP measurements in cells and muscle tissues)
- Determine mineral metabolism of mice lacking phosphate transporters in skeletal muscle or bone cells (ELISA and colorimetric assays to measure blood phosphate, FGF23, urine phosphate and creatinine)
- Determine the function of mutant phosphate transporters (adenoviral transduction of cells and life mice followed by Western blot analysis to measure transporter protein expression)
- Translational research to improve care of patients affected by Hereditary Hypophosphatemic Rickets with Hypercalciuria (HHRH) (review patient records with referring physicians, genotyping PCR and GENEIOUS software analysis of Sanger sequencing reads)

Mentorship

Jan-Feb 2019: Help with applications for summer student scholarships

June-August 2019: Provide written protocols and one-on-one supervision by PI or postdoc during 12 week summer internship, which enables students to independently conduct a small project, which will - if successful - earn them a co-authorship in a line of research that is close to publication

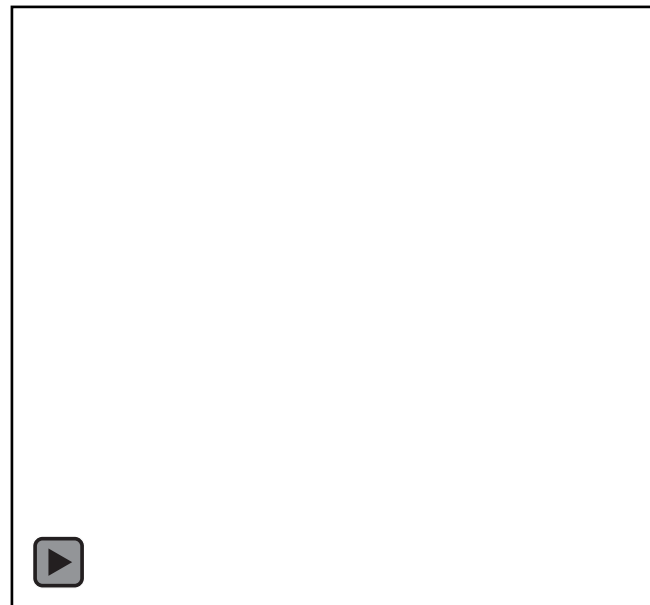
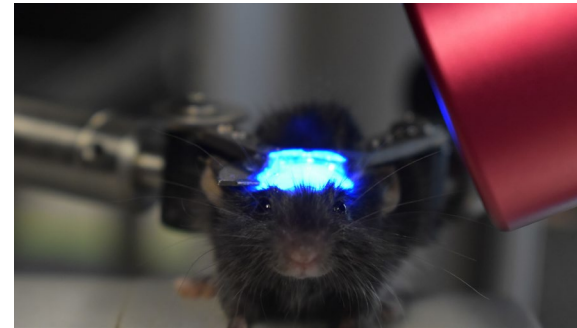
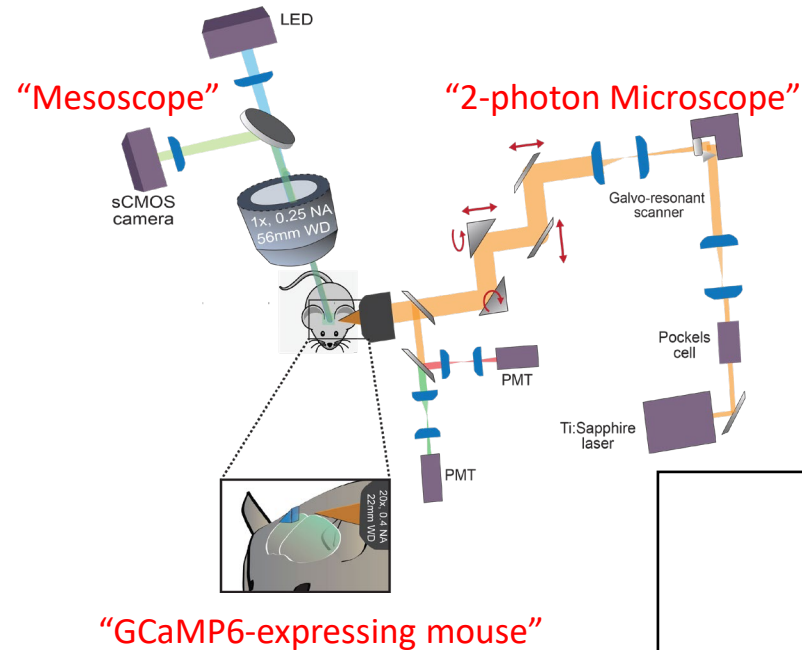
Sept 2019: Opportunity to continue independent research on a 12-hr/week basis during the semester

Track record 2014-2018: 10/10 undergraduate students received scholarship funding, 9/10 students published (3 in peer-reviewed journals, 1 as lead author, 9 abstracts in international conferences), 9/10 students continued in the semester or returned next summer (3 senior thesis, 1 postbachelaurate, 6 semester projects, see Lab website for names and contact info).

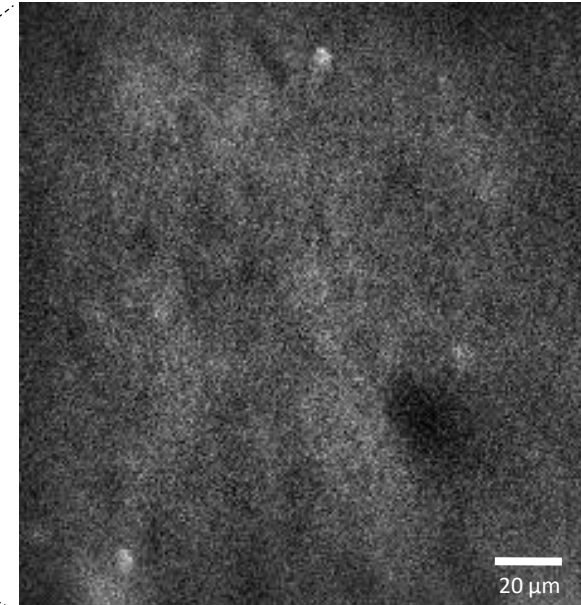
Imaging neuronal activity in the rodent brain

Laboratory of Michael Crair

Laboratory of Michael Higley



Mouse neocortex



Layer 2/3 Somatosensory Cortex

Presented by Daniel Barson
daniel.barson@yale.edu

Laboratory of Michael Crair

Department of Neuroscience

michael.crair@yale.edu

<https://medicine.yale.edu/lab/crair/>



Major Research Questions:

- How does spontaneous activity instruct the formation of sensory maps in the brain?
- How do sleep and other behavioral states contribute to the development of cortical circuits?
- How are different genetically-defined classes of neurons integrated into cortical circuits?
- How is the connectivity and function of neural circuits disrupted in neuropsychiatric disorders such as autism and schizophrenia?

Undergraduate Research Projects:

- Mesoscopic imaging of neural activity in mouse models of neuropsychiatric disease
- Virus-based anatomical tracing of developing brain circuits
- Analysis and computational modeling of cortical network dynamics based on recorded activity

Current students: Luke Newell (NSCI major, development of cortical modulation by behavioral state)
Seneca Oxendine (NSCI major, disruption of cortical activity in autism mouse models)

Laboratory of Michael Higley

Department of Neuroscience

m.higley@yale.edu

higleylab.org

@mjhigley



Major Research Questions:

- How are excitatory and inhibitory synaptic input integrated by single neurons?
- How are different types of neurons wired into large-scale networks?
- How do neurons and circuits in visual cortex mediate sensory perception and behavior?
- How is the connectivity and function of neural circuits disrupted in neuropsychiatric disorders such as autism and schizophrenia?

Undergraduate Research Projects:

- Implement novel genetic strategies for creating new mouse disease models
- Fluorescent imaging of neuronal structure across development and in disease
- Analysis of behavior in mouse models
- Fluorescent imaging of neural activity *in vivo*
- Analysis and computational modeling of cortical network dynamics based on recorded activity

Current student: Natasha Zaliznyak (MBB major, viral methods for genetic labeling of single neurons)

Enrique M. De La Cruz and his Awesome Research Team

enrique.delacruz@yale.edu

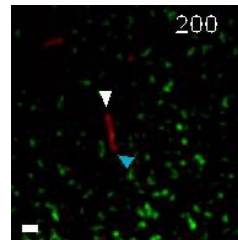


Enrique M. De La Cruz and his Awesome Research Team

enrique.delacruz@yale.edu



Actin filament assembly and cell motility

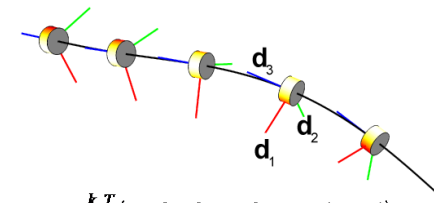
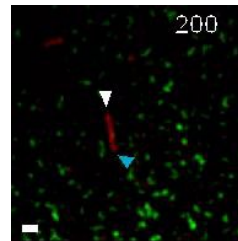


Enrique M. De La Cruz and his Awesome Research Team

enrique.delacruz@yale.edu



Actin filament assembly and cell motility



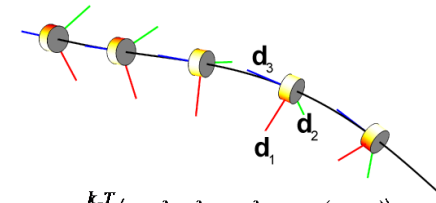
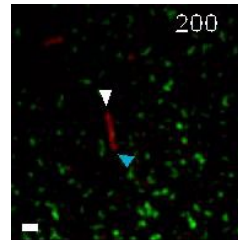
$$E_{\text{elastic}} = \frac{k_B T}{2} \left(L_p (\kappa_1^2 + \kappa_2^2) + L_T \kappa_3^2 + 2L_{TB} \kappa_3 (\kappa_1 + \kappa_2) \right)$$

Enrique M. De La Cruz and his Awesome Research Team

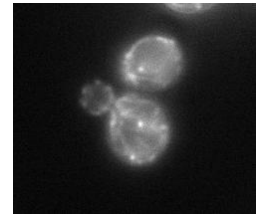
enrique.delacruz@yale.edu



Actin filament assembly and cell motility



$$E_{elastic} = \frac{k_B T}{2} \left(L_p (\kappa_1^2 + \kappa_2^2) + L_T \kappa_3^2 + 2L_{TB} \kappa_3 (\kappa_1 + \kappa_2) \right)$$

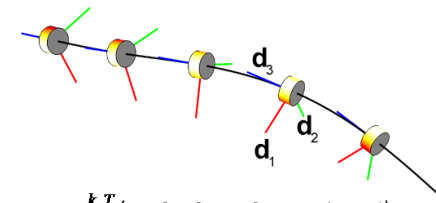
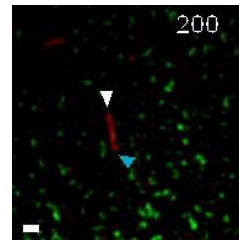


Enrique M. De La Cruz and his Awesome Research Team

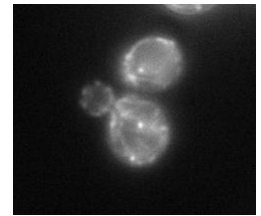
enrique.delacruz@yale.edu



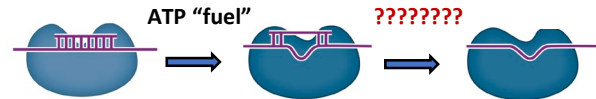
Actin filament assembly and cell motility



$$E_{elastic} = -\frac{k_B T}{2} \left(L_p (\kappa_1^2 + \kappa_2^2) + L_T \kappa_3^2 + 2L_{TB} \kappa_3 (\kappa_1 + \kappa_2) \right)$$



RNA helicase enzymes

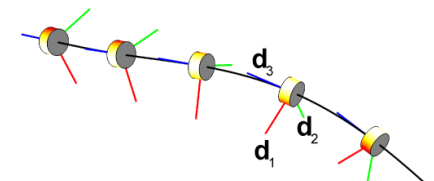
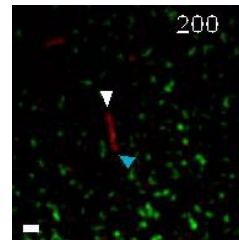


Enrique M. De La Cruz and his Awesome Research Team

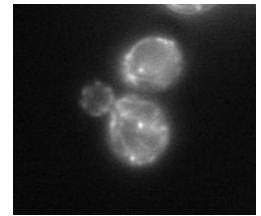
enrique.delacruz@yale.edu



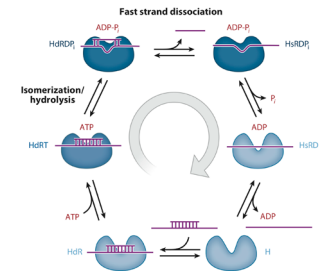
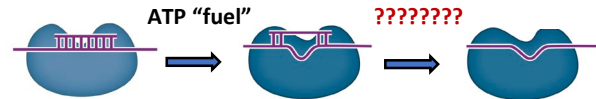
Actin filament assembly and cell motility



$$E_{elastic} = \frac{k_B T}{2} \left(L_p (\kappa_1^2 + \kappa_2^2) + L_T \kappa_3^2 + 2L_{TB} \kappa_3 (\kappa_1 + \kappa_2) \right)$$



RNA helicase enzymes

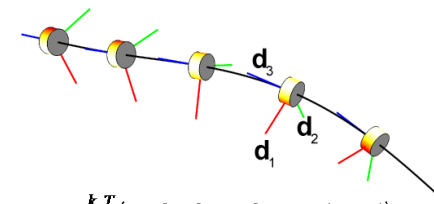
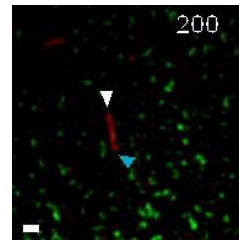


Enrique M. De La Cruz and his Awesome Research Team

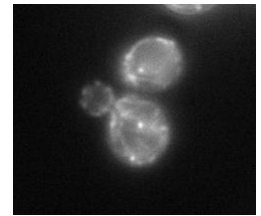
enrique.delacruz@yale.edu



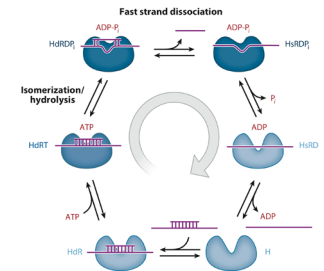
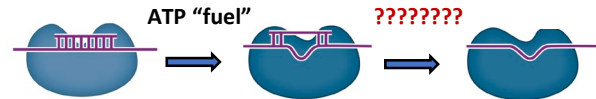
Actin filament assembly and cell motility



$$E_{elastic} = \frac{k_B T}{2} \left(L_p (\kappa_1^2 + \kappa_2^2) + L_T \kappa_3^2 + 2L_{TB} \kappa_3 (\kappa_1 + \kappa_2) \right)$$



RNA helicase enzymes



NPP signaling enzymes



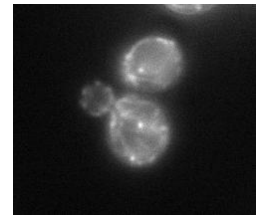
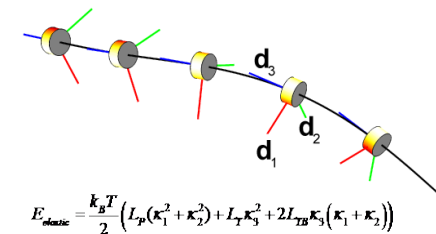
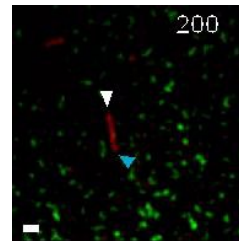
Demetrios Braddock
M.D./Ph.D.

Enrique M. De La Cruz and his Awesome Research Team

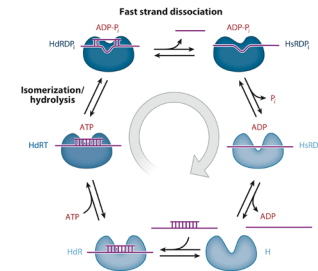
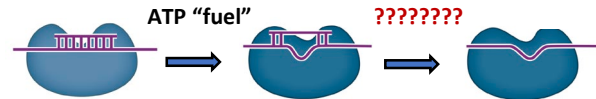
enrique.delacruz@yale.edu



Actin filament assembly and cell motility



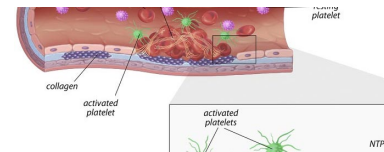
RNA helicase enzymes



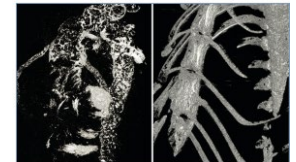
NPP signaling enzymes



Demetrios Braddock
M.D./Ph.D.



Blood clotting



Calcification of heart and vessels

What could you do with us this summer?



Marisa Michalchik

What could you do with us this summer?

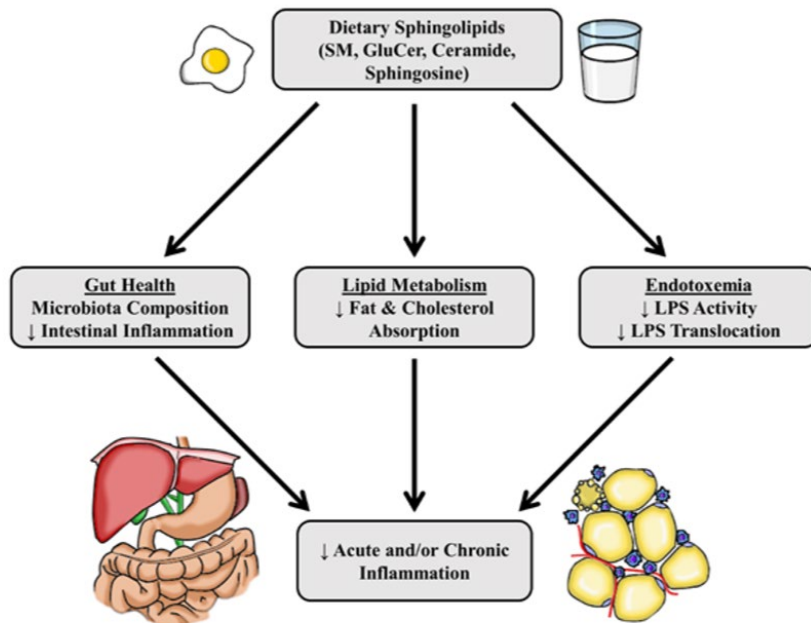
ENPP7 – a lipid cleaving enzyme in your gut



Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut

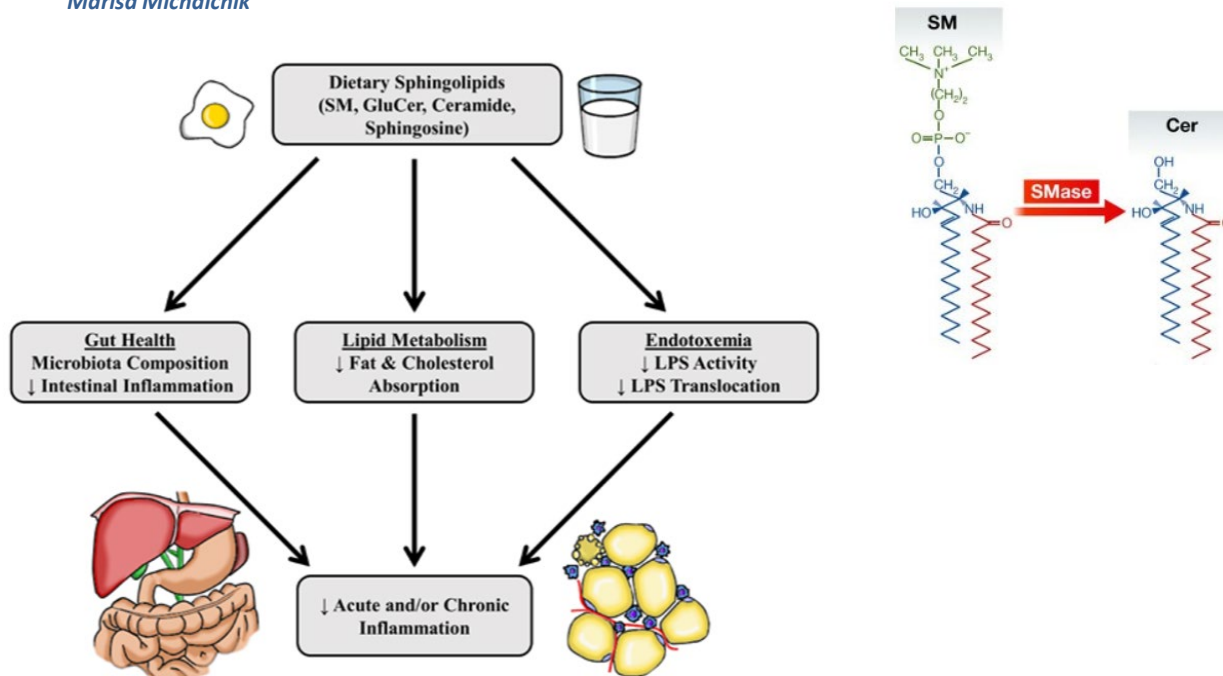




Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut

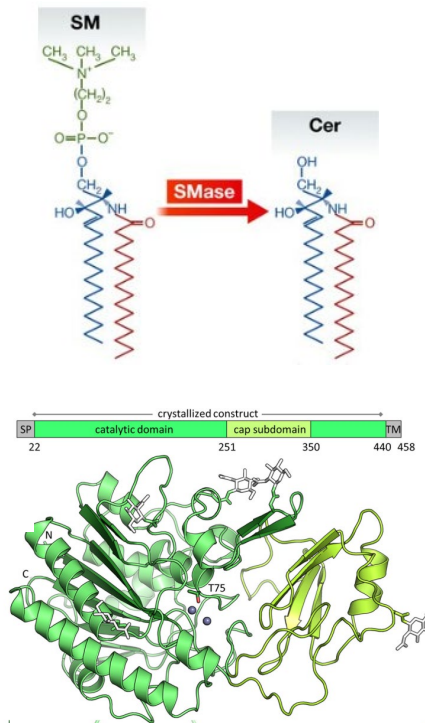
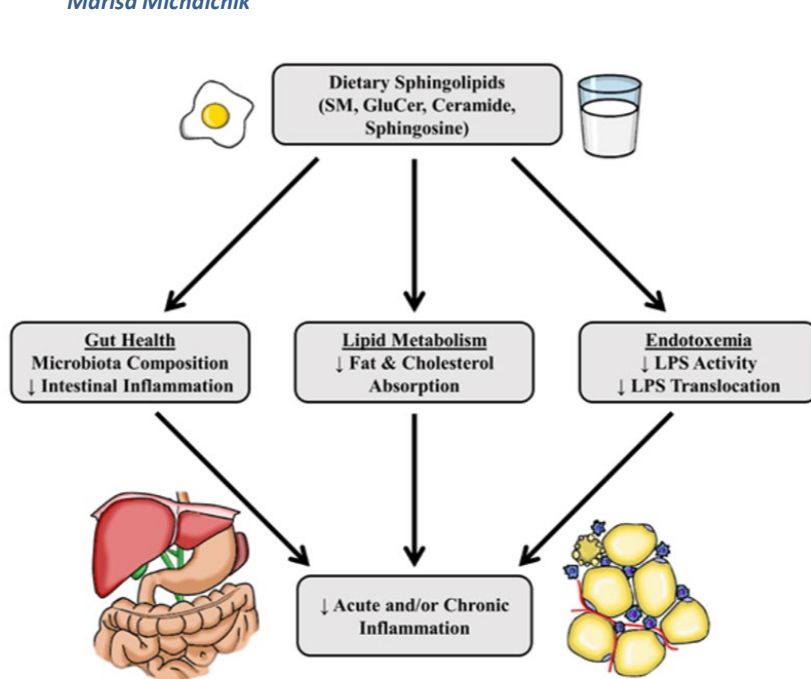




Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut

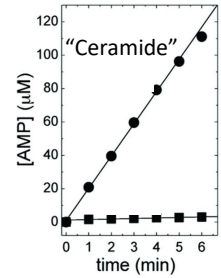
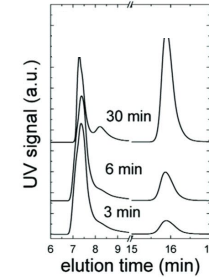
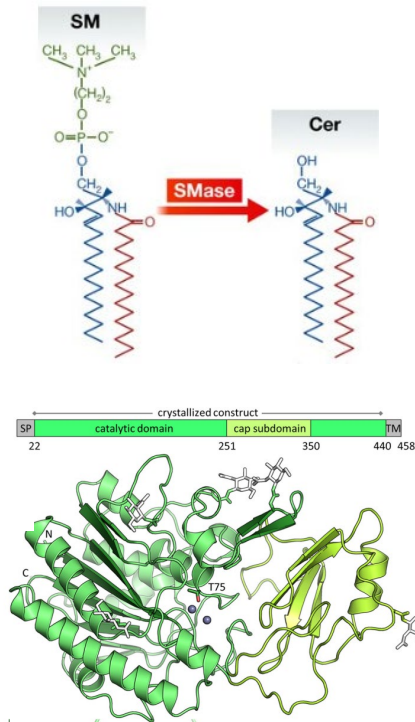
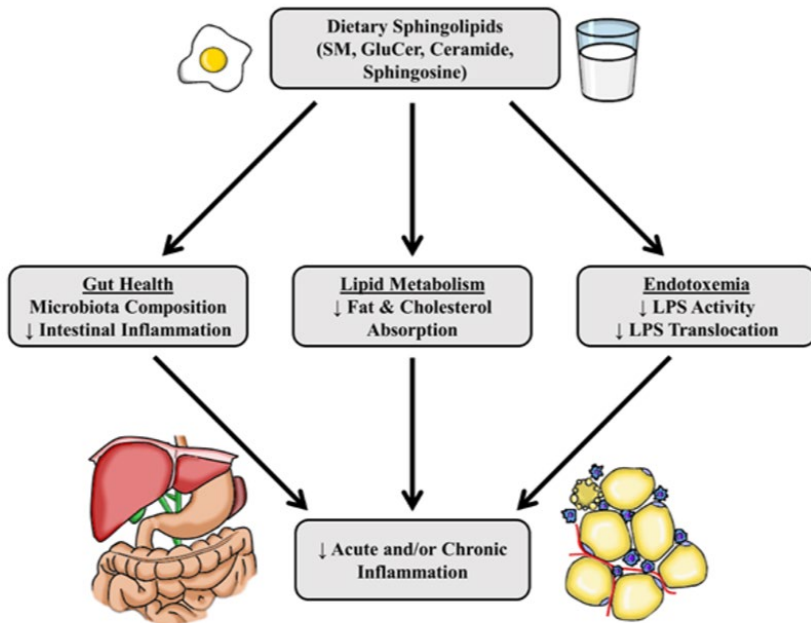




Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut

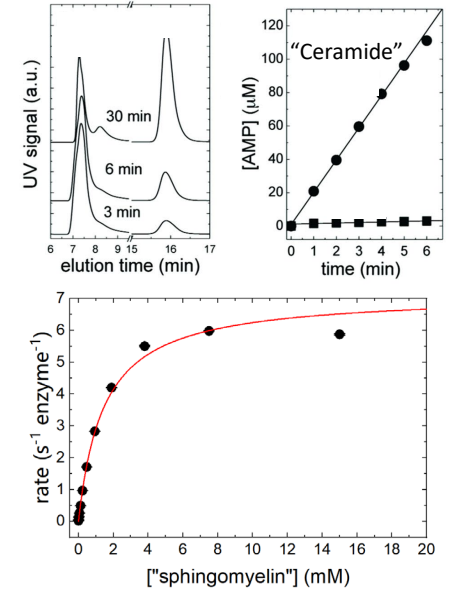
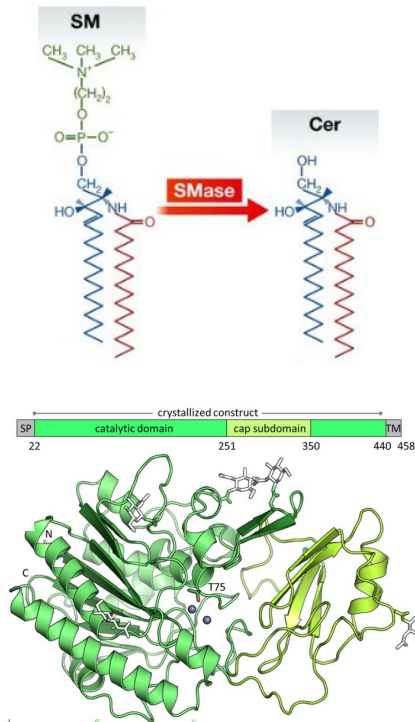
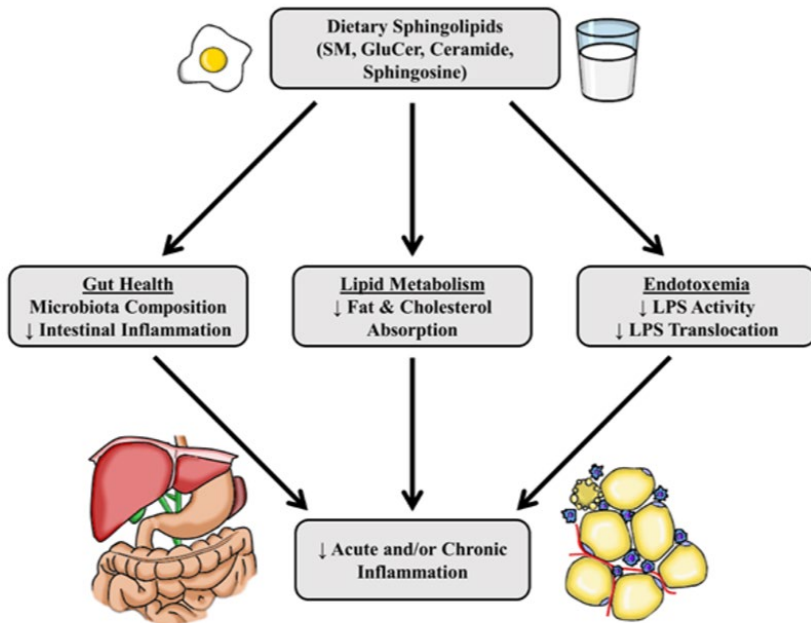




Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut

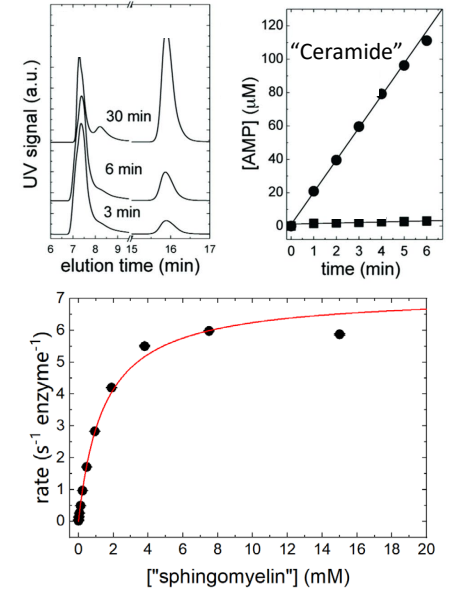
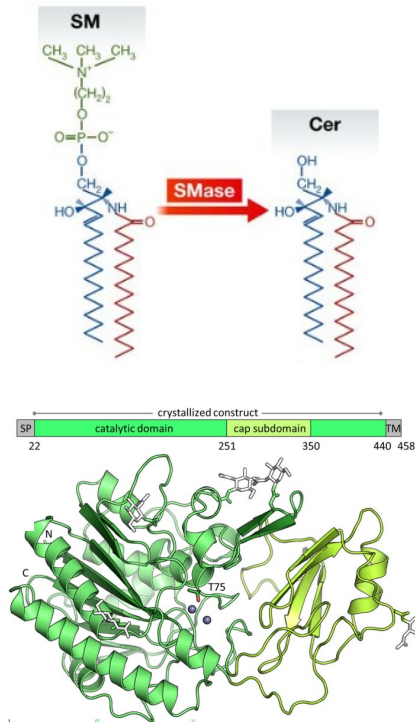
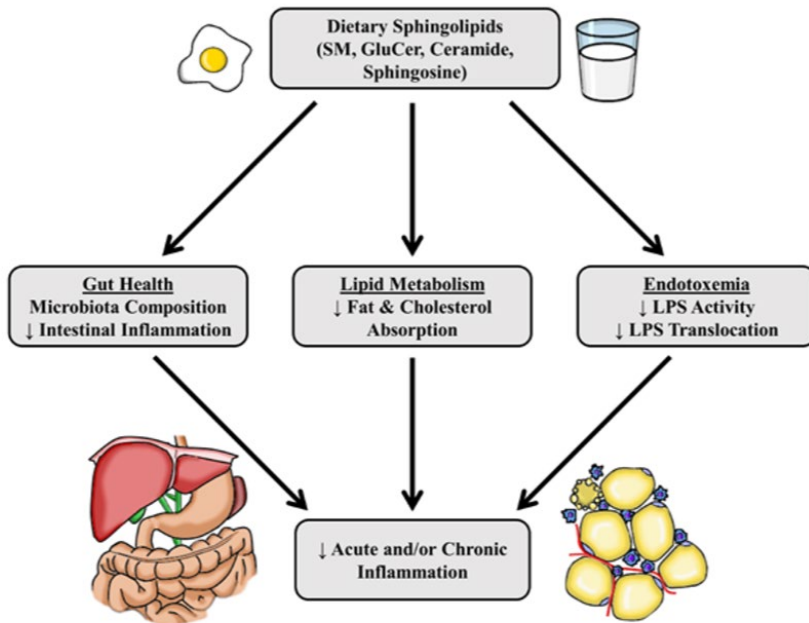




Marisa Michalchik

What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut



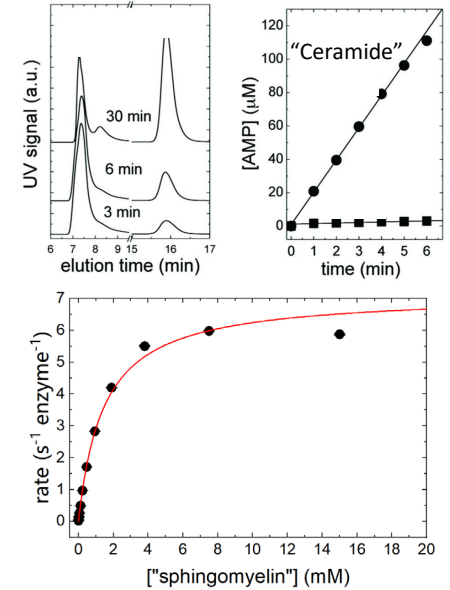
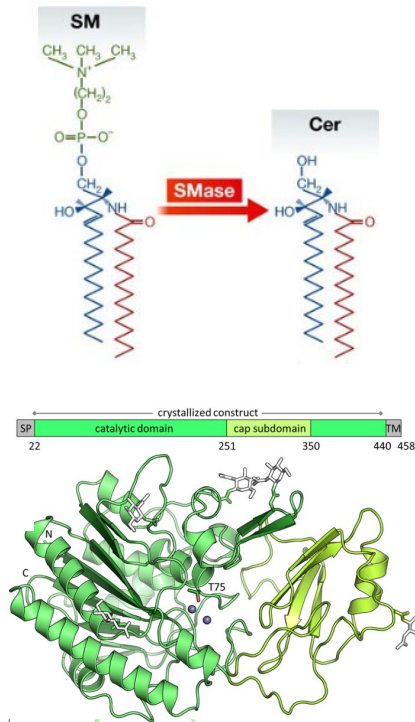
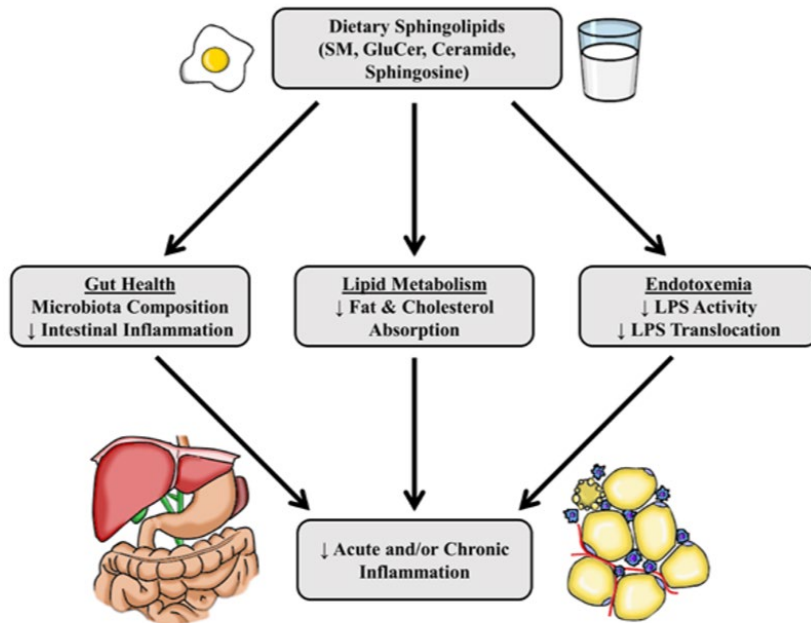
-Enzymology (Biochemistry!)



Marisa Michalchik

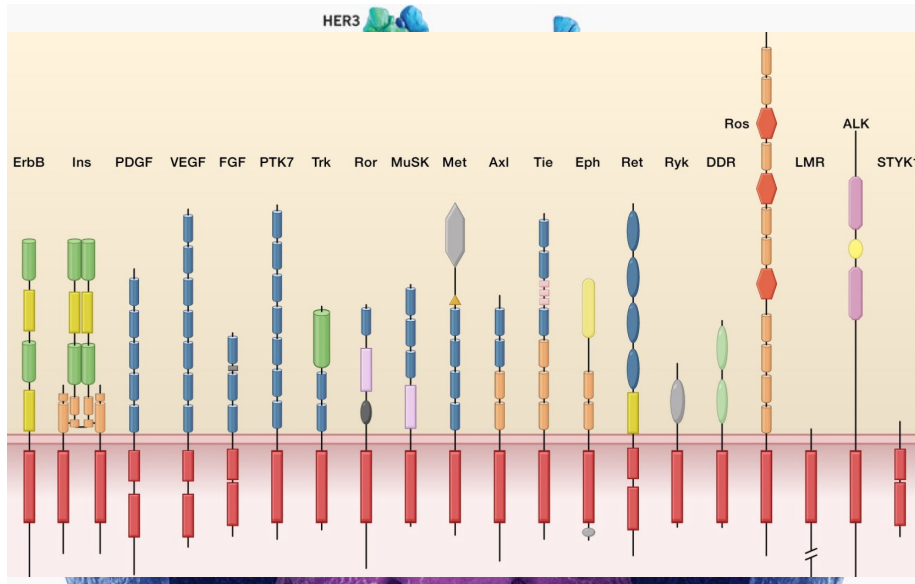
What could you do with us this summer?

ENPP7 – a lipid cleaving enzyme in your gut



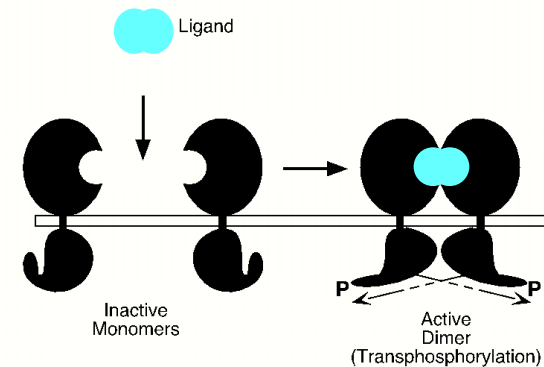
- Enzymology (Biochemistry!)
- Lab meetings
- Lunch discussions
- Coffee/Tea breaks
- Have fun and meet great people!

Regulation of Receptor Tyrosine Kinases (RTKs)



Regulate key cellular processes (proliferation, differentiation)
Disregulation can cause cancer and other diseases

“Textbook” view of receptor activation by
ligand induced receptor dimerization



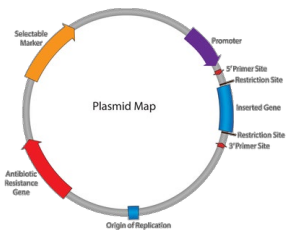
Elegant, but **it's more complex** for many of
the 20 families of RTKs –
larger oligomers, lifetimes, co-receptors

Kate Ferguson

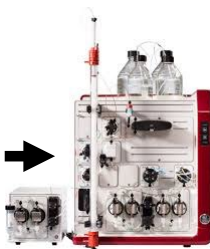
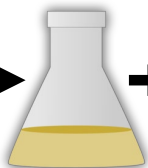
Yale Cancer Biology Institute & Department of Pharmacology,
Advanced Biosciences Building, Yale West Campus
kathryn.ferguson@yale.edu



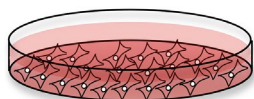
Yale University
School of Medicine



Generate/alter plasmids for protein expression



Recombinant protein purification



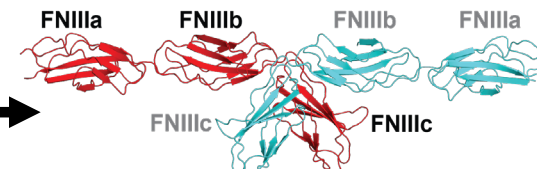
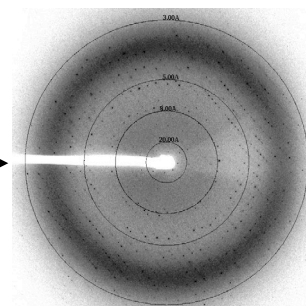
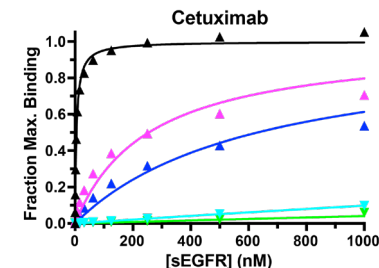
Expression in mammalian cell

Research Projects

Quantitative Biophysical Experiments – oligomerization, ligand binding analysis

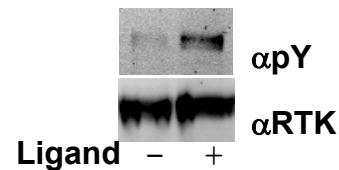
Low resolution shape information – SAXS, EM

Atomic resolution structure determination: X-ray crystallography, EM



Predict mechanism and test hypothesis in cellular assays

Functional assay



What to expect in the Ferguson Lab



- **Small lab - two postdocs, and a rotation student**
- **Share space and collaborate with Lemmon Lab**
- **You would**
 - **work directly with one of my postdocs on daily basis**
 - **meet with me at least weekly to discuss progress and plans**
 - **Participate in our weekly group meetings (with Lemmon lab)**
- **Expect to start with simpler techniques and shadow to learn more advanced procedures**
- **Please email if you are interested!**

THANKS!

Kate Ferguson

**Yale Cancer Biology Institute & Department of Pharmacology,
Advanced Biosciences Building, Yale West Campus
kathryn.ferguson@yale.edu**



**Yale University
School of Medicine**

Novel Approaches to evaluate and treat Gastrointestinal Diseases

John Geibel, MD, DSc, MSc, AGAF, FRSM

Vice Chairman Department of Surgery

Director of Surgical Research

Professor of Surgery and Cellular and Molecular Physiology

American Gastroenterological Association Fellow

Fellow of the Royal Society of Medicine



Projects Available for Summer 2019

- **Intestinal Repair**
 - Application of a nanoparticle based biological adhesive
 - Evaluate the tensile strength of the adhesive in *in vitro* and *in vivo* settings
 - Use of biological adhesive in 3D bioprinted intestine
 - Evaluate the use of “biopatches” for intestinal repair *in vivo*
- **Application of Novel Bioadhesives for Vascular Repair**
 - Use of unique bioadhesives for incorporation in vascular repair in animal models
- **Nanoparticle Therapy to treat Intestinal Diseases**
 - Using custom nanoparticles as a rapid therapeutic for intestinal diseases
 - IBD, Diarrhea, Crohns Disease



Mentoring Plan for Summer Students

- Dr. Geibel will conduct weekly laboratory meetings to review goals for coming week
 - Lab Meeting will discuss all research projects in the lab and allow for discussions with all lab members
- Student will be Assigned to Senior lab personal who will monitor and help student with project
 - Student will both assist and conduct studies in the designated project
- Dr. Geibel will be available all summer to provide additional insight as needed in the lab.



Plants have circadian rhythms

Joshua Gendron

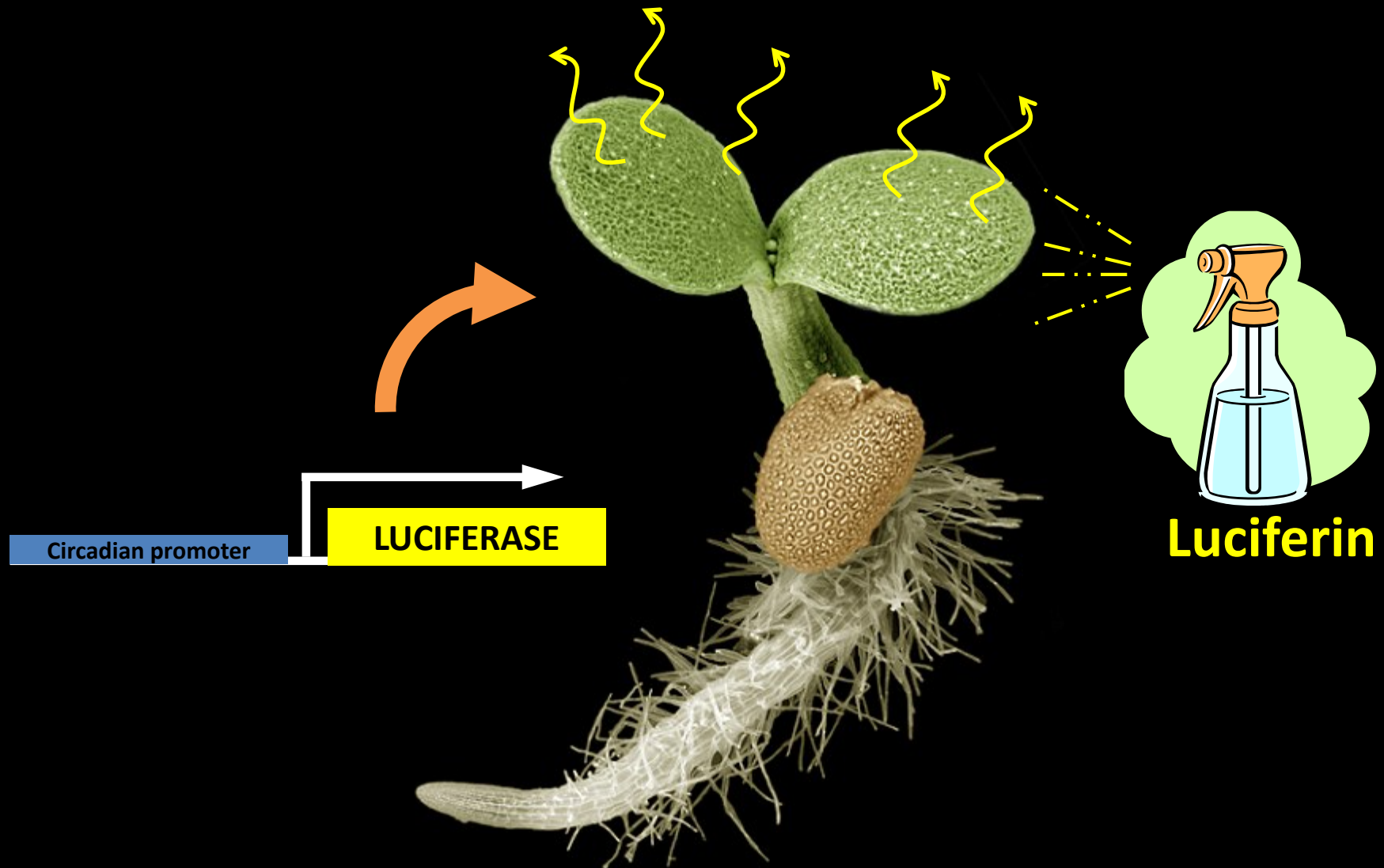
joshua.Gendron@yale.edu

Lab focus:

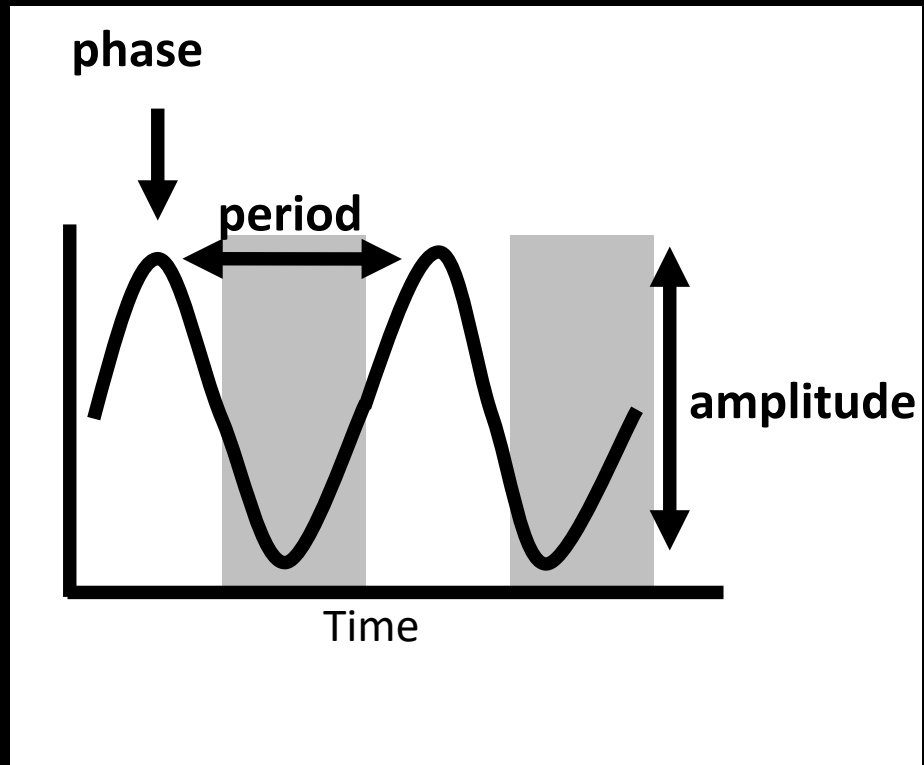
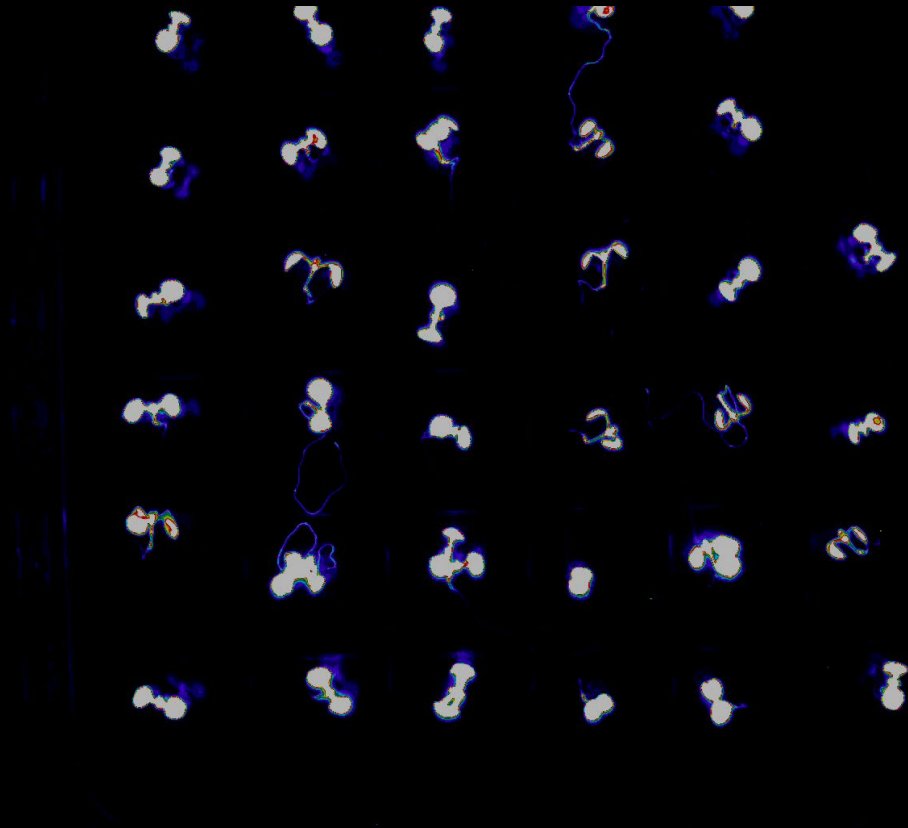
Daily and seasonal rhythms in plants



We can track circadian rhythms in real time



Students perform genetics, biochemistry, and molecular biology to understand daily or seasonal rhythms in plants



Students are actively performing experiments, mentored by me and a grad student or postdoc. Students are expected to participate similar to any lab member.

Gerstein Lab @ Yale (gersteinlab.org)

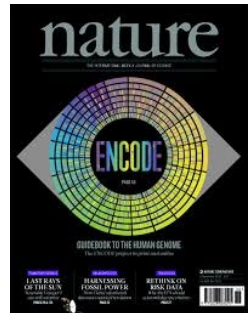
Program in Computational Biology & Bioinformatics (CBB)
Department of Biophysics & Biochemistry (MBB)

Lab focuses on biological data sciences

- Human Genomics
- Human Variation
- Functional Genomics

Participate in many big genomics consortia: **ENCODE**,
PsychENCODE, **TCGA**, **exRNA** ...

Contact: joel.rozowsky@yale.edu or mark@gersteinlab.org



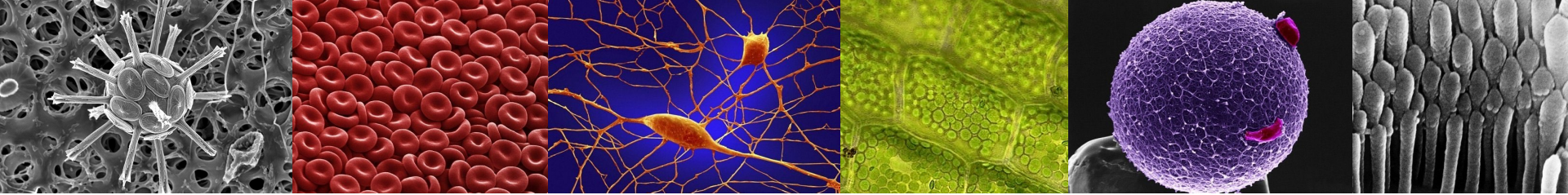
Projects in Gerstein Lab

- Human Genomics ([ENCODE](#) + [exRNA](#))
- Cancer Genomics & Human Variation ([TCGA](#) + [ENCODE](#))
- Brain Genomics ([PsychENCODE](#))
- Specific Topics: [Biological Networks](#), [Machine Learning Approaches](#), [Genome Annotation](#), [Pseudogenes](#), [Protein Structure](#), [Structural Variation](#), [Functional Genomics](#), [Tool Development](#)

Contact: joel.rozowsky@yale.edu or mark@gersteinlab.org

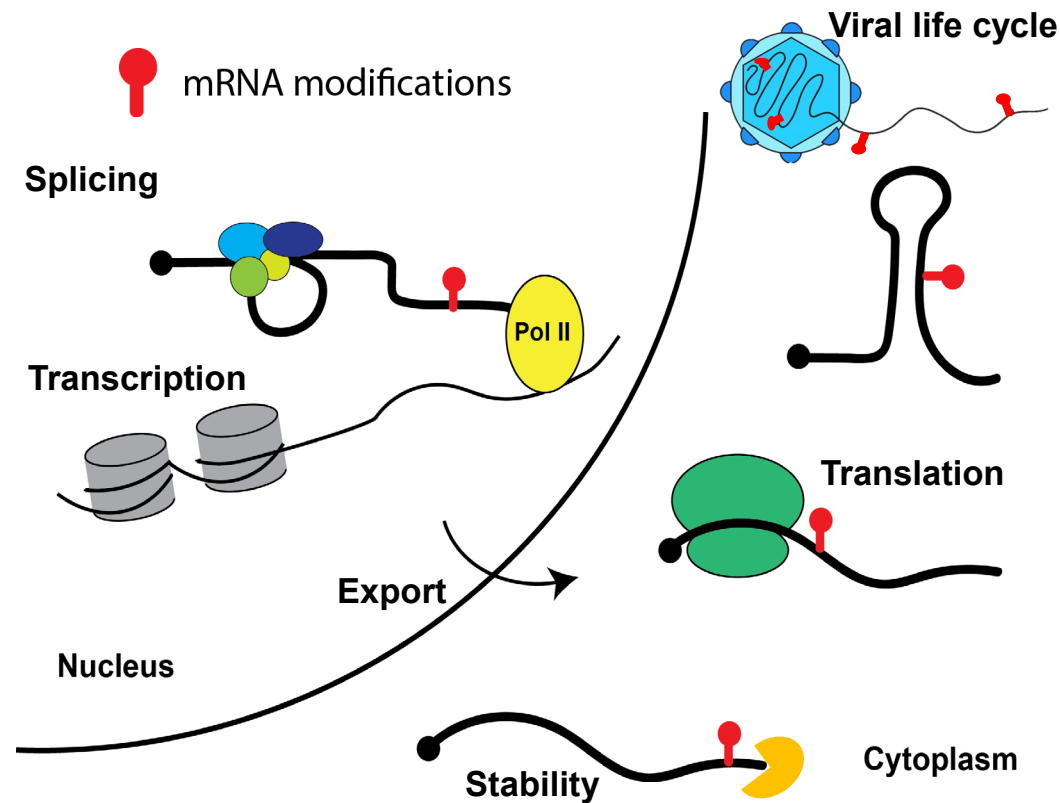
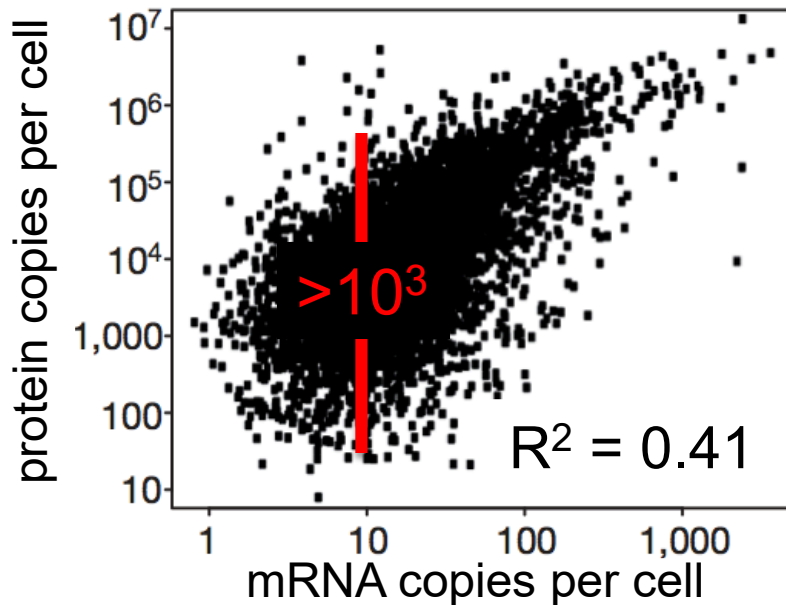
Gerstein Lab undergrad research experience

- Initially meet with MBG (will direct to potential mentors)
- Mentored by graduate students + postdocs + assoc. res. Scientists
 - big lab ~25 people + undergrads (5+ for summer)
- Weekly group meetings, journal clubs + subgroup meetings
- Lab Presentation (JC or GM) sometime during the summer presenting either a paper or summer work
- Many summer undergrad continue to work in the lab
- Many undergrads are included as authors on lab publications



DNA → mRNA → Protein

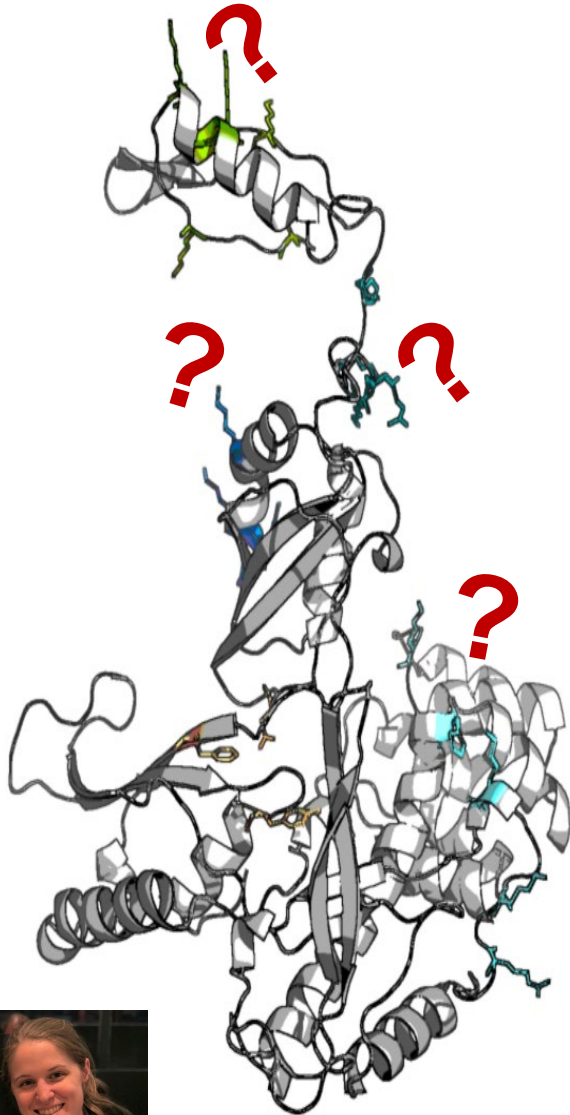
But did you know?



wendy.gilbert@yale.edu

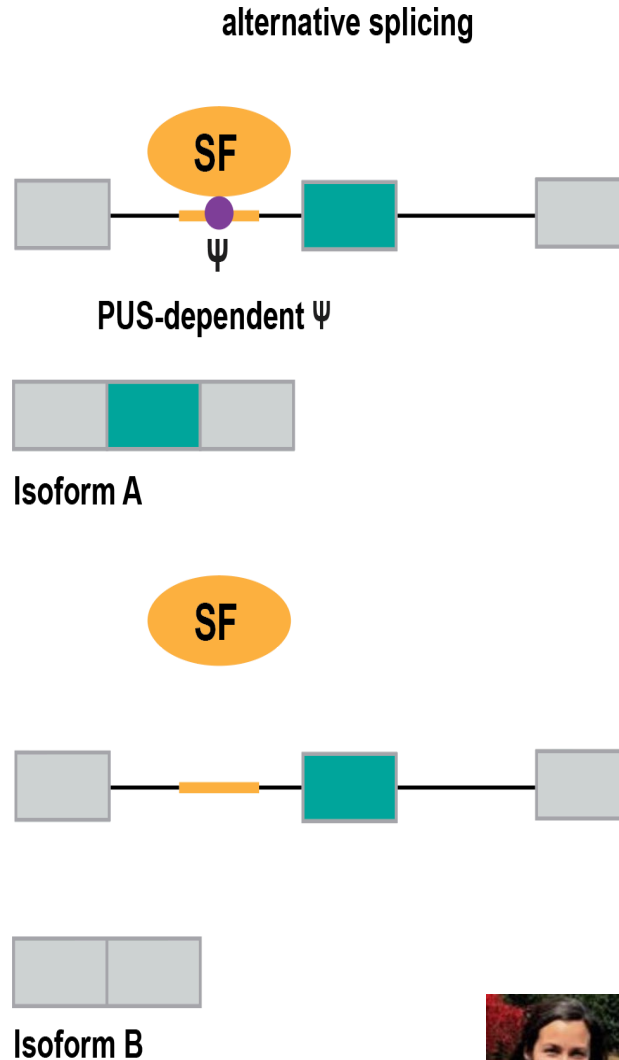
come visit us in SHM C129!

How does a PUS protein recognize it's RNA substrates?



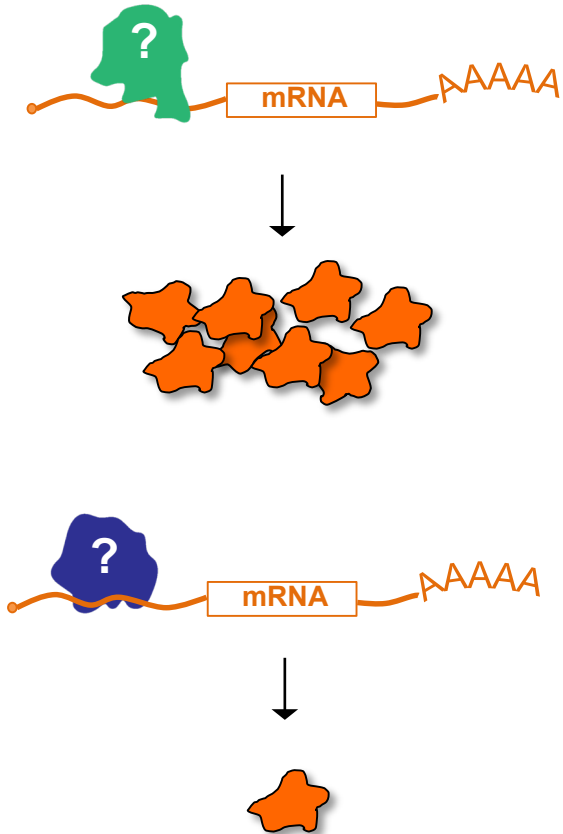
Erin Borchardt

How ψ does regulate alternative splicing?



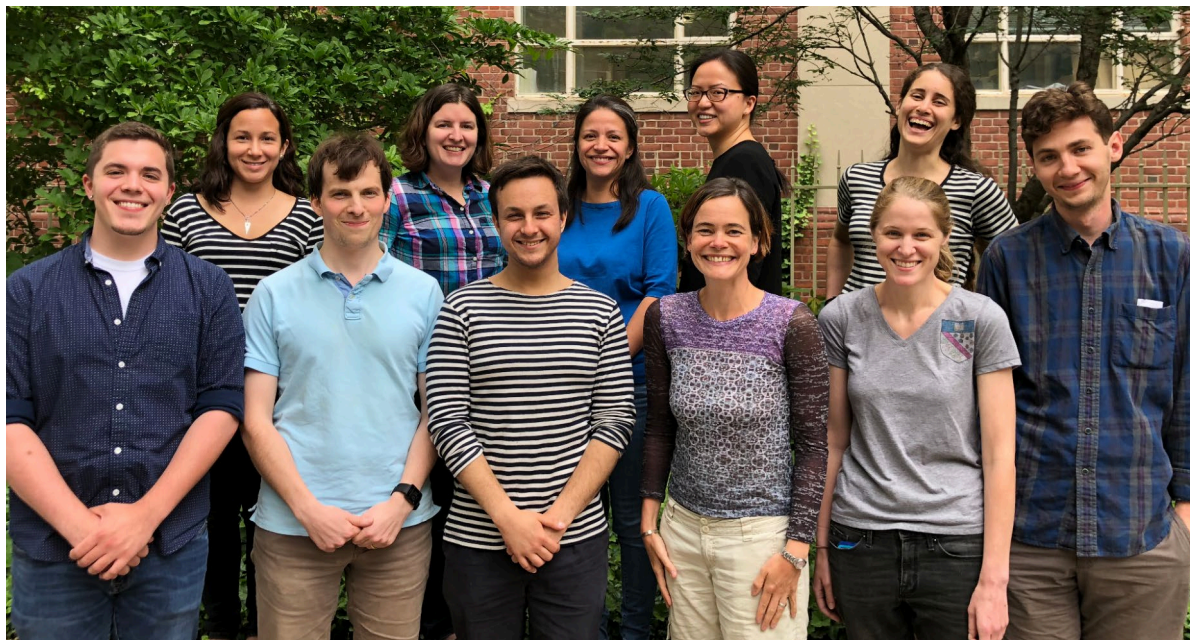
Nicole Martinez

Which proteins regulate mRNA translation?



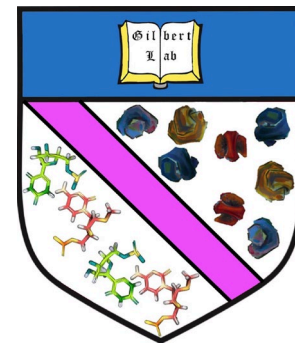
Rachel Niederer

Come work with us!



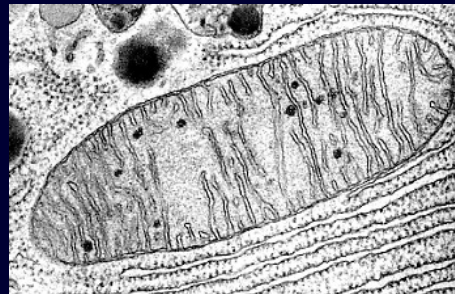
- we're looking for up to 3 students
- weekly lab meetings
- day to day mentoring by postdoc
- contact Dr. Gilbert if you are interested!

wendy.gilbert@yale.edu

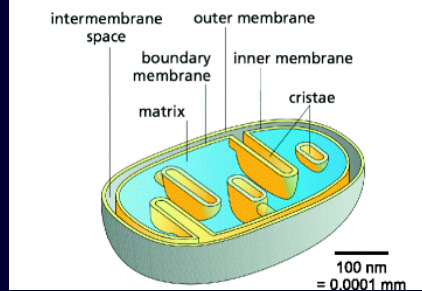


Elizabeth Jonas, elizabeth.jonas@yale.edu

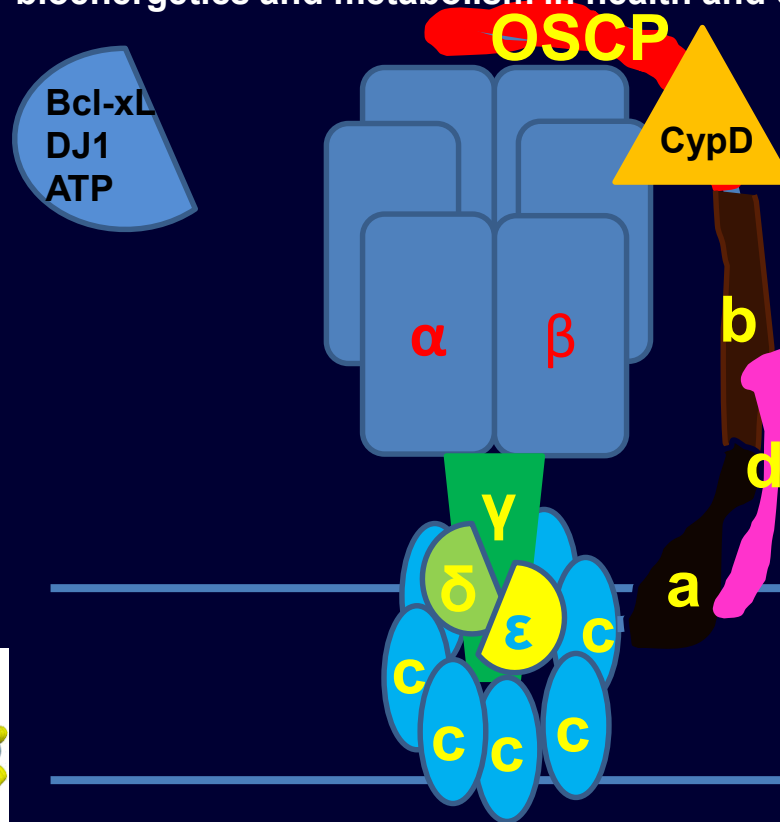
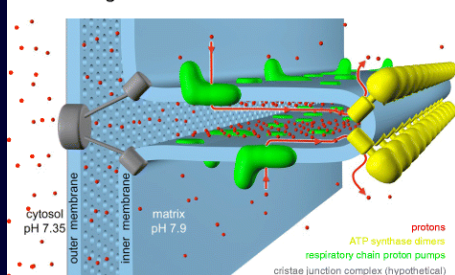
ATP synthase contains a non-selective leak that regulates cellular bioenergetics and metabolism in health and disease



Membrane compartments in the mitochondrion



Organisation of mitochondrial cristae

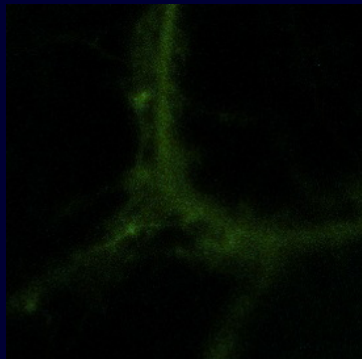


Projects for the summer:

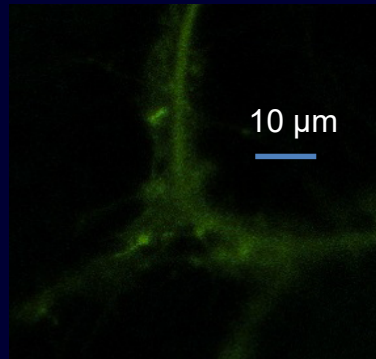
1. Mitochondria in LTP, the cellular basis of memory formation
2. Mitochondrial regulation of mRNA translation in the synapse and in translation of cell cycle proteins in Parkinson's disease.
3. Mitochondrial regulation of neuronal development: Changes in metabolism during normal development and in Fragile X Syndrome.
4. Regulation of synaptic vesicle positioning and acute changes in cytoskeleton during presynaptic plasticity.

Long term increases in ATP levels occur in synaptic spines after chemical NMDAr stimulation

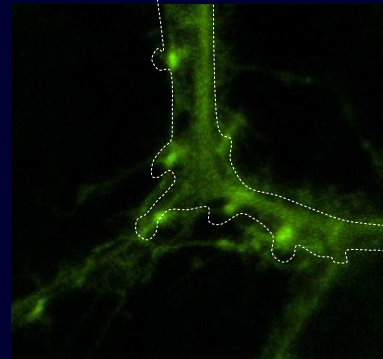
Dendritic spines



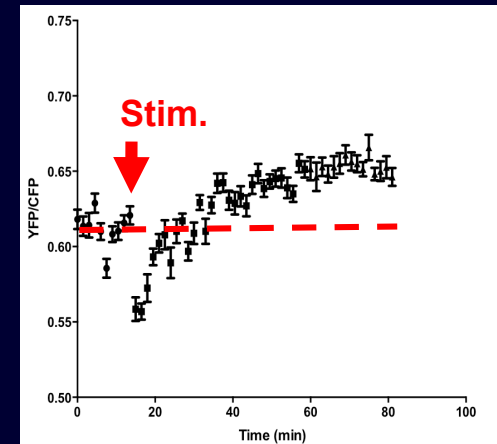
Before treatment



15 minutes after
Stim.



27 minutes after
Stim.



Details

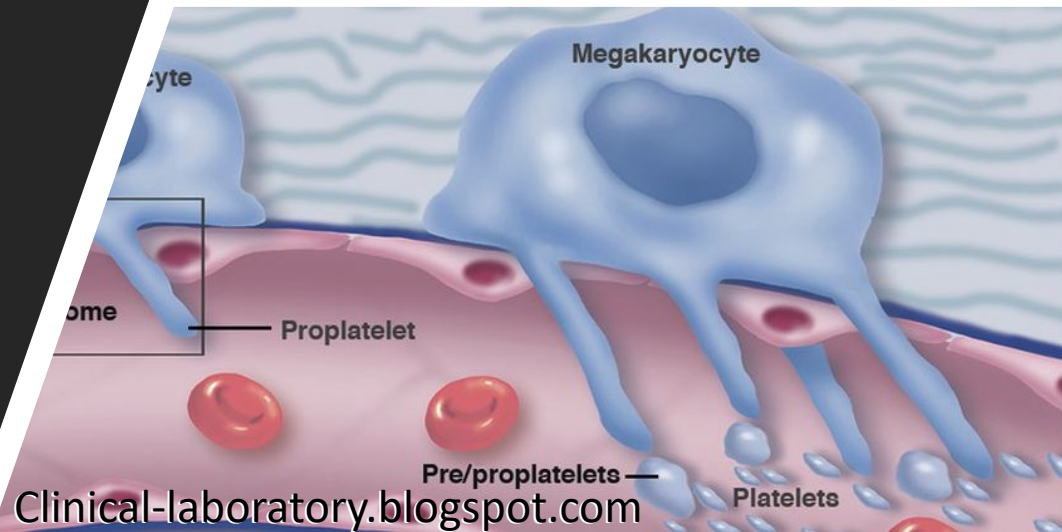
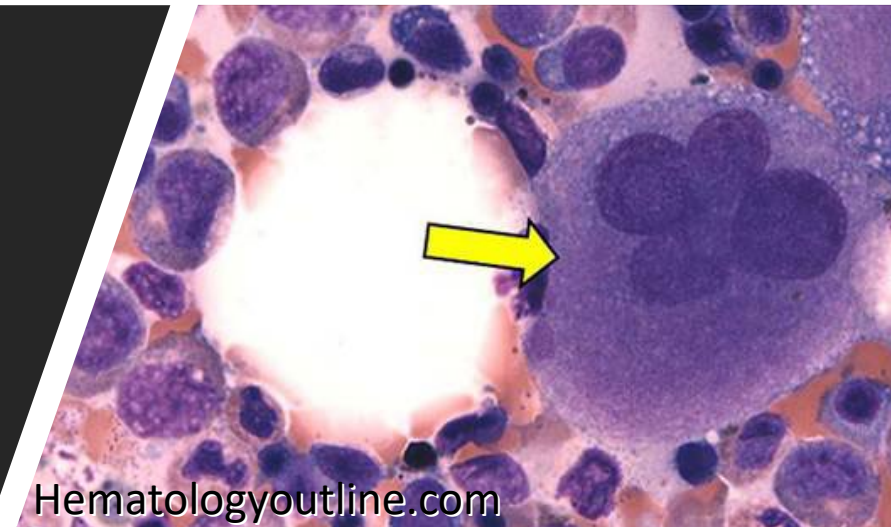
- Will meet with student weekly at least.
- Student will work closely either with me or with a post-doctoral fellow, depending on the project.
- Ideally, the student will participate in lab meetings and design a power point presentation on his/her work at the end of the summer.
- We would like to host 1-2 students.
- For more information, please email and meet with me to discuss. **elizabeth.jonas@yale.edu**.

Gene Regulation in Megakaryopoiesis

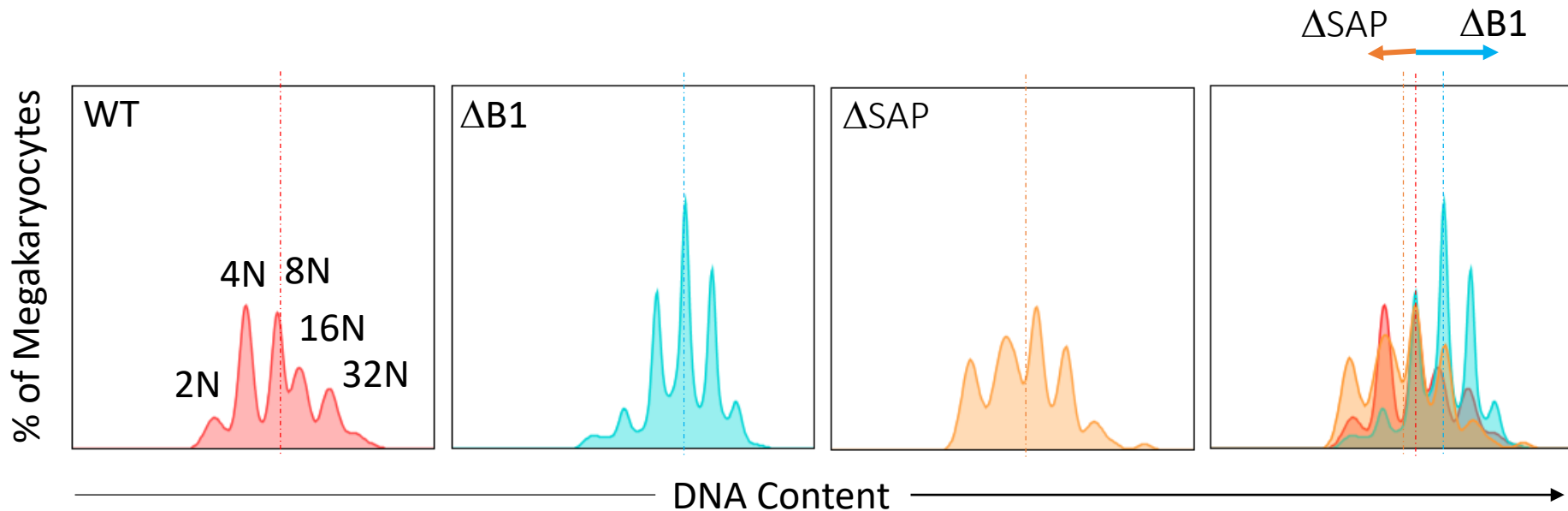
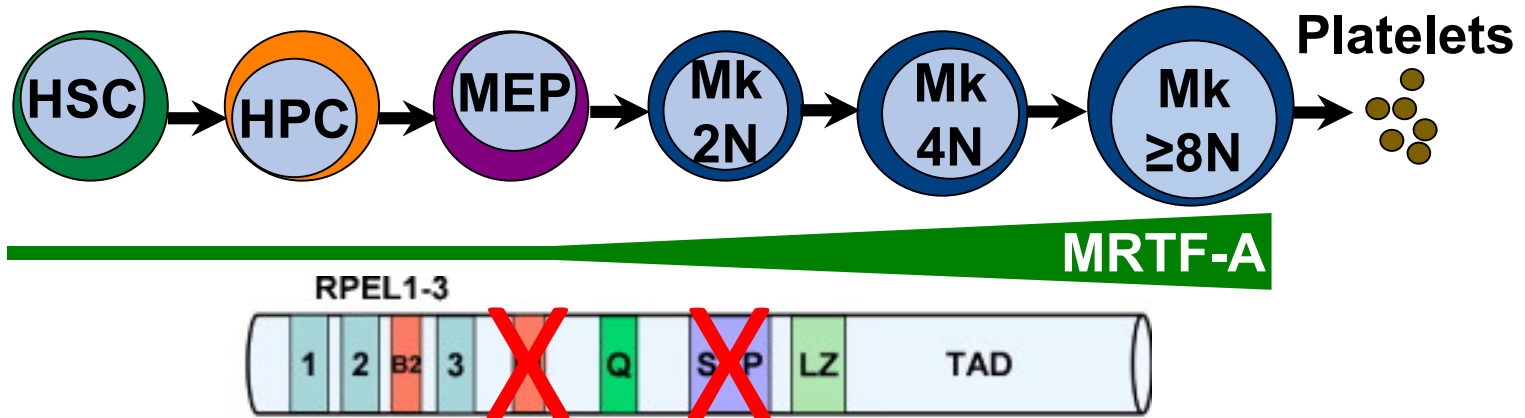
Vanessa Scanlon, PhD

Vanessa.Scanlon@yale.edu

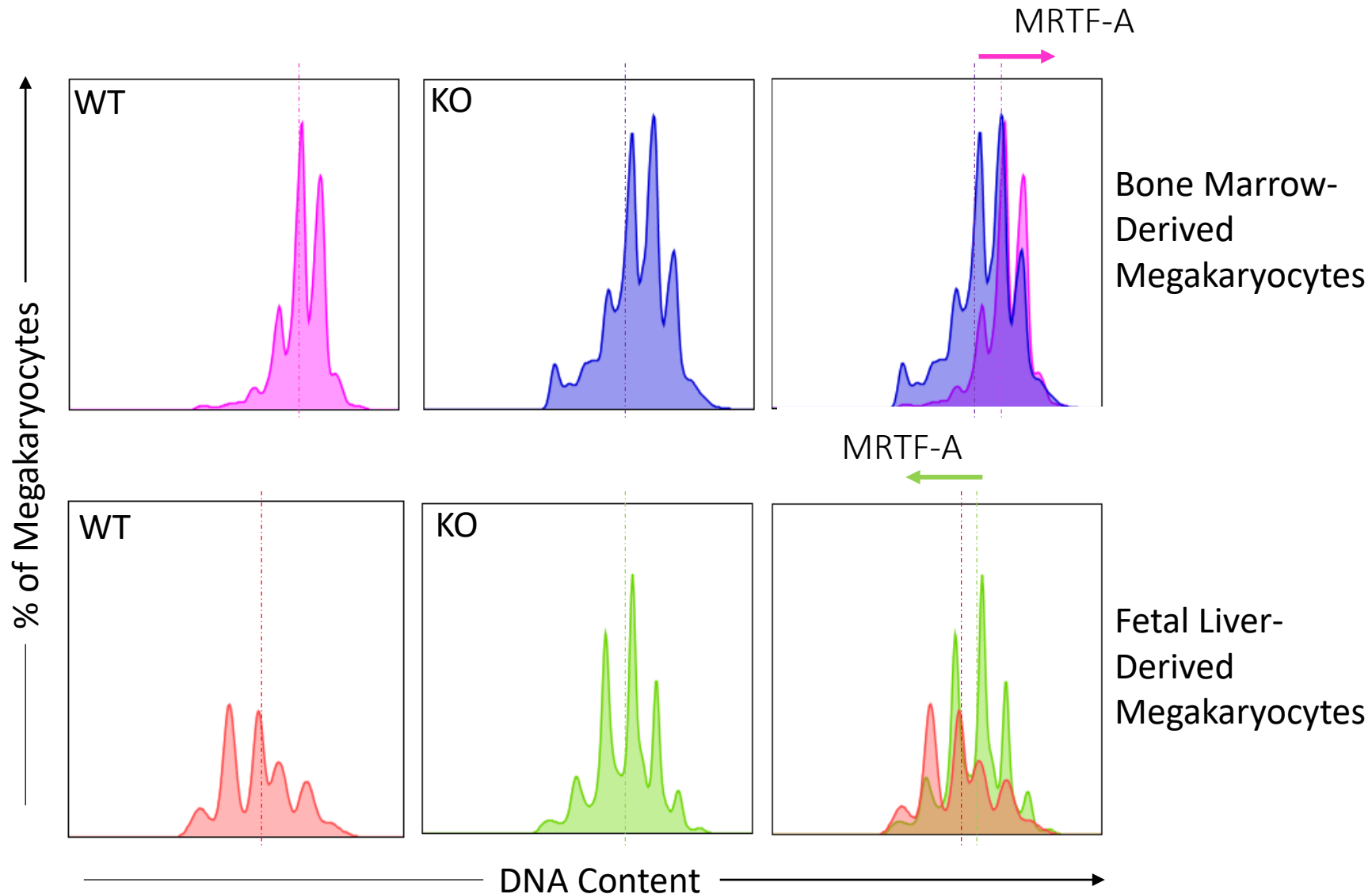
- Megakaryocytes (Mk)= Large polyploid bone marrow cells - generate platelets
- Critical for blood clotting
- Acute Megakaryoblastic Leukemia - increased megakaryoblasts (immature Mk cells) in the bone marrow



Which domains of MRTF-A are responsible for promoting megakaryopoiesis?

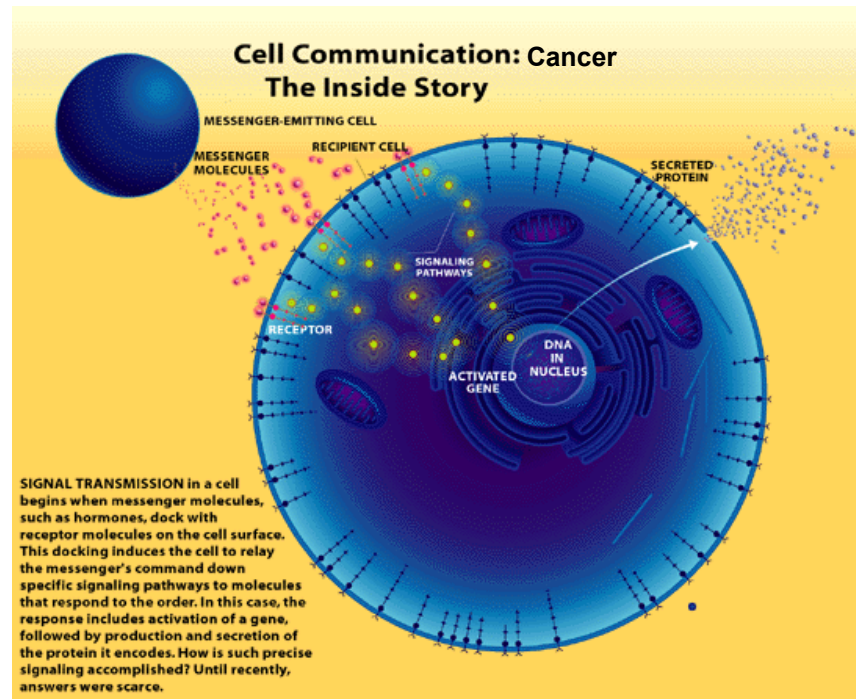


Does MRTF-A have opposite effects on megakaryopoiesis during development?





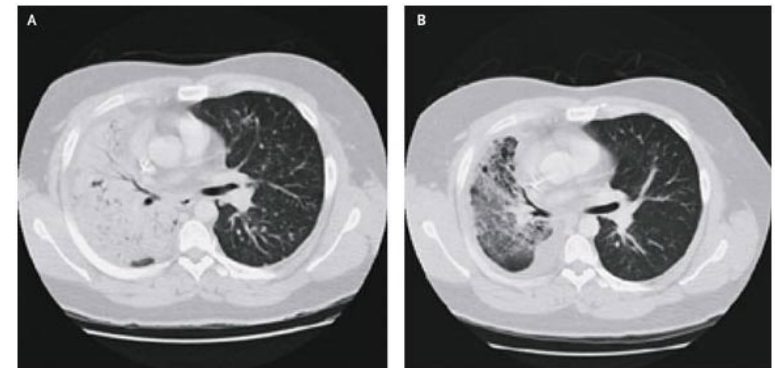
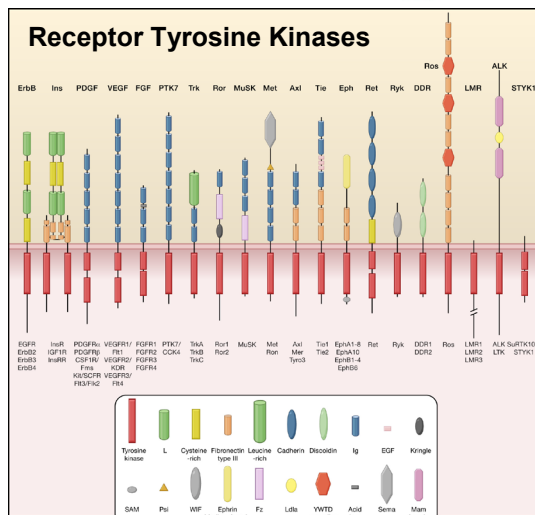
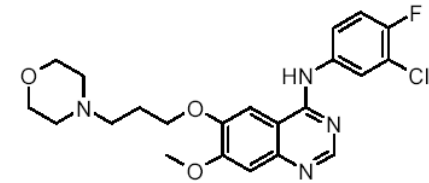
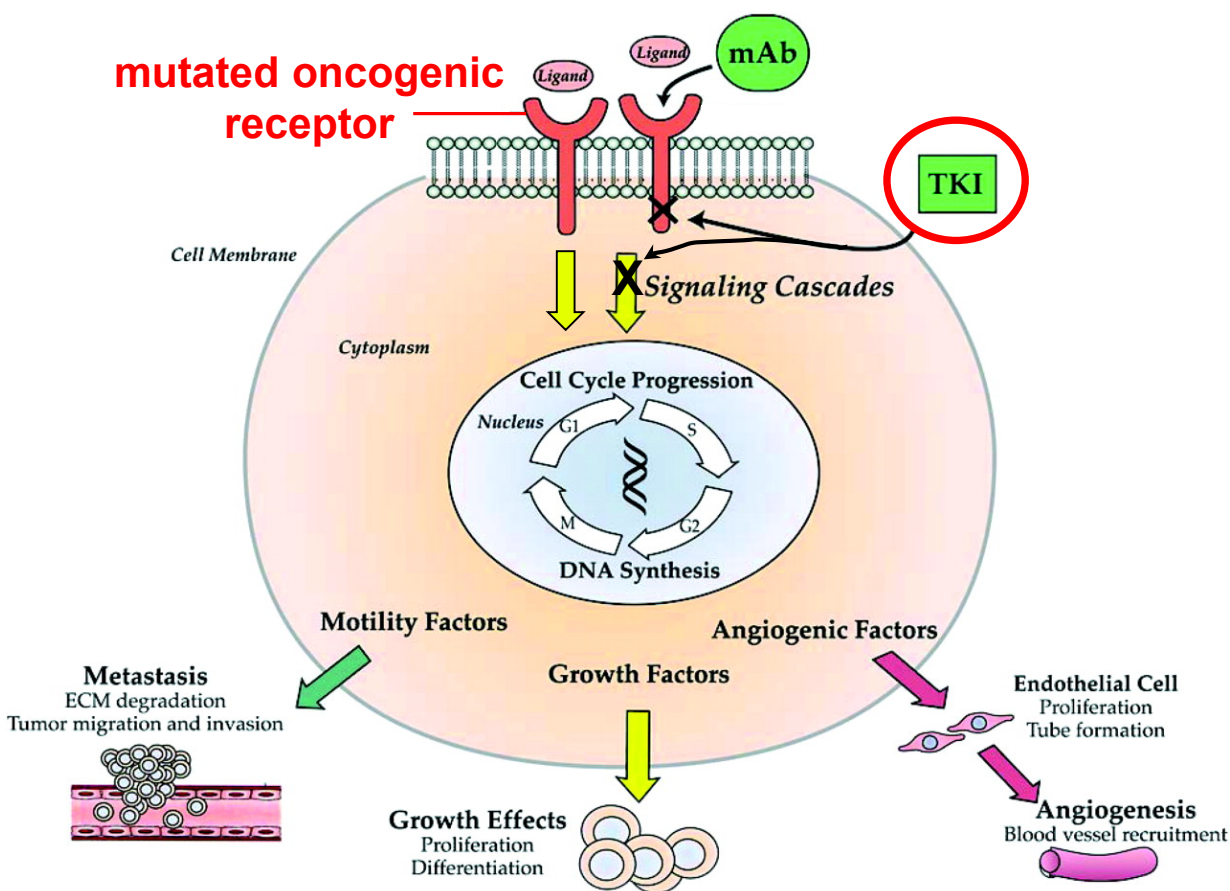
Signaling by cell-surface growth factor receptors: in biology and disease (cancer)



Mark A. Lemmon:

**Yale Cancer Biology Institute
Department of Pharmacology, YSM
mark.lemmon@yale.edu**

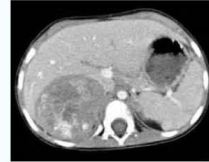
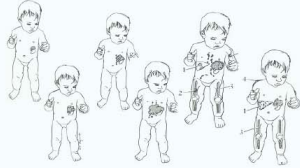
mutated oncogenic
receptor



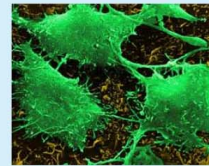
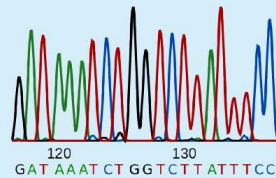
Lynch *et al.*, *N.E.J.M.* **350**, 2129 (2004)

**Diagnosis
and staging**

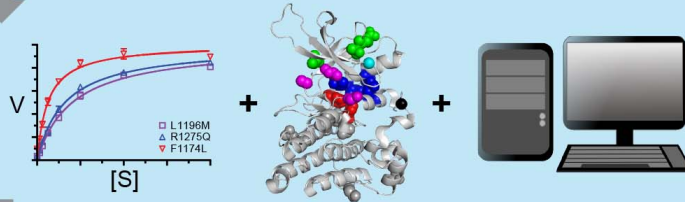
Neuroblastoma



ALK sequencing



**Biochemical,
structural, and
computational
analyses**



Identification of ALK-dependent tumors

Therapy

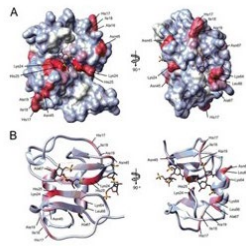
ALK kinase inhibitors

Chemo-therapy → Surgery → Stem cell transplant → Cis-Retinoic acid + immunotherapy



- Lab of 5 grad students, 5 post-docs, 3-4 undergrads, plus lab manager
- Direct Supervision by 1-2 grad students or postdoc
- Lab meetings Thurs afternoons
- Learn to make mutated recombinant protein and assess activity – for mutations found in lung cancer rebiopsy program at YNHH
- Help identify ‘rules’ for inhibitor application and choice

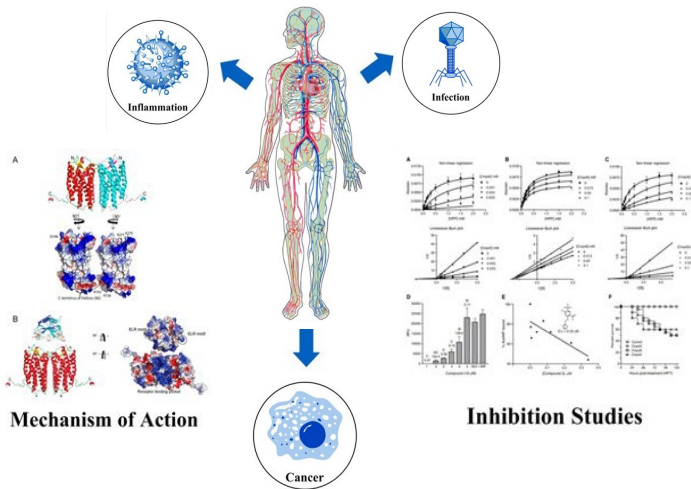
Elias Lolis (elias.lolis@yale.edu)



Structural Biology

Lab background: Understand mechanism of macromolecules and small molecules in inflammation, cancer, and microbial infections.

- Techniques:
 - Protein purification
 - Enzyme kinetics
 - X-ray crystallography
 - Yeast and mouse models



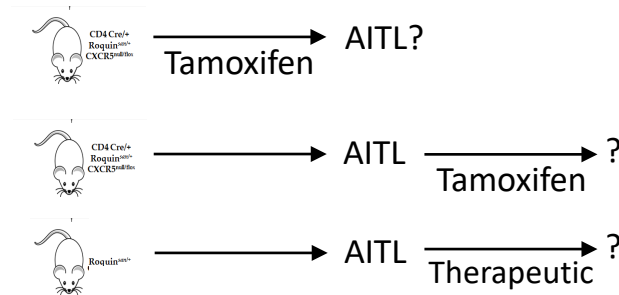
Elias Lolis (elias.lolis@yale.edu)

Macrophage migration inflammatory factor (MIF)



1. Molecular Dynamics defined an allosteric site at Y99
2. NIH researcher called about Y99C mutant in JSIA
3. **Characterize Y99A: enzyme kinetics, receptor binding, inhibitor binding.**

CXCR5 in angioimmunoblastic T-cell lymphoma (AITL)



1. AITL mouse model found in Australia
2. Inducible Cre/lox CXCR5 KO to determine if
(a) **CXCR5 is involved in lymphoma development** and (b) **is a therapeutic target.**
3. **Develop therapeutics**

Mentoring

- My office is at the very end of the lab with an open door policy. I also have *ad hoc* meetings with individuals
- A senior member of the lab (two graduate students and two postdoctoral associates) will mentor the undergraduate
- Students present their work at lab meeting every other week
- Students present journal manuscript
- One undergraduate this summer
- Seven previous Yale undergrads: 2 of 3 MDs went to Yale Medical School, 2 worked at Science/Health companies, 1 Stanford Law, 1 Deceased

FIXING THE NUCLEUS

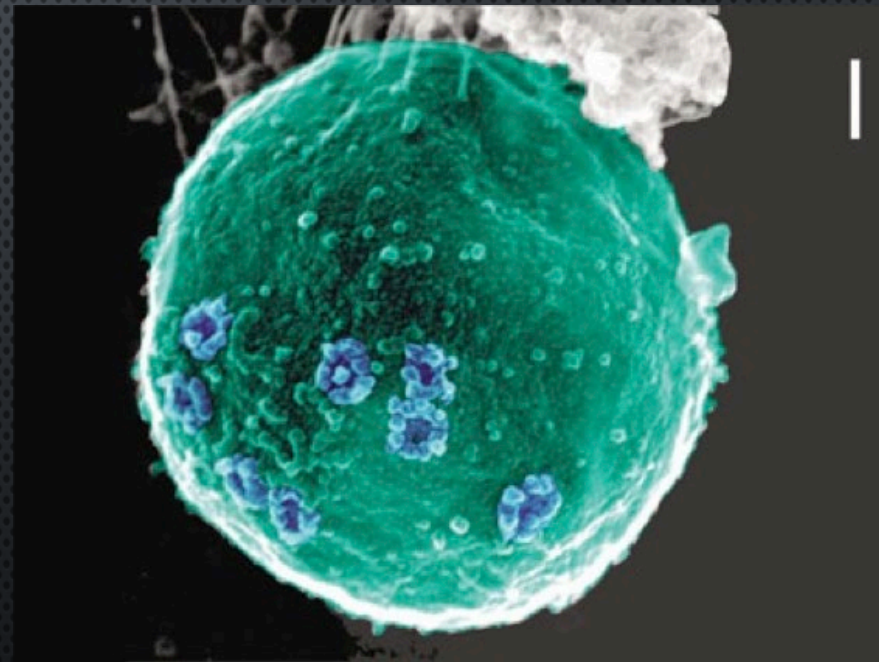
PATRICK LUSK

DEPARTMENT OF CELL BIOLOGY

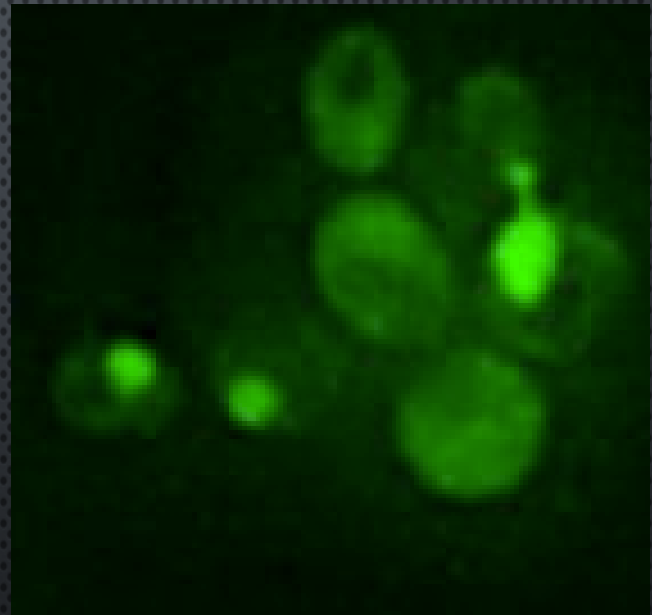
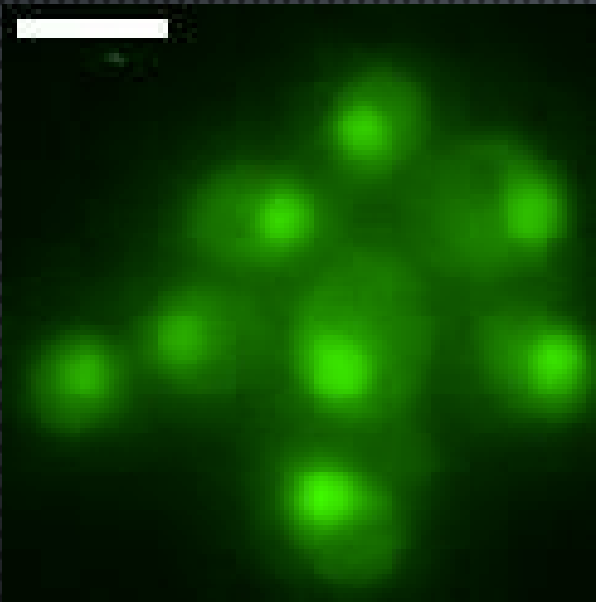
PATRICK.LUSK@YALE.EDU

BCMM 254B

On twitter: @Plusk4u



WHAT ARE THE MECHANISMS OF NUCLEAR MEMBRANE REPAIR?



WHAT PEOPLE ARE SAYING ABOUT THE LUSK LAB

"5 stars...working with
Dr. Lusk's group was nothing short of a revelation!"
-Bob Fluorophore, class of '19

"Two Thumbs Up! It was like living a dream, how
can I work anywhere else?"
- Andrew Petri, class of '19

"Who knew yeast could be so beautiful?"
-Emma Pipette, class of '21

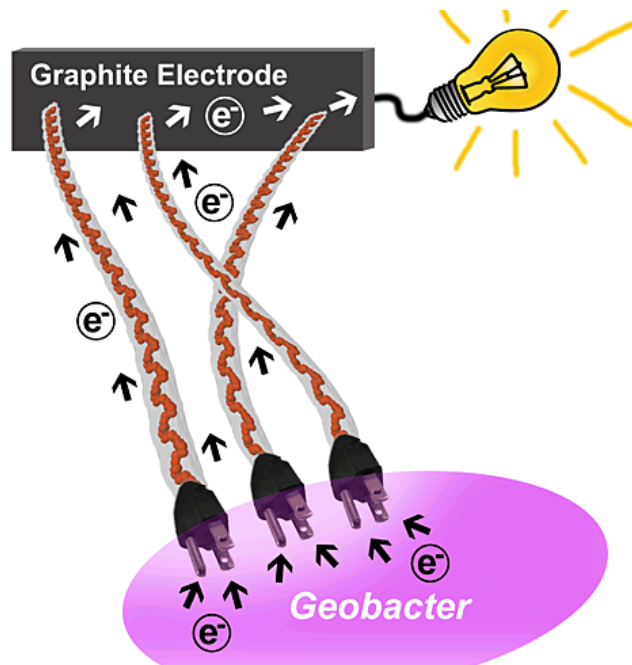
"Biology, mechanism, evolution....SCIENCE, so cool."
-Doug Yestbudder



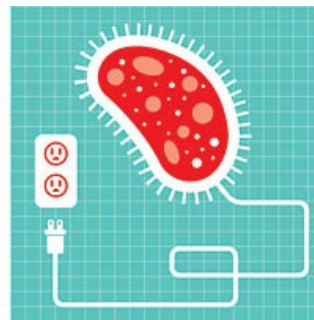
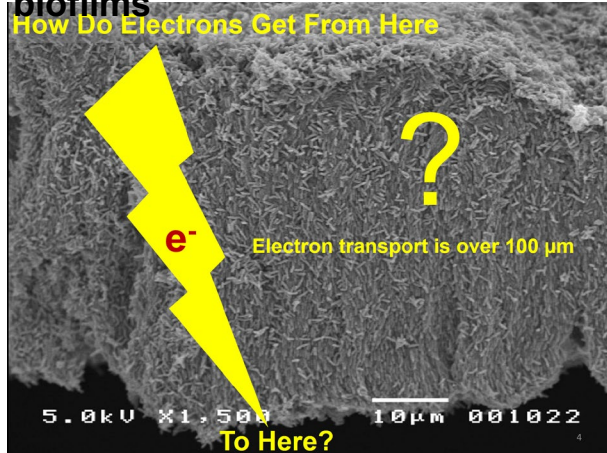
Nikhil Malvankar Lab

New imaging and measurement technologies to define the mechanisms by which microbes interact with and manipulate their environment. The ultimate goal is to engineer these interactions to control microbial pathophysiology and ecology.

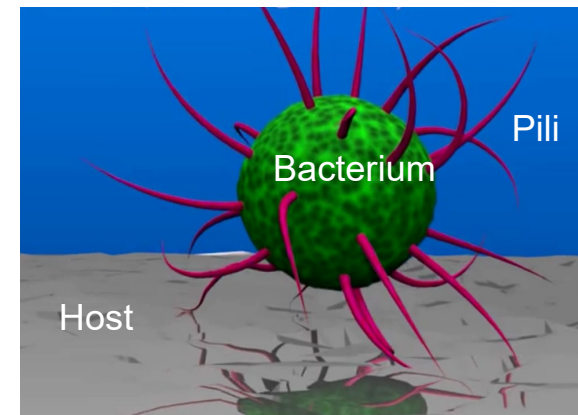
Electric Bacterial Nanowires



Bacterial communities in biofilms



Host-pathogen Interactions in single cells and biofilms

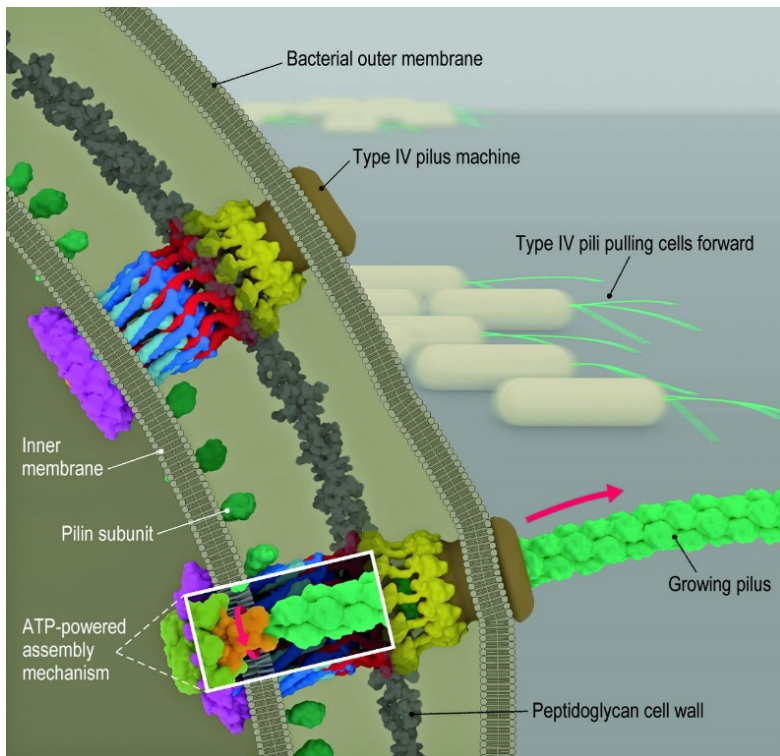


Nikhil Malvankar

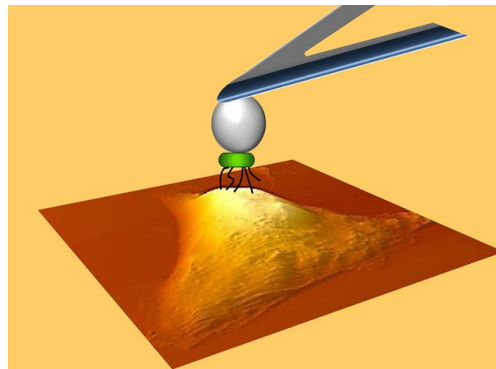
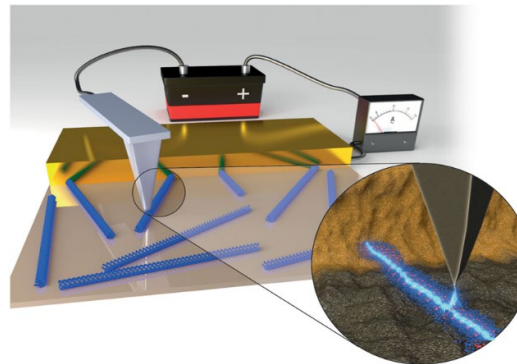
Mol. Biophysics & Biochem.
Microbial Sciences Institute
nikhil.malvankar@yale.edu

Rotation Projects – Physics, Chemistry, Biology & Engineering

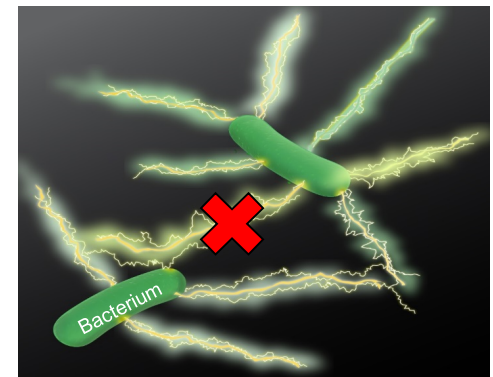
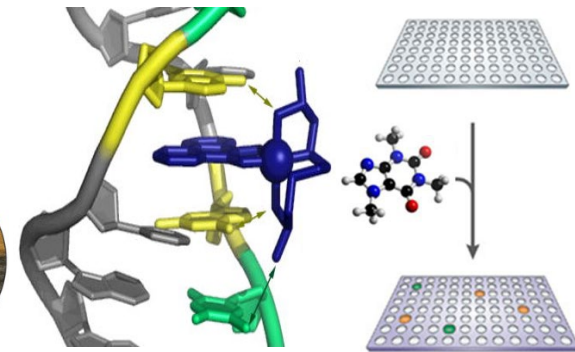
Cryo Electron Microscopy & Tomography
of bacterial nanowires and assembly



Targeting bacterial
survival mechanisms

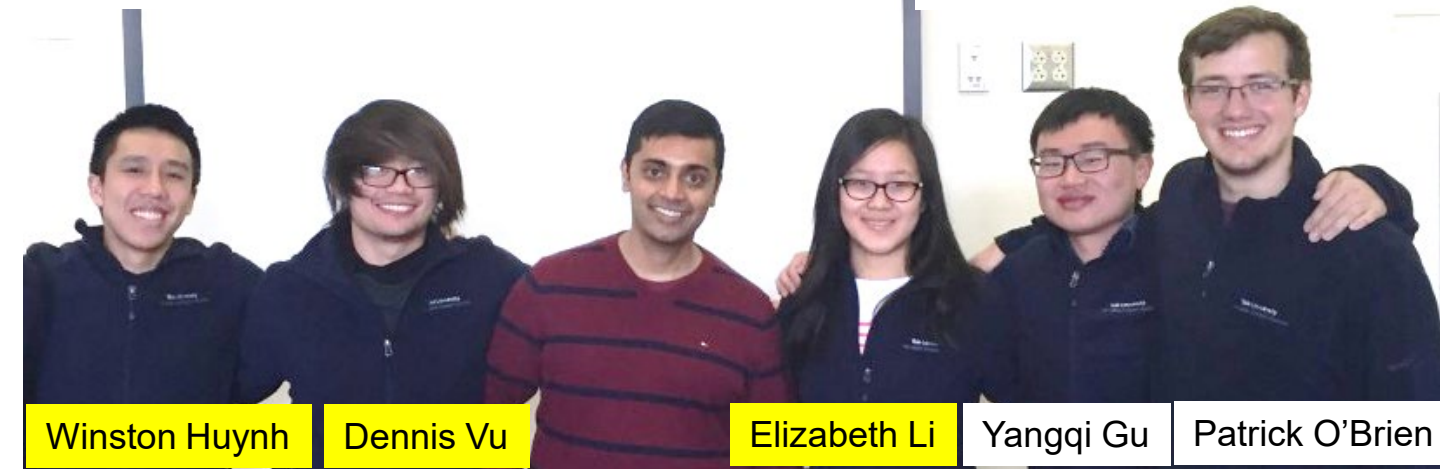


Novel drugs to neutralize
host-pathogen interactions

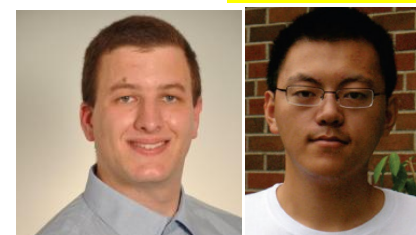




Sibel Yalcin Catharine Shipps Sophia Yi Peter Dahl Vishok Srikanth Cong Shen Ruchi Jain Dan Shapiro



Winston Huynh Dennis Vu Elizabeth Li Yangqi Gu Patrick O'Brien



Jen s Xinglong Ren



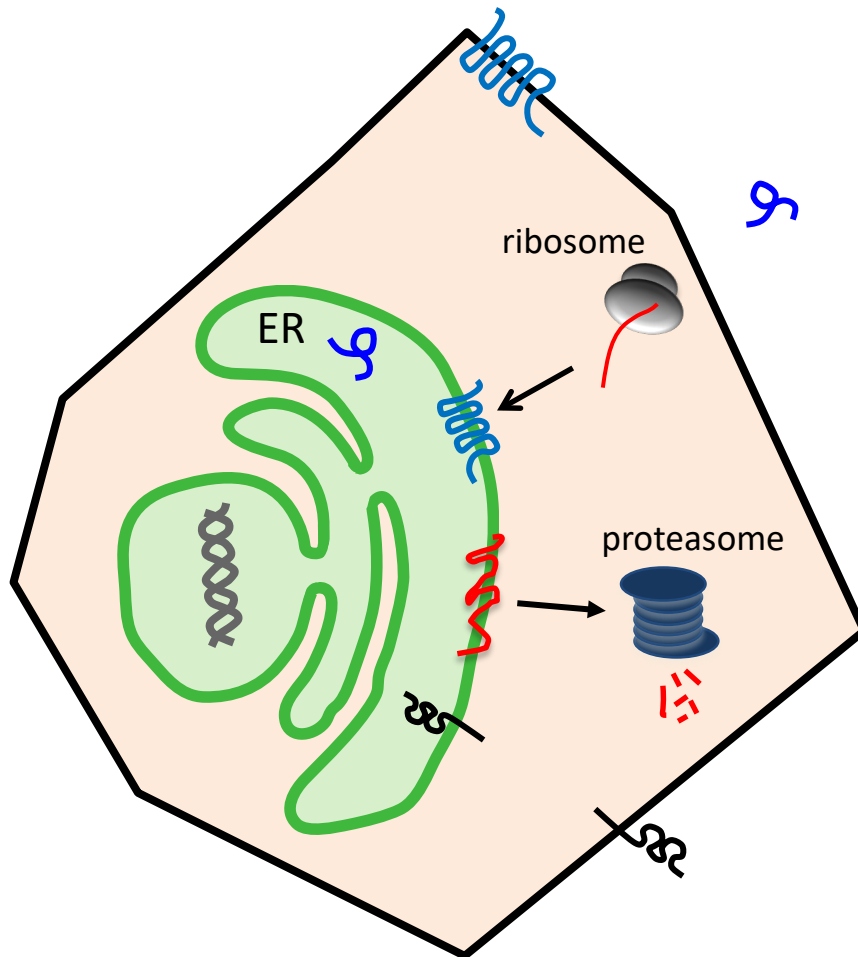
Claire Sheen

Undergraduate students



Protein biosynthesis and quality control at the ER

The Mariappan Lab (malaiyalam.mariappan@yale.edu)



The endoplasmic reticulum (ER)

➤ responsible for synthesizing ~ 30% of human proteome, which includes insulin, antibodies, channels and receptors.

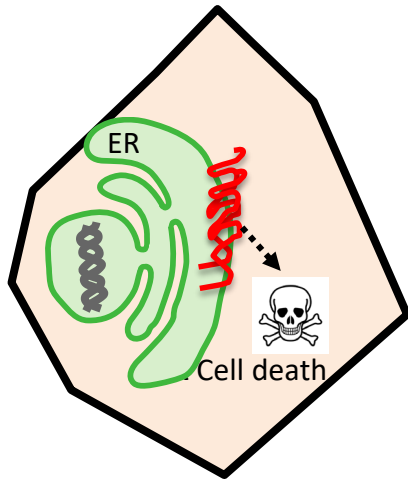
We are investigating:

- How are proteins targeted to the ER?
- How are misfolded proteins eliminated from the ER?
- How do the ER maintain its homeostasis ?

Why are we interested?

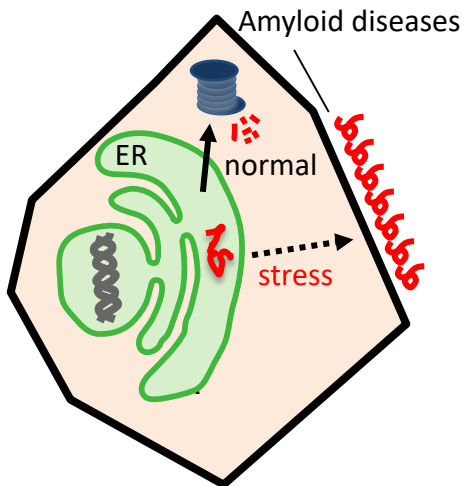
➤ Defects in protein biogenesis are associated numerous human diseases: Cystic Fibrosis, Diabetes, and Retinitis Pigmentosa.

Summer Projects for Undergraduate Students



1. How do **Rhodopsin mutants** cause blindness?

- Create mutations in the Rhodopsin gene by PCR and cloning
- Test the expression of mutants in HeLa cells by Western Blotting and Immunofluorescence
- Monitor the Unfolded Protein Response by luciferase assay
- Perform cell-based apoptosis assays



2. How are **misfolded proteins** secreted and cause amyloidosis?

- Test the secretion of misfolded proteins into the cell culture media by Western Blotting
- Test whether stress conditions stimulate the secretion of misfolded proteins
- Identify chaperones that retain misfolded proteins under normal conditions by mass spectrometry

The Marippan Lab (Two positions available)
malaiyalam.mariappan@yale.edu

Current Members

Jacob Culver (Graduate Student)
Sha Sun (Post-Doctoral Associate)
Xia Li (Post-Doctoral Associate)
Zhouping Hong (Rotation Student)

Previous undergraduate members

Sneha Mittal,
Keith Anderson
Nisha Rajamohan
Jessica Maddela

Previous high school student

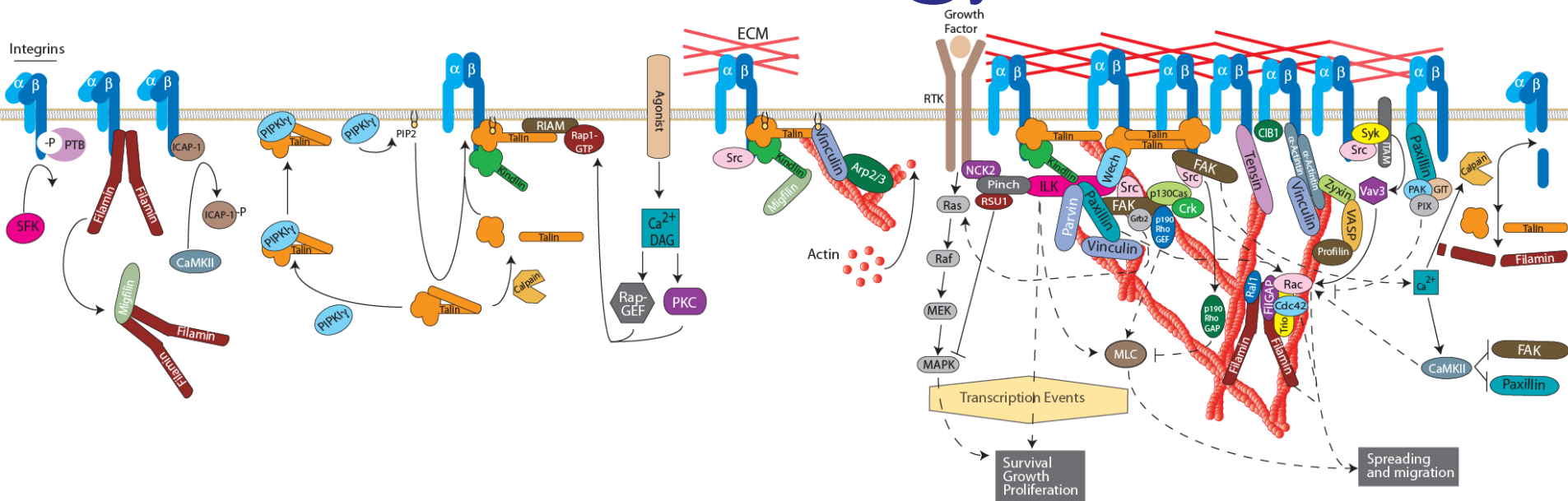
Nikhil Jaiswal
Poorna Balakumar

Mentoring of undergraduates

- Independent project
- Weekly one on one meeting with PI
- Working with post-docs or PI
- Presentation in lab meeting as well as in institute seminars

Adhesion Signaling and the Cytoskeleton

David A Calderwood
david.calderwood@yale.edu

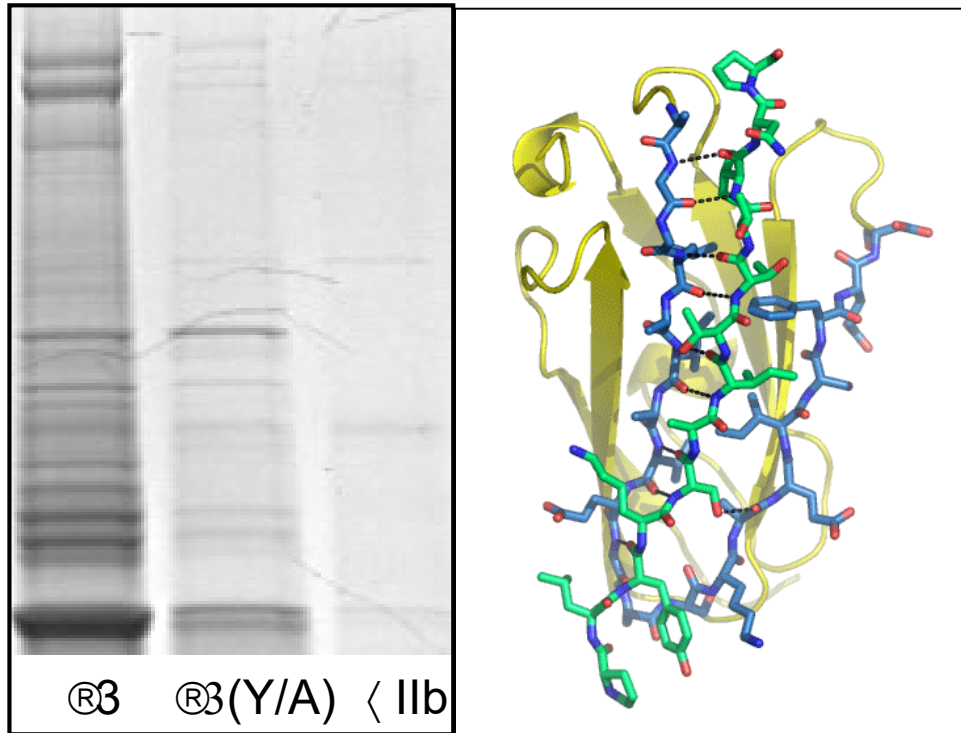


Functional significance of protein-protein interactions in controlling cell signaling, migration and morphogenesis

Structural Biology / Biochemistry / Cell biology

Adhesion Signaling and the Cytoskeleton

David A Calderwood



Structural Biology / Biochemistry / Cell biology

Adhesion Signaling and the Cytoskeleton

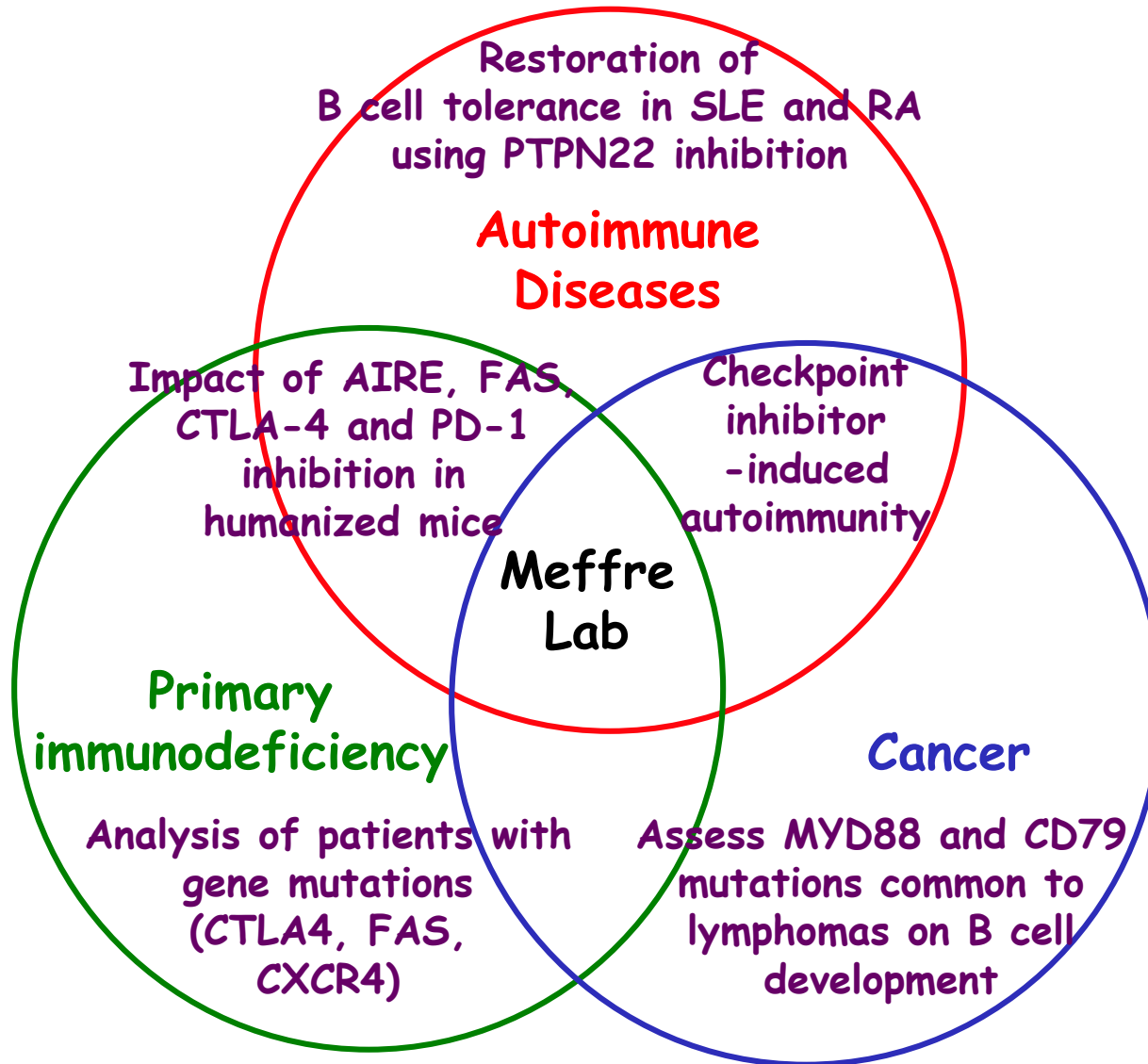
David A Calderwood
david.calderwood@yale.edu

- Dept. Pharmacology, Medical School
- <https://medicine.yale.edu/lab/calderwood/>
- Lab meetings 9AM Wednesday SHMB395D
- Mentoring by postdoc or senior grad-student
- 1 position open

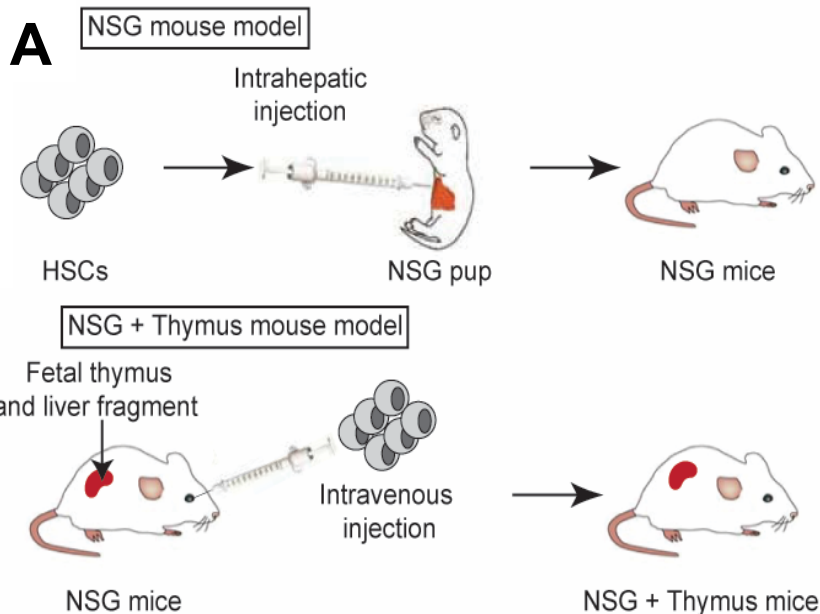
The regulation of human B cell tolerance

Eric Meffre, Department of Immunobiology

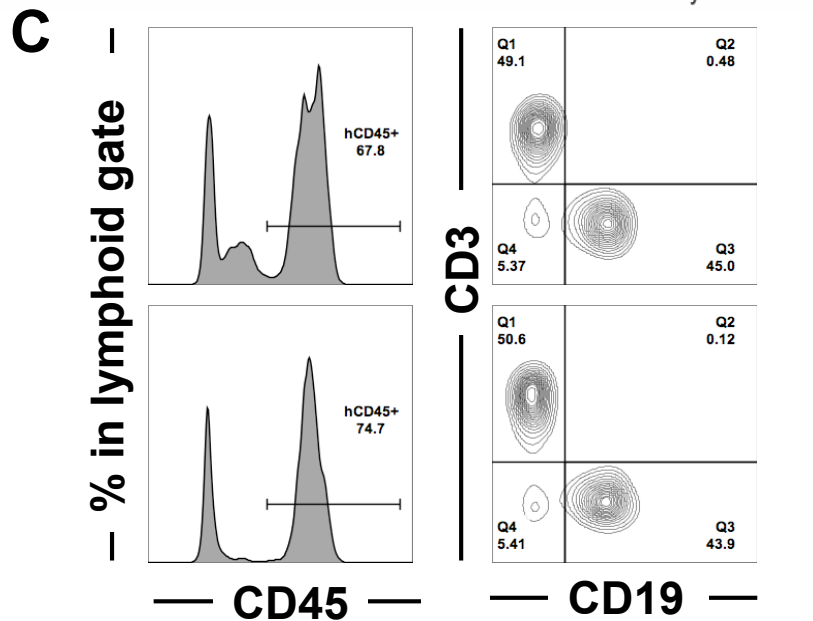
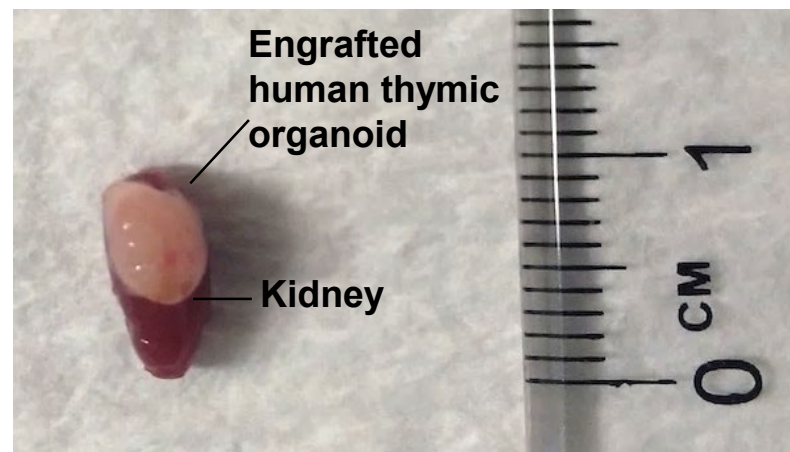
Eric.meffre@yale.edu



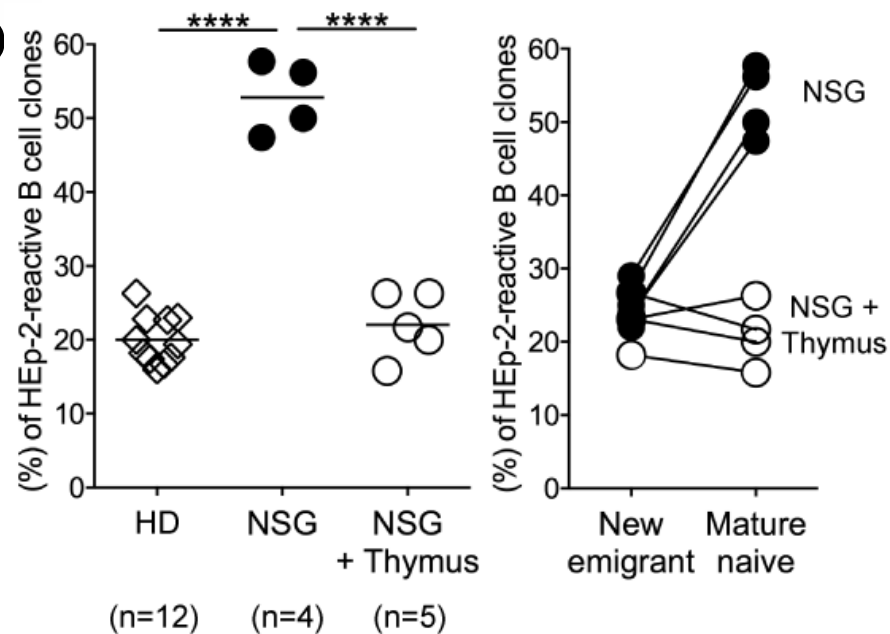
Humanized mouse models to study B cell tolerance



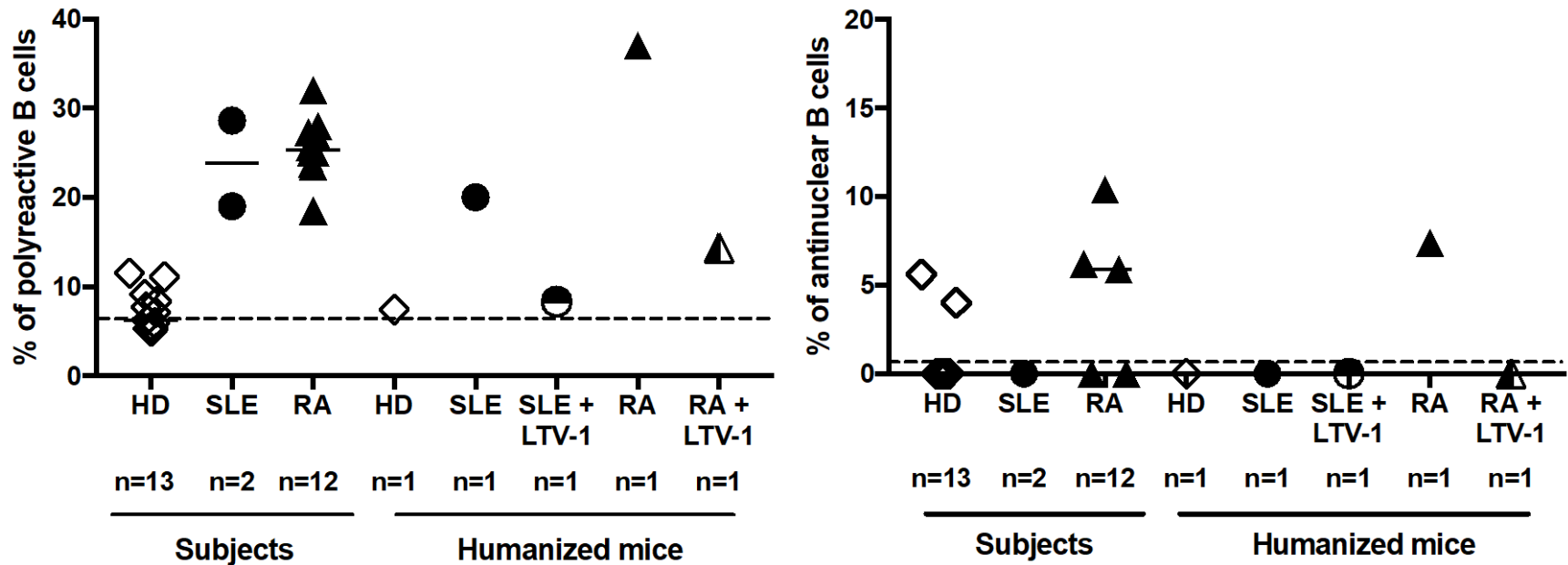
B



D



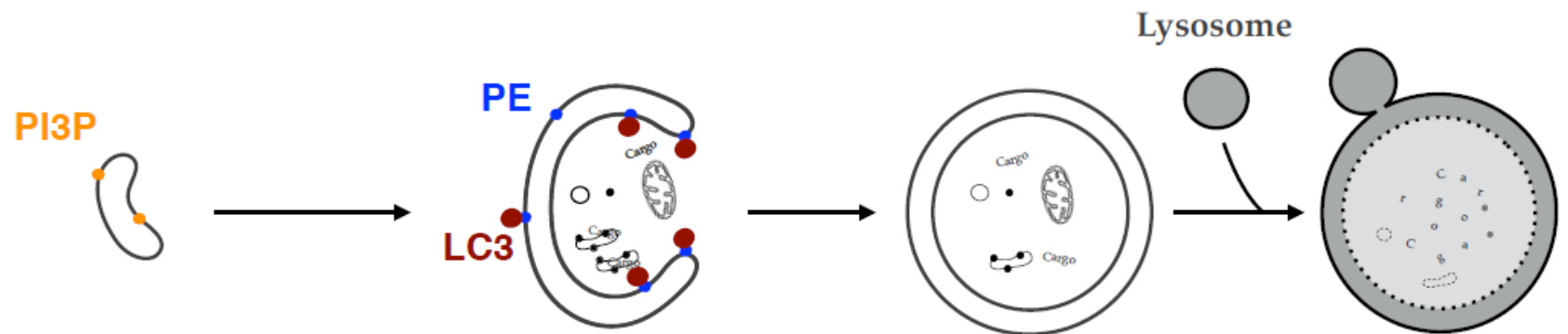
PTPN22 blockade prevents autoreactive B cell production in the bone marrow



PTPN22 inhibition can correct defective central B cell tolerance in systemic lupus erythematosus and rheumatoid arthritis

How cells fight intracellular challenges with the Macroautophagy pathway

The problem: things go catastrophically wrong in the cytoplasm all the time, how do cells get rid of dying organelles, invading pathogens or protein aggregates?



Initiation

Expansion

Closure

Degradation

Early-acting proteins
form the PAS structure

Enzymes attach
ubiquitin-like LC3 to
the growing
membrane

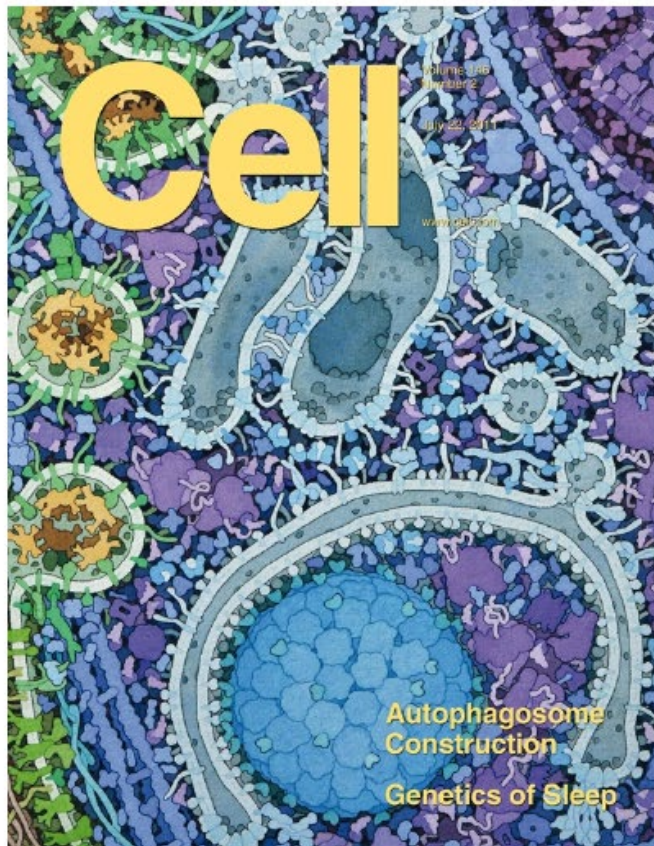
Release of many
autophagy proteins
other than LC3

Fusion with
lysosomes to
degrade cargo

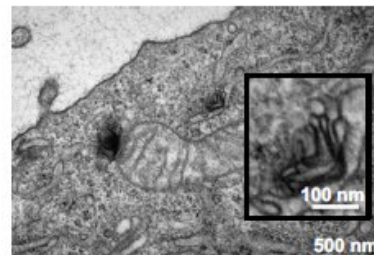
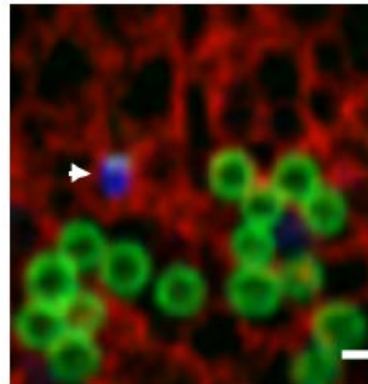
Tom Melia,
Cell Biology Department
thomas.melia@yale.edu
203-785-2165

How cells fight intracellular challenges with the Macroautophagy pathway

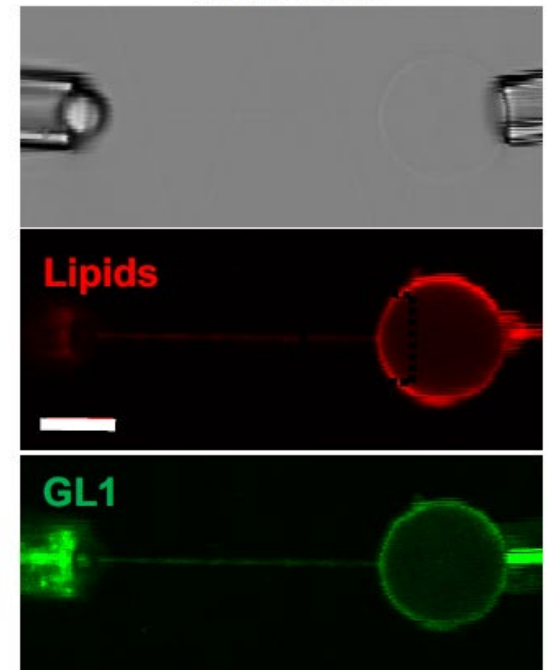
Our approaches: To figure out mechanism, we reconstitute protein activity on artificial membranes that mimic autophagosome structure. To establish biology, we test elements of these mechanisms in gene-edited cell lines.



Follow membrane
formation in cells



Mimic membrane
formation on artificial
membranes



Tom Melia,
Cell Biology Department
thomas.melia@yale.edu
203-785-2165

How cells fight intracellular challenges with the Macroautophagy pathway

What have undergraduates done: Most summer students start off learning proteo-membrane reconstitution and explore fundamental activities of simple protein machines.

Yale **Other Institutes**

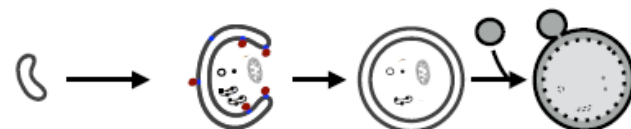
Worked in my lab for a summer

Then decided to continue into school year

Have appeared on at least 1 paper

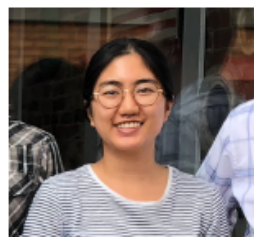
Have data for papers not yet in submission

11	6
8	2*
4	4
3	1



Summer project: used DNA-scaffolded liposomes to study enzyme activity on membranes with curvature similar to parts of autophagosome (collaboration with Lin lab)

Thesis project: Describing first ever disease phenotype of a patient discovered in Yale Genomics facility with mutations in Laurie's favorite autophagy enzyme.

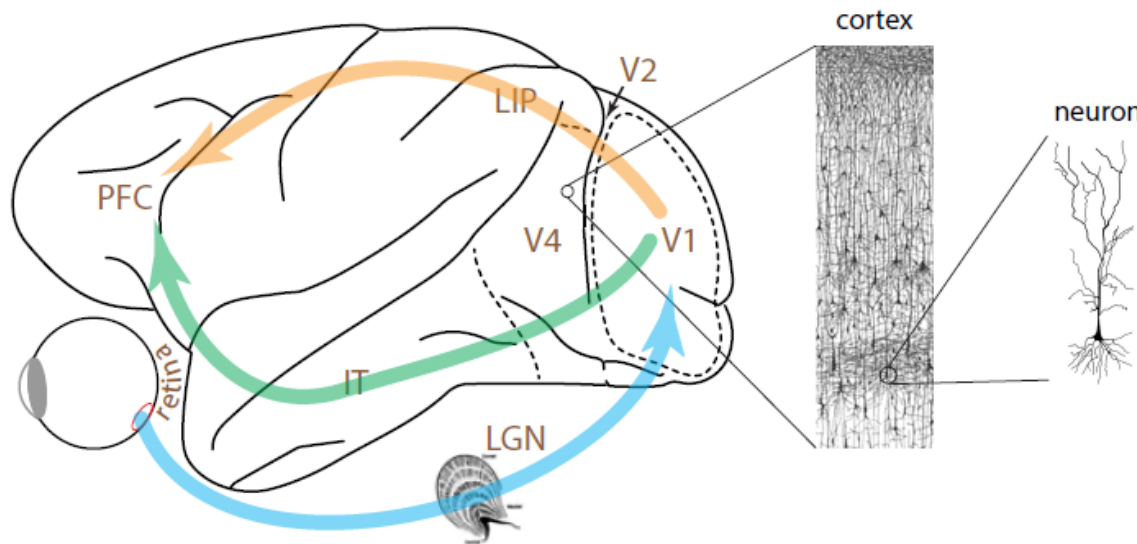


Laurie Wang
Class of 2019

PI: Anirvan Nandy (anirvan.nandy@yale.edu)
Graduate Student Mentor: Feng Xing (feng.xing@yale.edu)

Dept. of Neuroscience

Neural mechanisms of perception, attention & social cognition

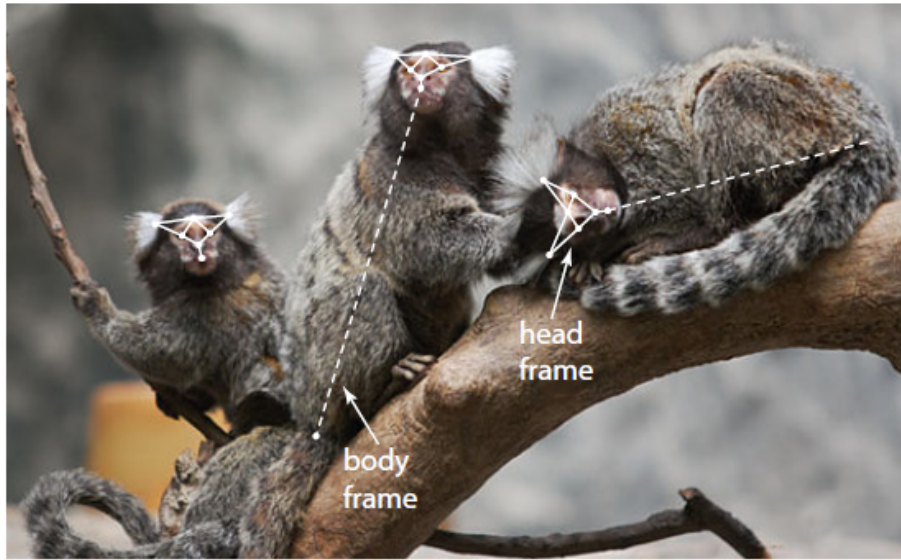


How do neurons in the visual cortex encode information about shapes of objects?

How is this encoded information changed by attention?

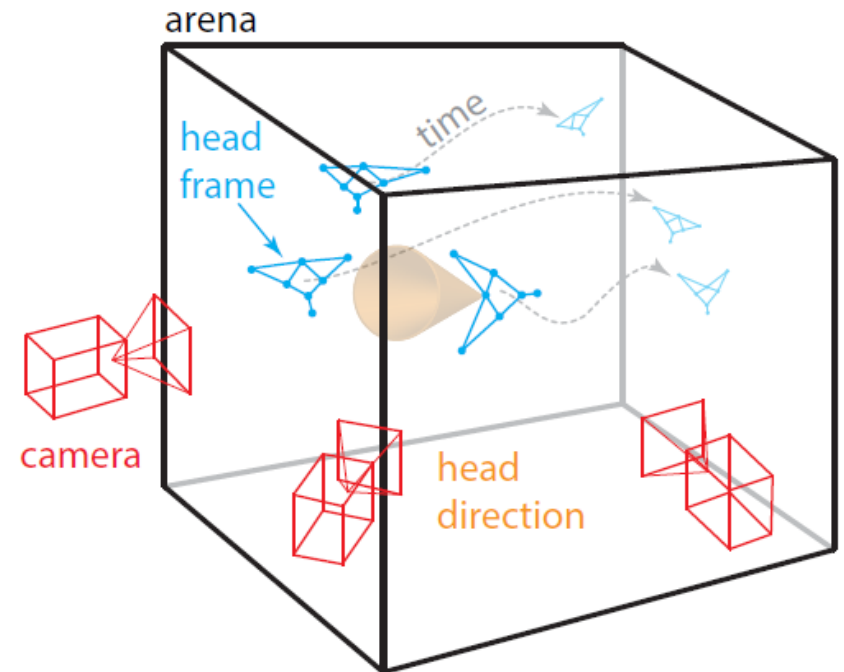
How do neurons in the pre-frontal cortex encode information about complex social interactions?

An important first step in understanding complex social cognition is an accurate quantification of behavior



We will use marmoset monkeys as a model system since they are highly pro-social animals

We will track interacting groups of marmosets using multiple cameras and use advanced computer vision and machine learning methods to accurately quantify their behavior



Roles & Expectations:

Student will be actively mentored daily by the graduate student mentor

Student will meet with PI weekly

Student will participate in weekly lab meetings

Position is available for one funded student

Thank You!

<http://nandylab.org>

Functional genomics of adaptive radiation in Antarctic fishes



thomas.near@yale.edu



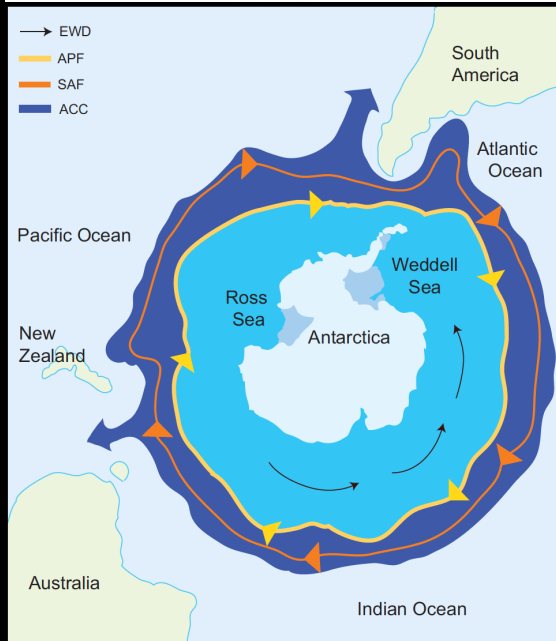
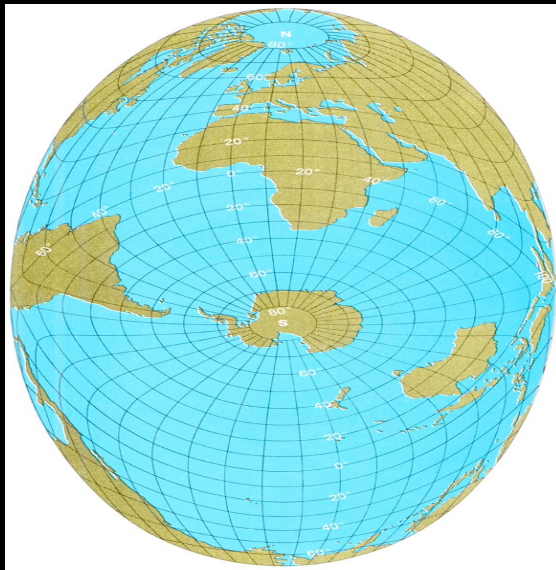
@tjnear



Thomas J. Near

*Dept. Ecology and Evolutionary Biology and Peabody Museum of Natural History
Yale University*

Southern Ocean Fish Fauna



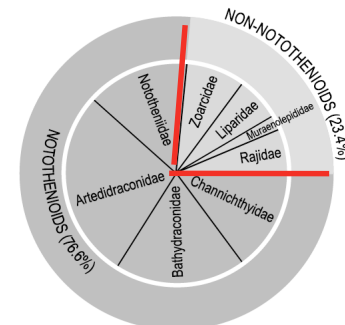
Pogonophryne marmorata



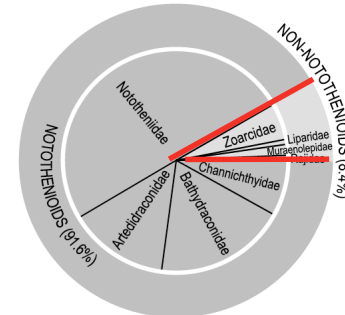
Chionodraco myersi



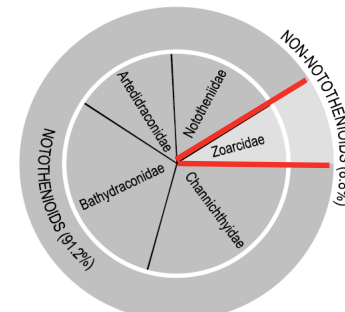
Trematomus borchgrevinki



% By Species



% By Number of Individuals



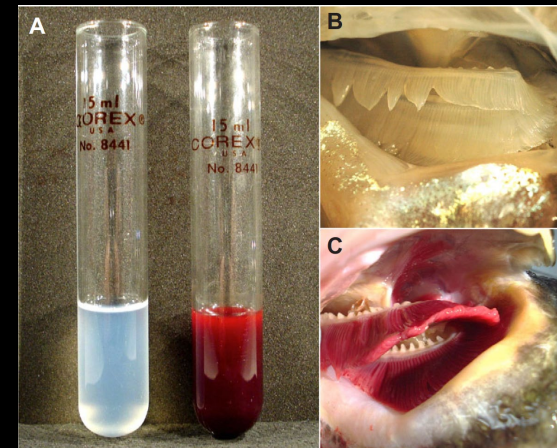
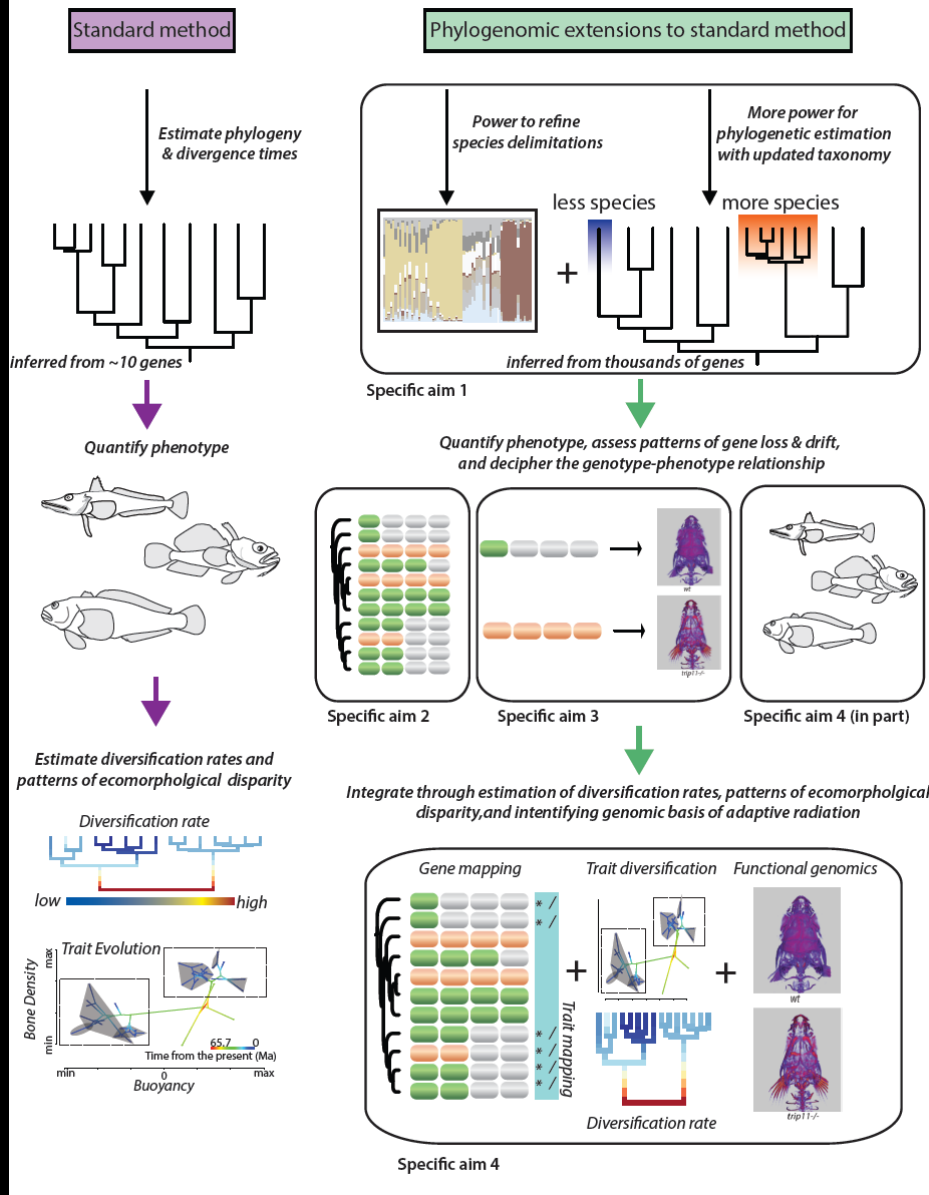
% By Weight

Diversity
76.6%

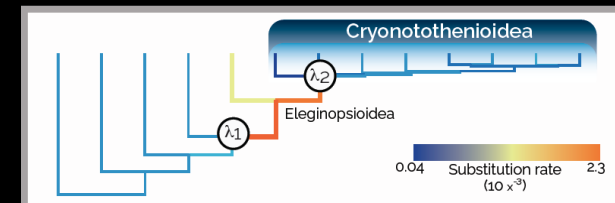
Abundance
91.6%

Biomass
91.2%

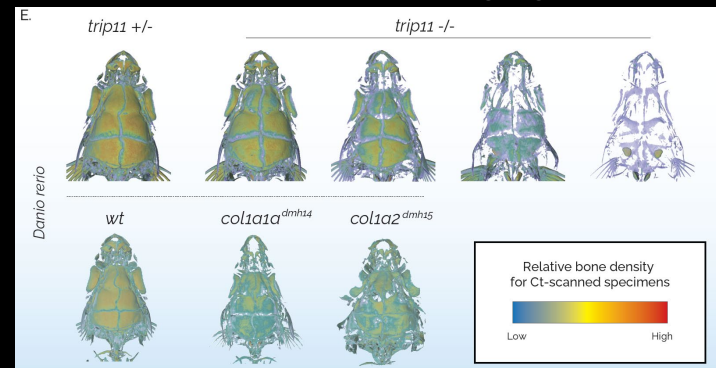
The study of adaptive radiation: the phylogenomic approach



Phylogeny showing high rates of genomic evolution

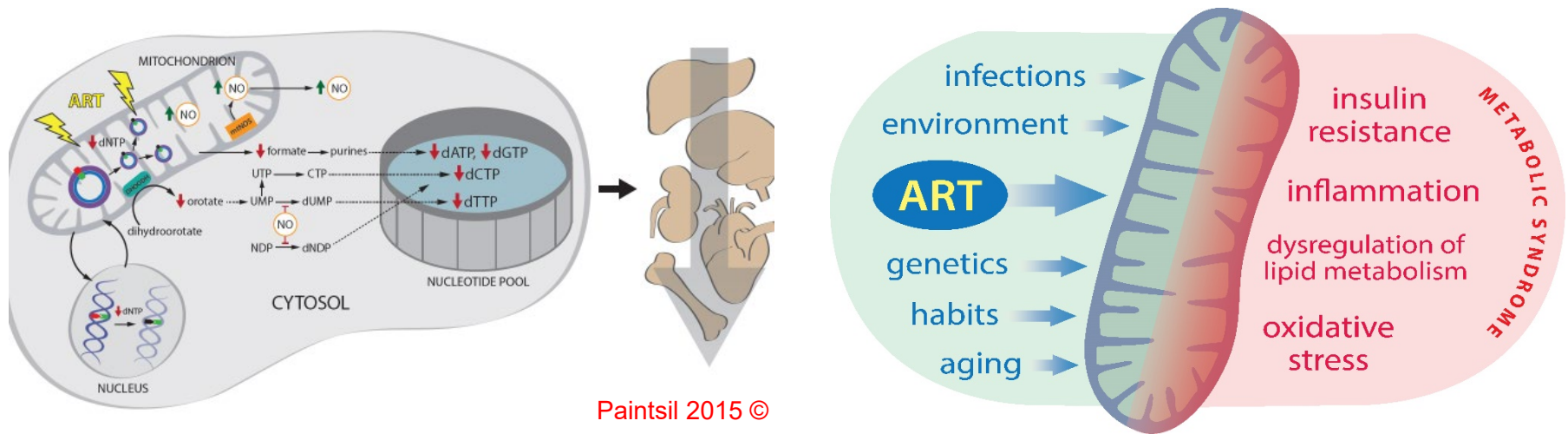


CRISPR-Cas9 induced mutations on target genes in Zebrafish



Paintsil Lab

Effects of Antiretroviral Therapy on Mitochondrial Function



“Our central hypothesis is that Antiretroviral Therapy (ART)-induced mitochondrial dysfunction is in the causal pathway of ART-associated toxicities:- metabolic syndrome, accelerated aging, and aging-related disorders such as cardiovascular diseases, cancer, and neurodegenerative diseases”

Elijah Paintsil, MD
 Yale Child Health Research Center
 464 Congress Avenue, New Haven
 Email: Elijah.paintsil@yale.edu

Projects Available for Students

- **Comparative effects of older and new generation nucleoside analogs on mitochondria DNA polymerase gamma expression**
- **Comparative effects of older and new generation nucleoside analogs on mitochondrial content in peripheral mononuclear cells and T-cell lines**
- **Identifying biomarkers of ART-induced mitochondrial toxicity**
- **Effects and underlying mechanism(s) of ART on metallothionein 1 gene expression**
- **Effects and underlying mechanism(s) of ART on cholesterol biosynthesis**

Mentoring Expectation

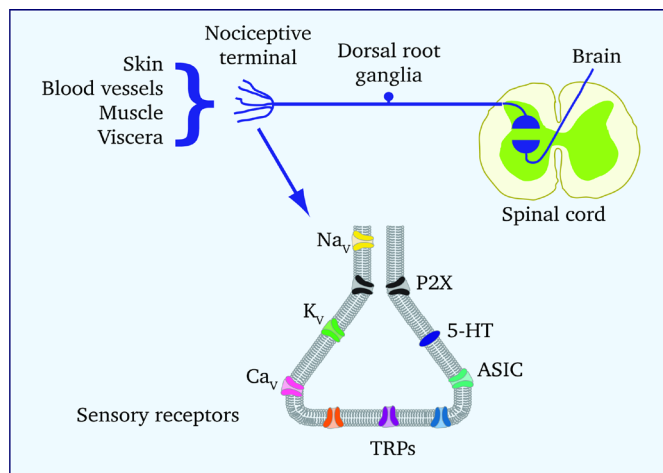
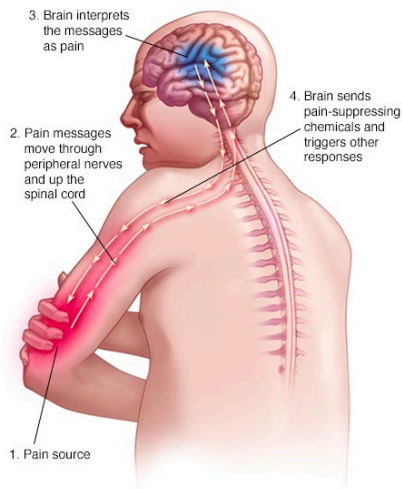
- I will meet with student daily to discuss progress
- Day to day mentoring of student by a Postdoc in the Lab
- The student is expected to participate in weekly Lab meetings
- We can take on 2 students
- The last 2 summers, we have hosted one Yale undergrad
- Their work resulted in manuscripts



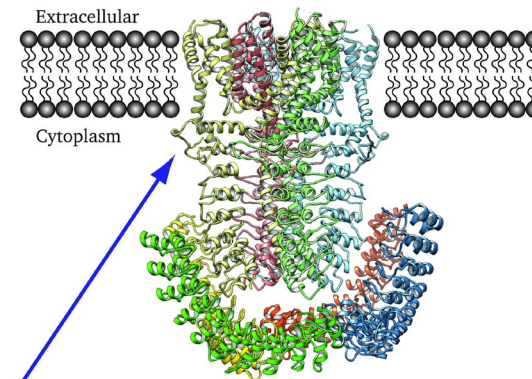
The Paulsen Lab



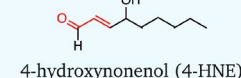
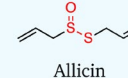
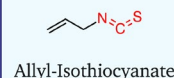
Studying molecular mechanisms of pain at Yale University



TRPA1



Activators



Candie Paulsen

Candice.Paulsen@yale.edu

Ongoing projects in the Paulsen lab

1. Determine the structure of TRPA1 in its open, active conformation (Peter Niimi)
(membrane protein biochemistry, negative stain EM, cryo-EM)
2. Identify TRPA1 interacting protein partners (Justin Sanders)
(chemical biology, biochemistry, fluorescence microscopy, electrophysiology)
3. Express and purify the TRPA1 intracellular N-terminus for structural and biochemical analyses (Samantha Schaefer)
(biochemistry; ultimately X-ray crystallography and pull down experiments)
4. Determine the structure of a human-rattlesnake TRPA1 chimera (Justin Sanders)
(molecular biology, fluorescence microscopy, membrane protein biochemistry)
5. Characterize TRPA1's role in signal transduction cascades (TBD)
(biochemistry)

Summer research structure

During the summer, the students (2) will...

- Meet with me **daily** to touch bases and establish an experimental plan
- Be mentored in the lab by **Justin**, **Peter**, and **Samantha**
- Attend all lab group meetings and journal clubs
- Give an end of summer presentation on their work to the lab

Death begets a new beginning

Carla V. Rothlin

HHMI Faculty Scholar

Associate Professor of Immunobiology
and Pharmacology

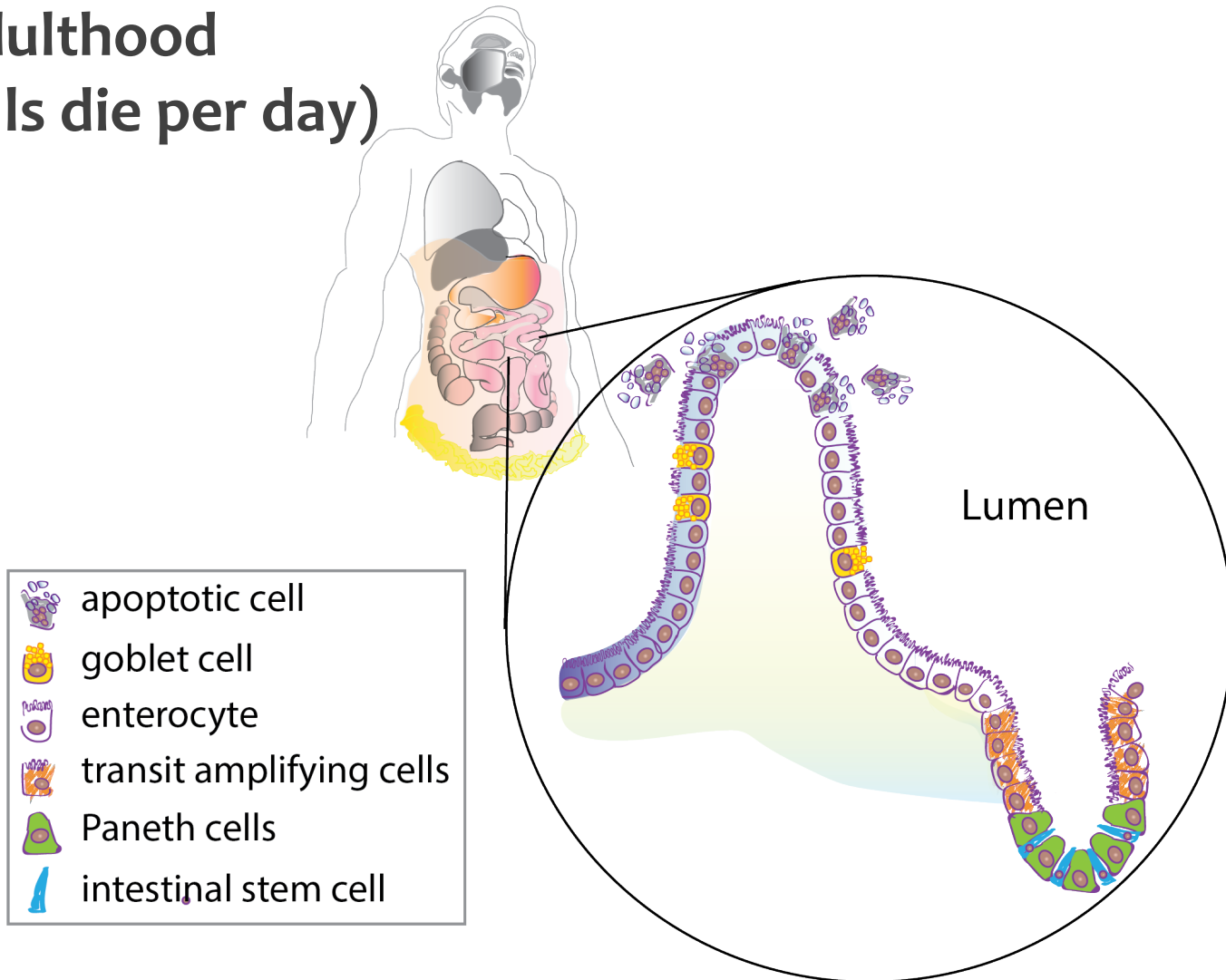
Yale School of Medicine



What happens after death?

What happens after cell death?

Adulthood ($>10^9$ cells die per day)



Development

Injury

Homeostasis

Degeneration

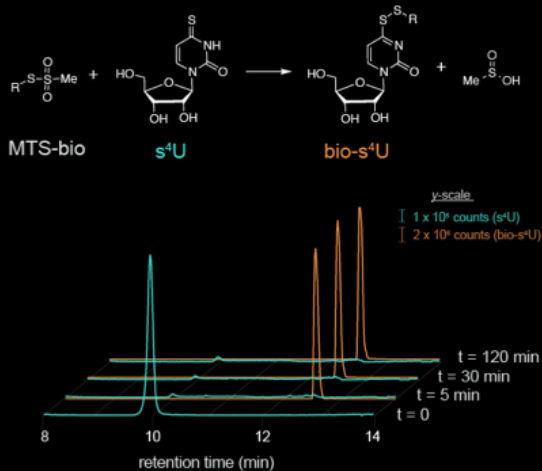
Infection



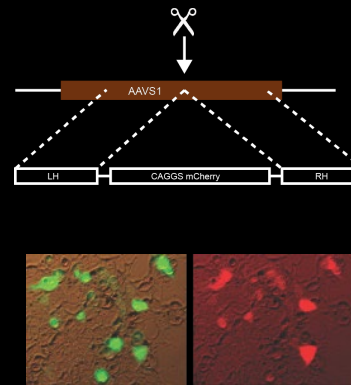
Cell death recognition code

Research in the Simon Lab: An integrated approach to studying RNA and chromatin biology

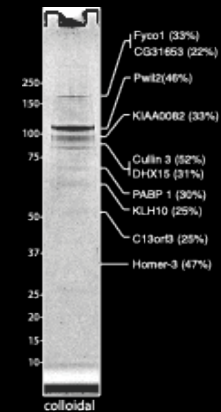
nucleoside chemistry



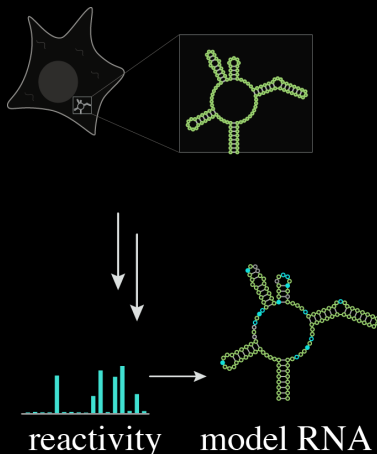
genome editing



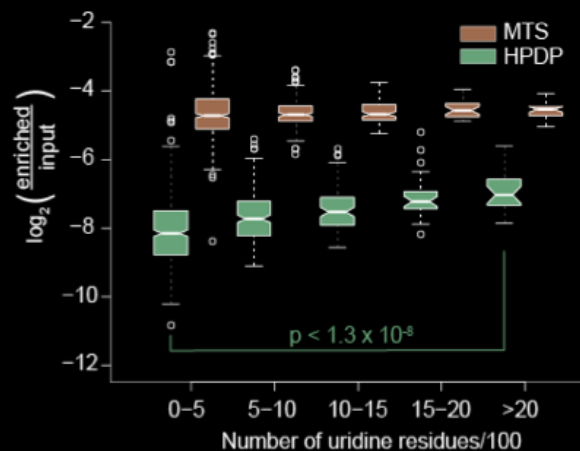
biochemical purifications



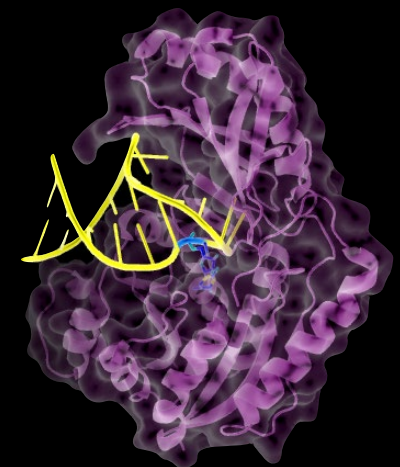
RNA structural probing



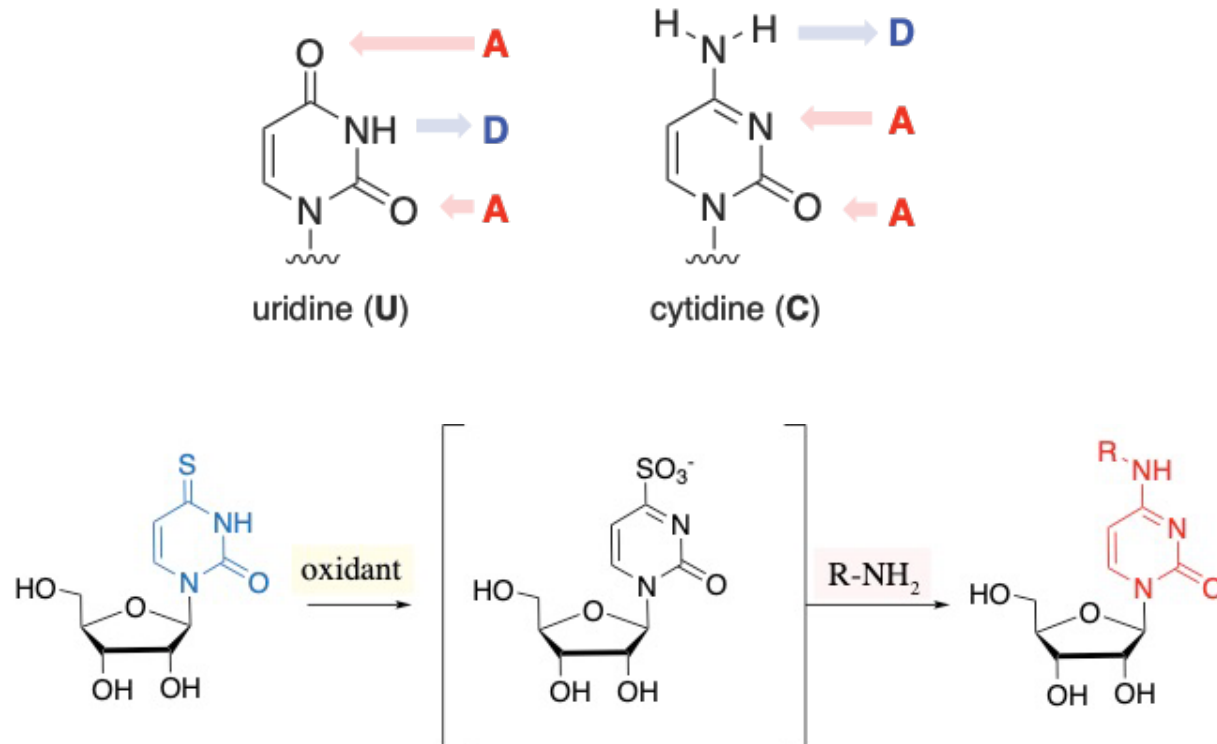
genomics



protein engineering



Project: Nucleotide recoding to reveal RNA population dynamics



- Learn about the **chemical biology of nucleotides**.
- Learn **computational tools** to examine changes in global RNA populations.
- Use these tools to **understand the biology of how cells adapt** to changes in their environment.

Two undergraduate success stories: Catherine Stark and Peter Wang



Duffy, E. E., Rutenberg-Schoenberg, M., **Stark, C. D.**, Kitchen, R. R., Gerstein, M. B., and Simon, M.D., (2015) Tracking distinct RNA populations using efficient and reversible covalent chemistry. *Mol Cell*, 59(5), 858-66.

Sexton, A.N., **Wang, P.Y.**, Rutenberg-Schoenberg, M. and Simon, M.D. (2017). Interpreting Reverse Transcriptase Termination and Mutation Events for Greater Insight into the Chemical Probing of RNA. *Biochemistry*, 56(35):4713-4721.

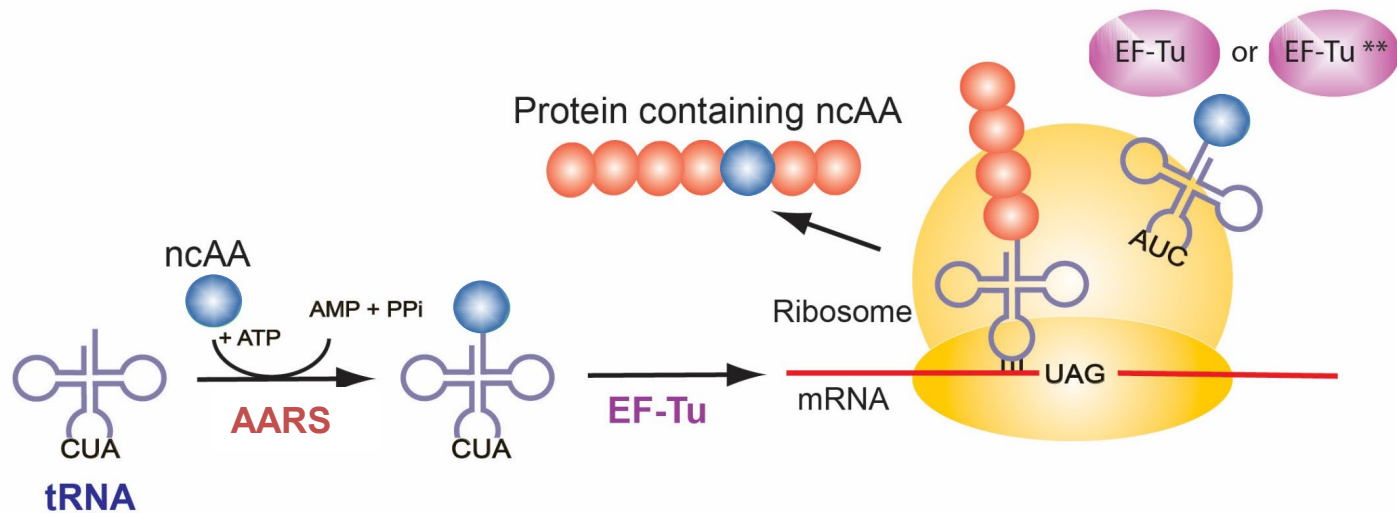
Wang, P. Y., Sexton, A. N., Culligan, W. J., & Simon, M. D. (2018). Carbodiimide reagents for the chemical probing of RNA structure in cells. *RNA*, rna.067561.118.

Dieter Söll / Kyle Hoffman

Department of Molecular Biophysics & Biochemistry

Yale University

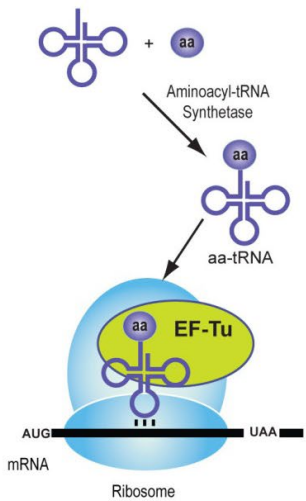
Adding amino acids to the genetic code



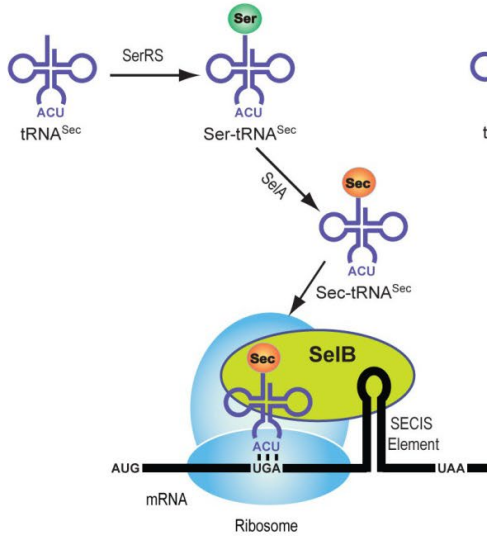
Develop variants of tRNA, aminoacyl-tRNA synthetases, elongation factors, and ribosomes

Re-wire translation for site-specific Sec insertion

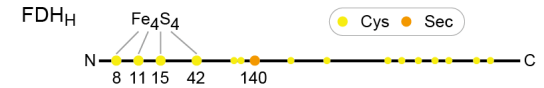
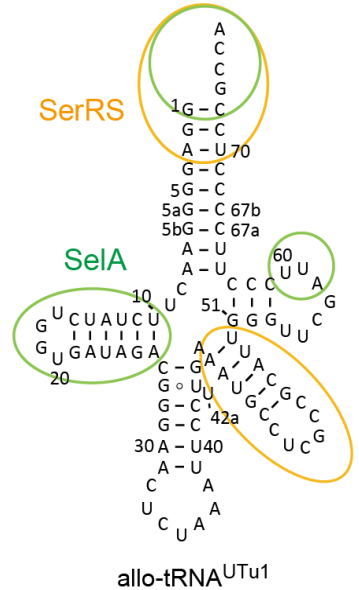
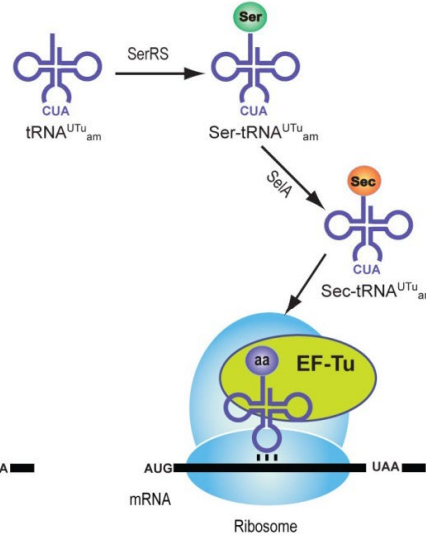
Normal AA insertion



Normal Sec insertion



Re-wired Sec insertion



		As SerA											
		-	+	+	+	+	+	-	+	+	+	+	+
allo-tRNA		+	+	+	+	+	+	+	+	+	+	+	+
FDH _H	8	Cys	Cys	Am	Am	Am	Am	Cys	Cys	Ser	Cys	Cys	Cys
	11	Cys	Cys	Cys	Am	Am	Am	Cys	Cys	Cys	Ser	Cys	Cys
	15	Cys	Cys	Cys	Am	Am	Am	Cys	Cys	Cys	Cys	Ser	Cys
	42	Cys	Cys	Cys	Am	Am	Am	Cys	Cys	Cys	Cys	Cys	Cys
	140	Am	Am	Am	Am	Am	Am	Am	Am	Am	Am	Am	Ser
			●	●	●	●	●		●				

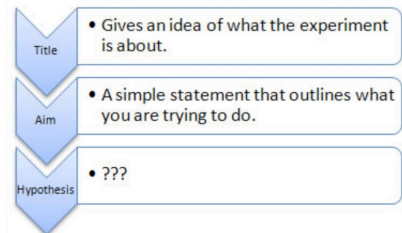
Challenge: Design of synthetic tRNA^{UTu}

How you will be trained

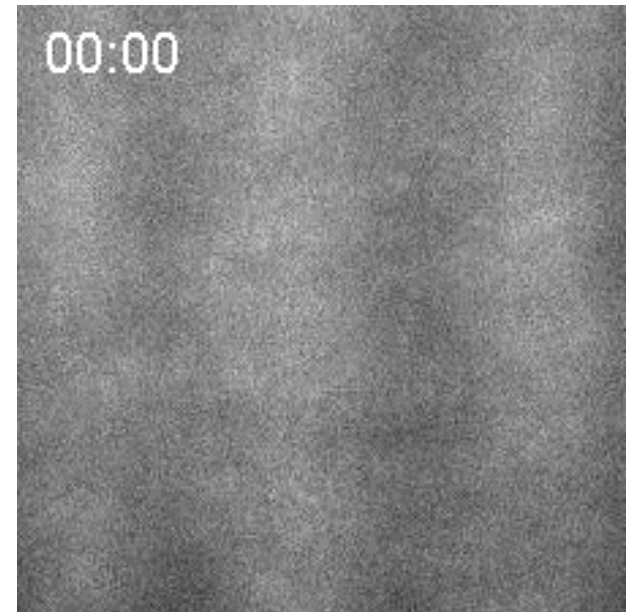
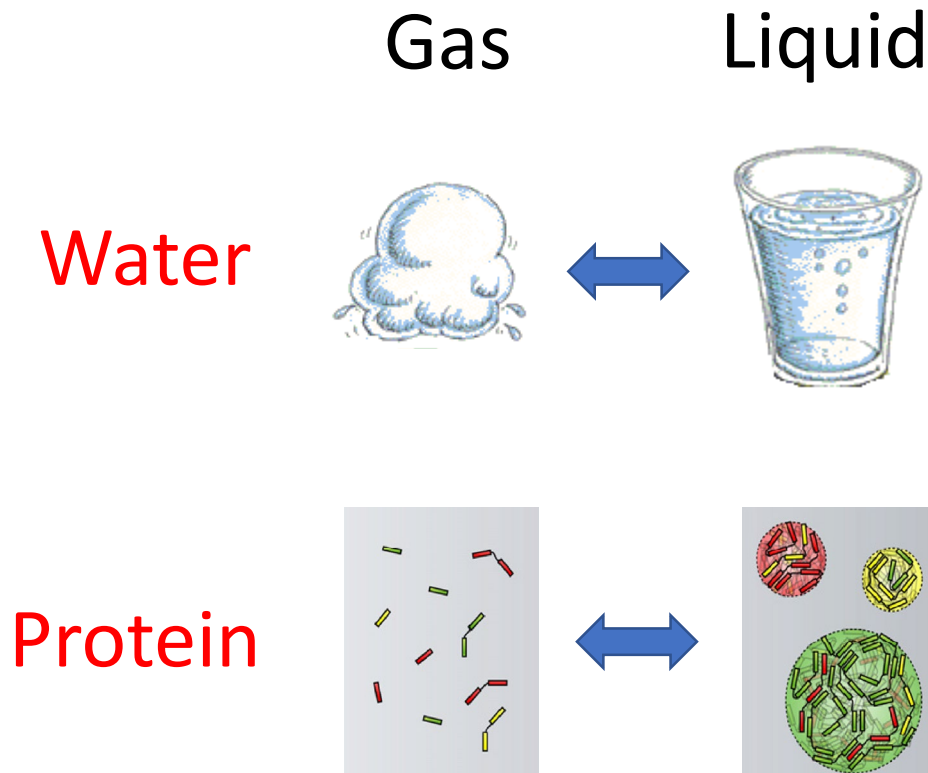
- Doing research: working side by side with a postdoc who will train you to become 'independent'
- Participate in weekly lab meetings
- Writing a short final report
- Giving a PowerPoint presentation to the group before departure
- Undergraduate seminar presentation
- GOAL: self confidence that allows you to tackle research in the future



Writing scientific reports



Phase Transition in Cell Signaling



Su*, Ditlev*, *Science* 2016

Xiaolei Su

xiaolei.su@yale.edu

www.sulab.net

Summer Projects

- How do lipids regulate phase transition during T cell activation?
- How does phase transition promote chimeric antigen receptor (CAR) activation?

Mentoring

Summer students last year:

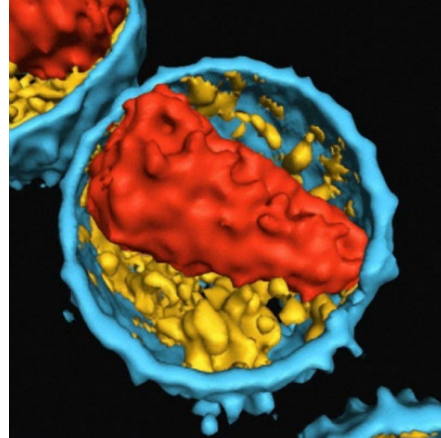
Kendra Libby (class '21, kendra.libby@yale.edu)

Hannah Triscott (class '19, hannah.triscott@yale.edu)

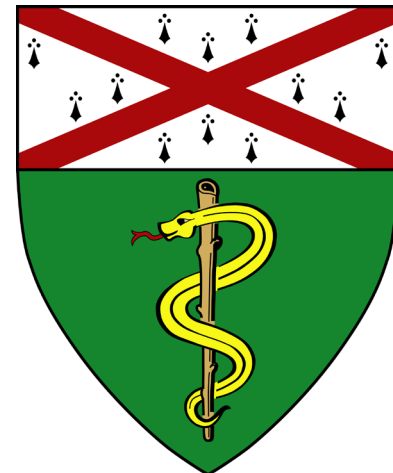


- Independent project
- I (PI) am the mentor
- Daily interactions
- Bi-weekly presentation
- Working together on proposal and progress report

Host Genetic Control of HIV/HIV Cure



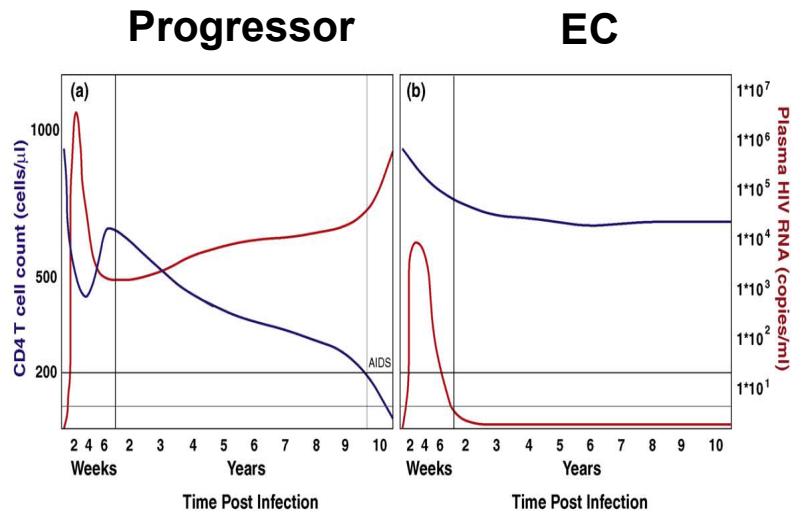
My
office



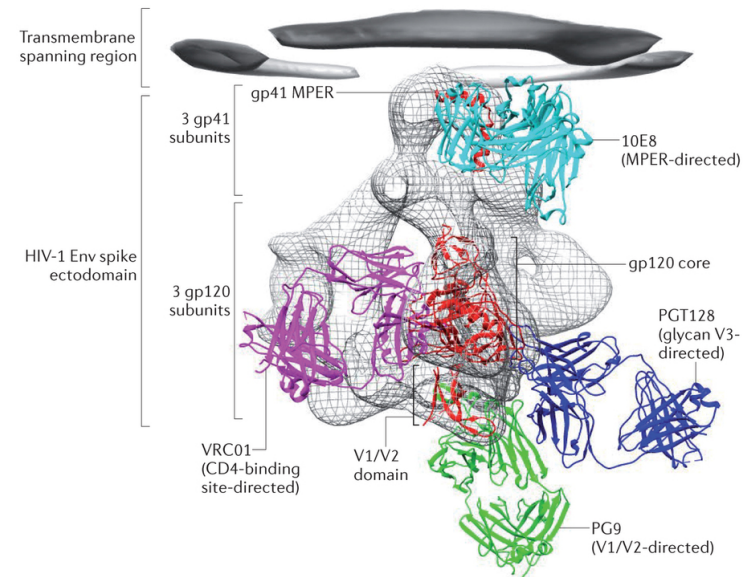
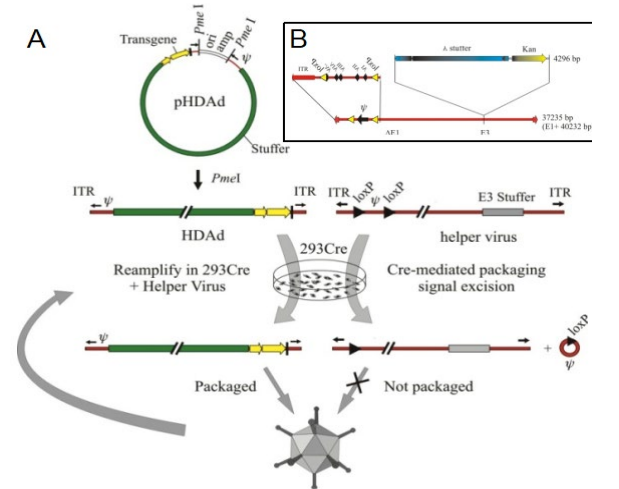
Richard Sutton, MD-PhD
Section of Infectious Diseases
& Department of Microbial
Pathogenesis
Yale University School of Medicine
Richard.Sutton@yale.edu

Possible Projects:

- Post-transcriptional regulation of *ccr5*
- Vectored delivery of anti-HIV bnAbs
- Study of *cnpy4* in HIV elite control
- KO of *ccr5* in hematopoietic cells



TRENDS in Pharmacological Sciences



The student experience:

- Will work directly with post-doc or me (I am usu around)
- Typically we have 2-3 undergrads in the lab
- BSL1 or BSL2, focus is molecular biology
- Get to give lab meeting!
- Recent former undergrads: med school, applying to med school, working in translational or clinical research (NYU & Pitt), grad school in chemistry (here)

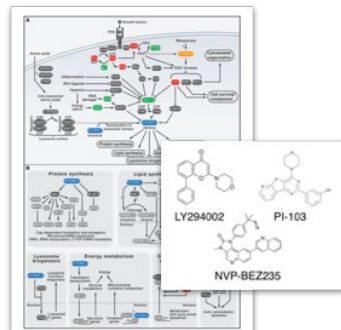
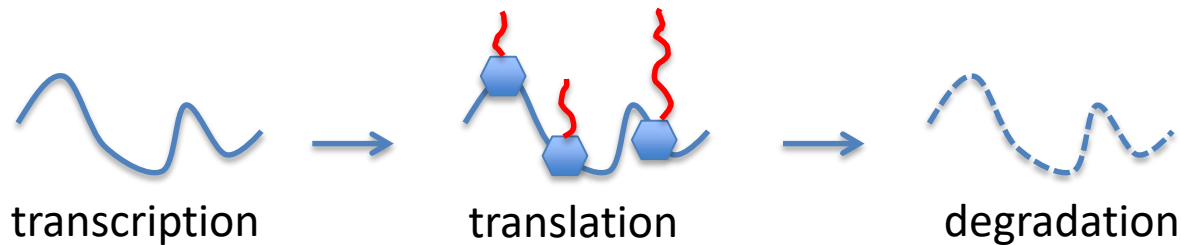
Carson Thoreen, PhD

Cellular and Molecular Physiology

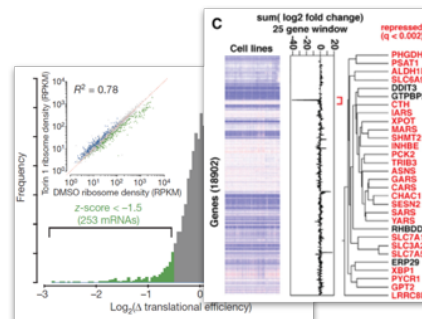
carson.thoreen@yale.edu

campuspress.yale.edu/thoreenlab

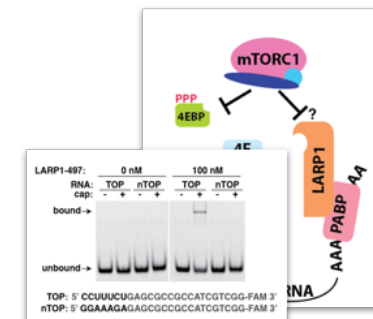
Post-transcriptional regulation of complex programs of gene expression



Signaling



Bioinformatics



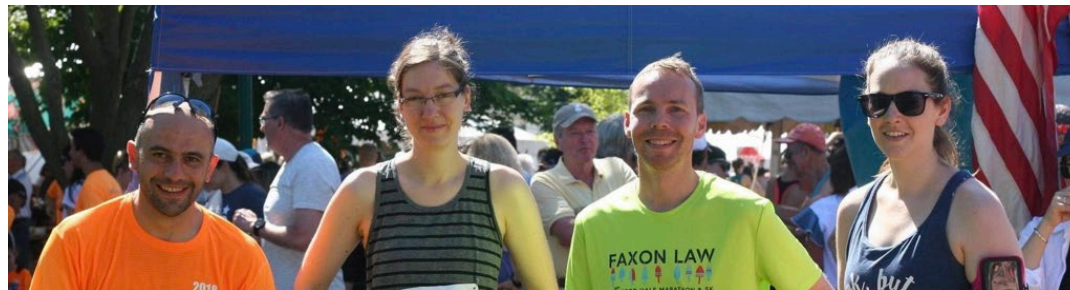
Biochemistry

Potential summer projects

1. Identify mRNA sequences that control translation using a reporter system.
2. Bioinformatic discovery of sequences that destabilize mRNAs.
3. Biochemical study of interactions between a regulatory protein and its RNA targets.

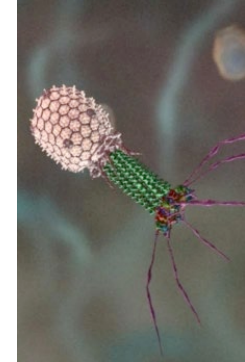
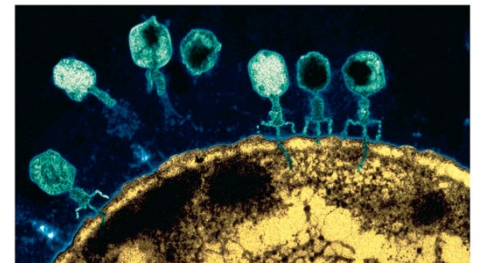
How the lab works

- Paired with post-doc.
- Participate in bi-monthly meetings that alternate between data and journal club.
- Meet with me on a weekly basis.



Turner Lab for Virus Evolution and Virotherapy

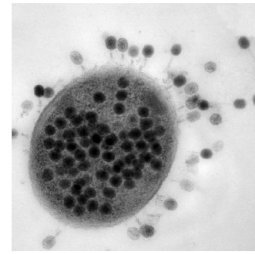
- How do viruses evolutionarily adapt to new environments, such as novel hosts?
- Which virus traits are useful for biotechnology, such as developing novel therapies?



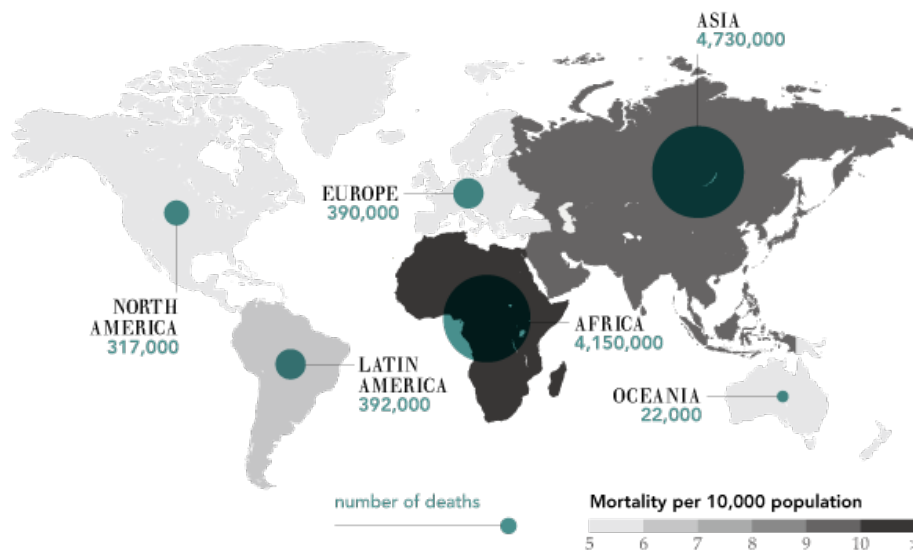
Paul E. Turner
Professor of Ecology & Evolutionary Biology, Yale U.
Microbiology Faculty, Yale School of Medicine.
paul.turner@yale.edu

Summer Project

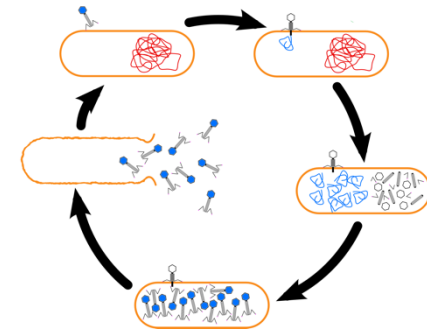
- Discover and characterize novel viruses (bacteriophages).
- Test how the phages select for evolution of reduced virulence in bacteria.
- Determine best phage candidates to develop as novel therapies targeting drug-resistant bacterial pathogens.



Annual deaths from drug-resistant infections by 2050

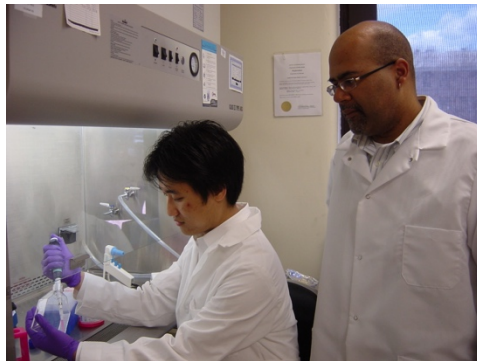
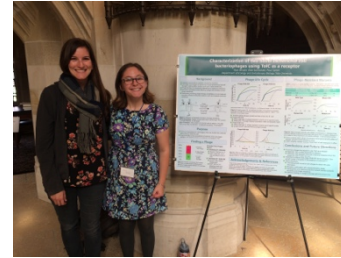


Source: The Review on Antimicrobial Resistance



Mentoring Plan

- Daily interaction with direct research mentor (graduate student, or postdoc)
- Weekly meetings with Professor Turner
- Weekly group lab meetings
- Goal: Train you to become a creative and independent scientist!



Dr. Yul Yang, MD
Yale Class '07



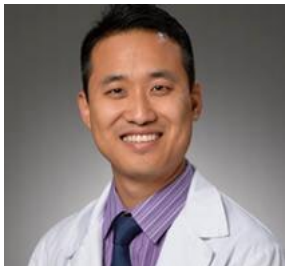
Dr. James Pease, PhD
Assistant Prof, Wake Forest U
Yale Class '07



Mary Ann Santucci
Yale Class '18



Cecilia Sanchez, Derek Park,
Kelly Diaz, Kristen Brao
Yale Class '13

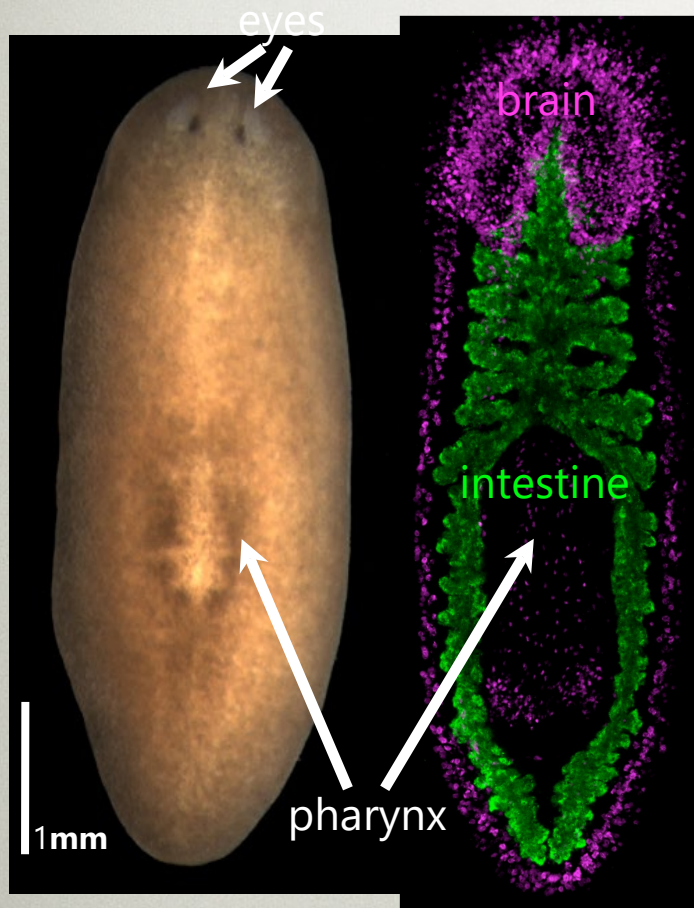


Earl Chism
Yale Class '18

Josien van Wolfswinkel
josien.van.wolfswinkel@yale.edu
vanwolfswinkellab.org

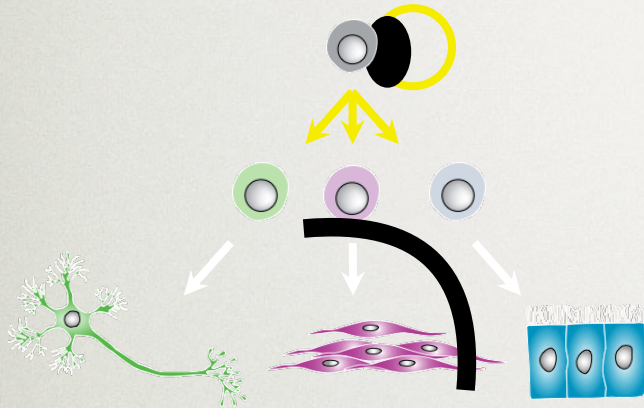
Planarian regeneration and stem cell biology

Schmidtea mediterranea



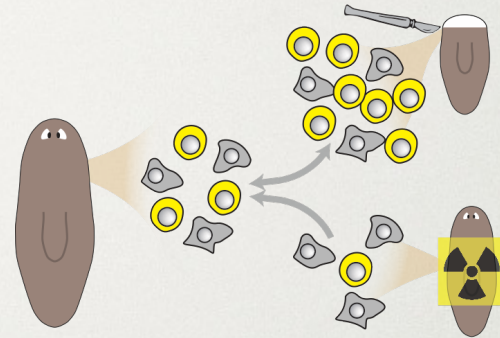
Lab projects

Regulation of pluripotency and cell specification



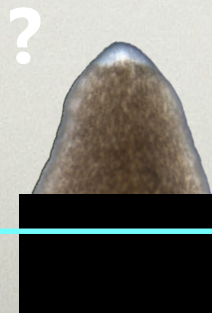
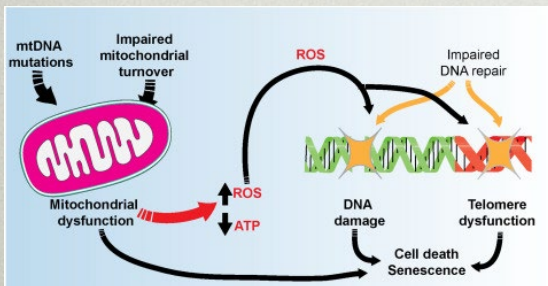
How do stem cells make decisions?

Stem cell control



How do planarians avoid cancer and aging?

Metabolism and regeneration



Making transgenics!

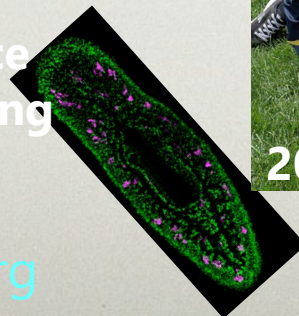


Visit us on the 10th floor of KBT!

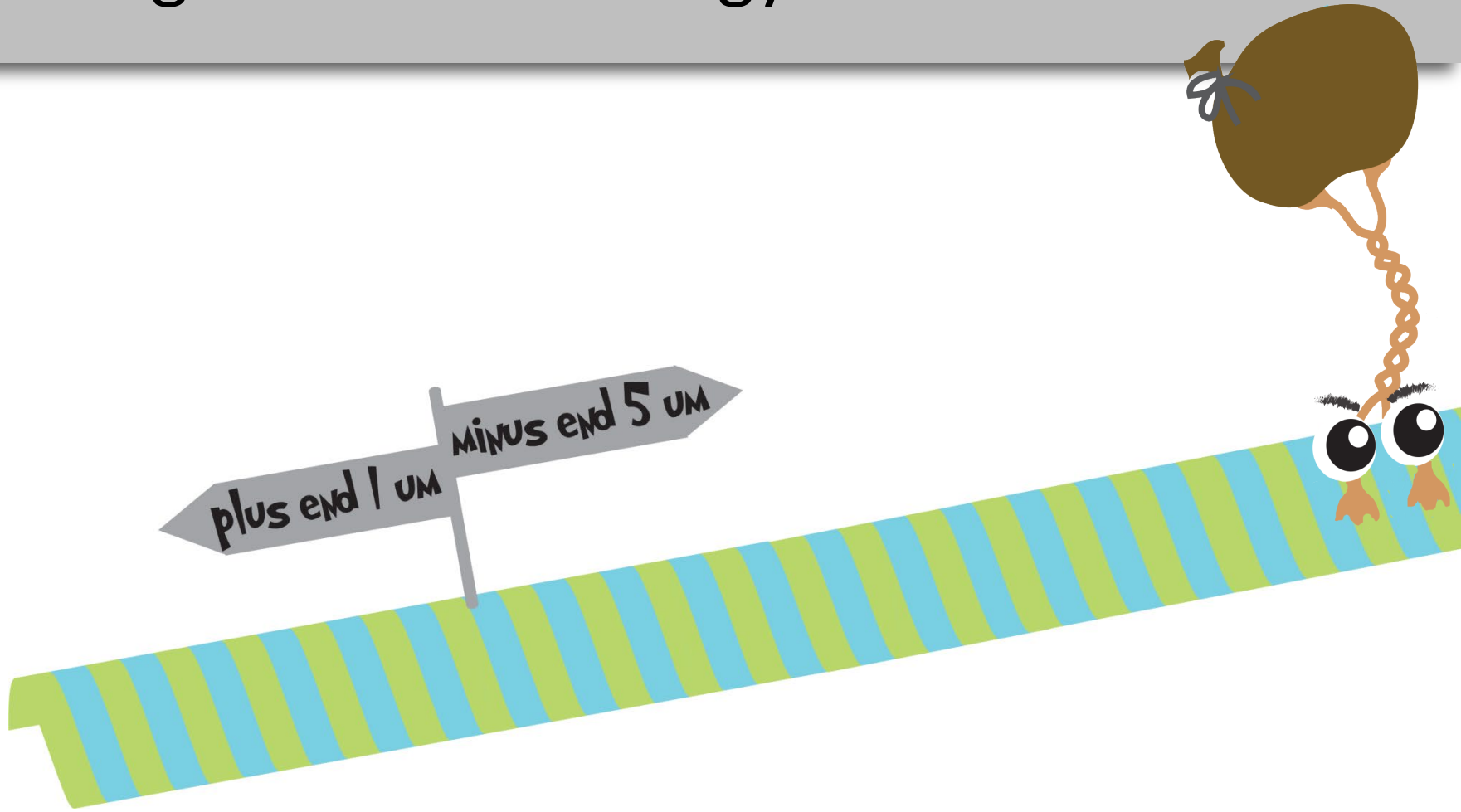


Yale stem cell center
Yale center for RNA Science
Yale Pepper center for Aging

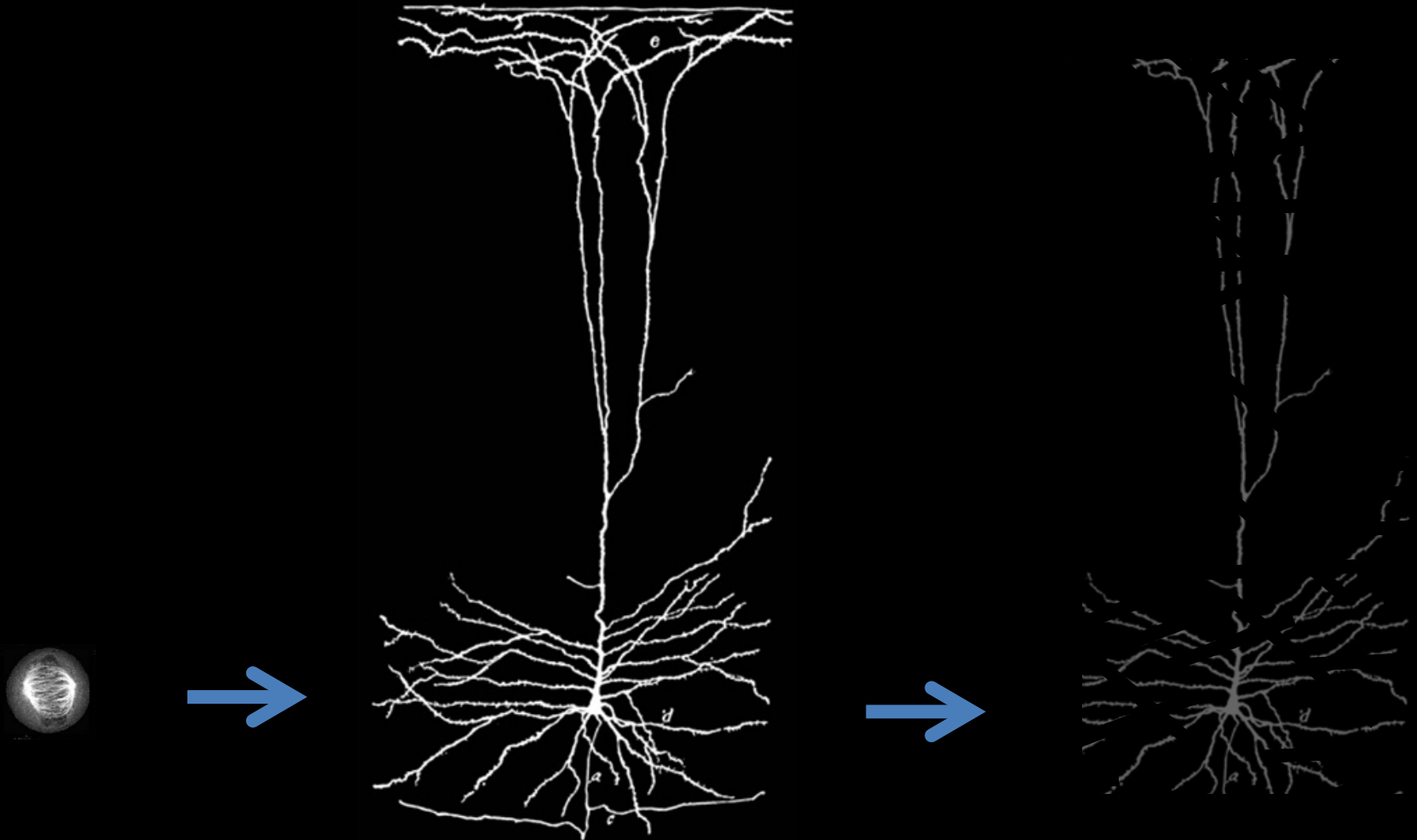
www.vanwolfswinkellab.org



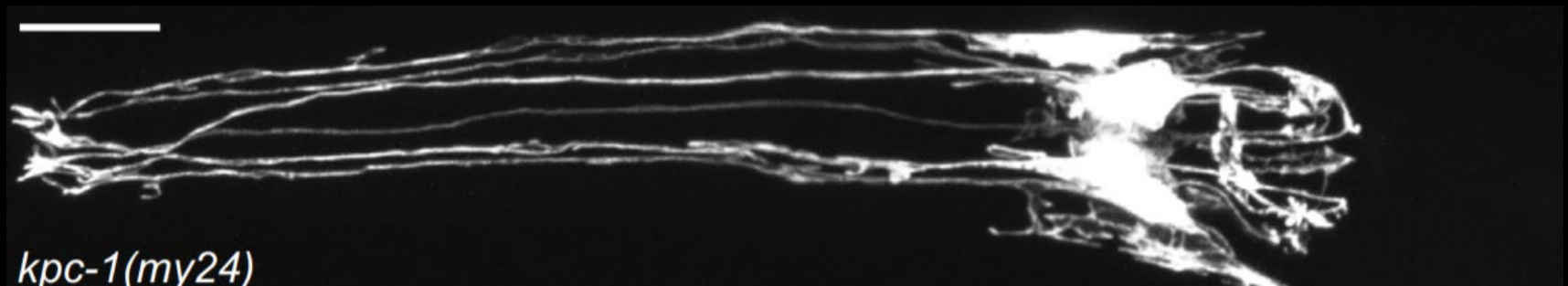
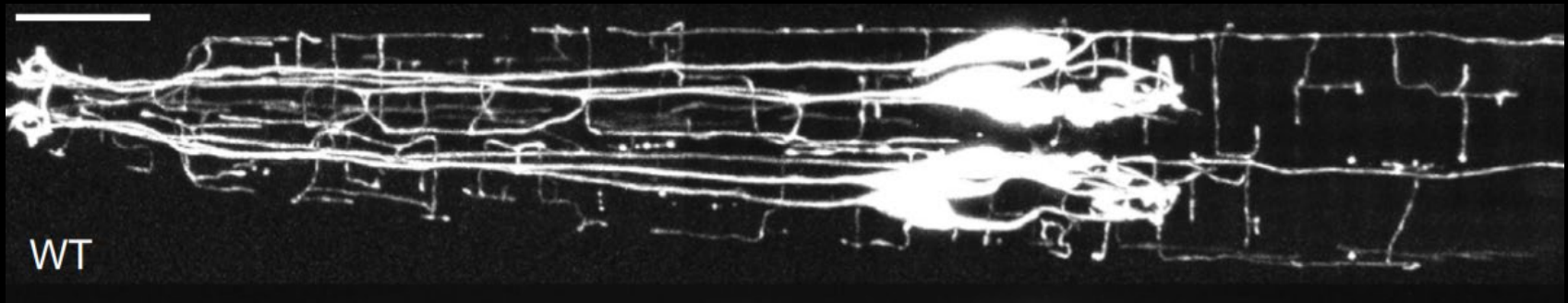
Yogev lab: Cell biology of the neuron



We are interested in neuronal cell biology – how neurons polarize, grow and remodel



Summer project: study the mechanisms of neuronal remodeling in *C. elegans*

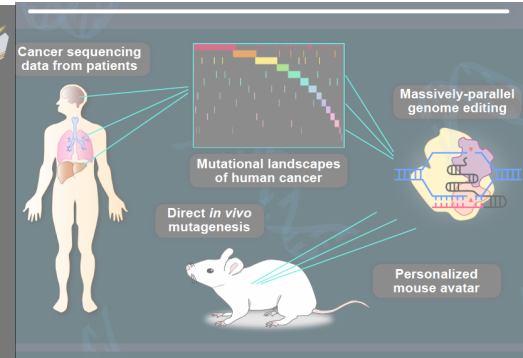
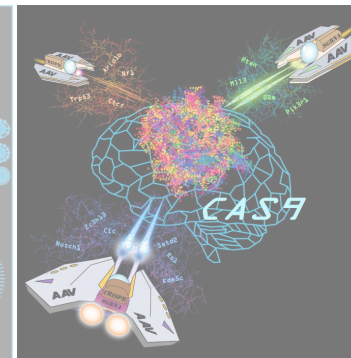
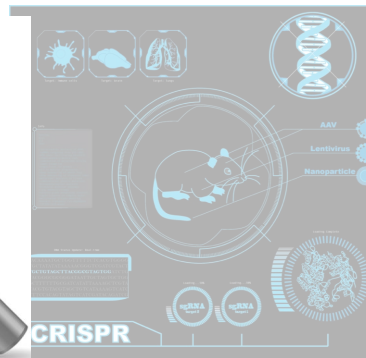


Methods: genetics and imaging

Mentoring by PhD student (daily) and PI (weekly)

Faculty – Student / Yale 2019-01-04

Lab Focus: Genome editing and Cancer Immunology



Sidi Chen, PhD

Group Leader, Assistant Professor

NIH Director's New Innovator (2018)

Department of Genetics & Systems Biology Institute

Yale Cancer Center & Stem Cell Center

Yale University School of Medicine

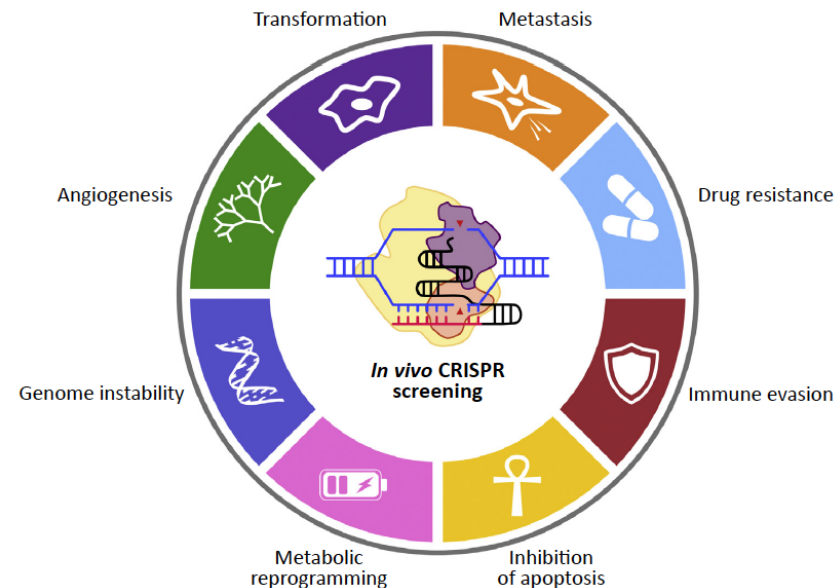
sidi.chen@yale.edu



Yale University

Student projects

- High-throughput mapping of functional drivers
- In vivo screens for novel drug targets
- Immune engineering
- Genetic interaction mapping
- Tumor immunity
- Development of novel biotechnologies
- Single cell sequencing



Trends in Cancer

Student mentoring

Each UROP paired with a PhD, MD/PhD or postdoc level mentor

Meeting with PI weekly

Presentation of research at the end

2-4 UROP students in 2019 summer

Can attend all lab activities. Other opportunities

Funding



NIH DIRECTOR'S
NEW INNOVATOR
AWARD

High-Risk, High-Reward
Research Symposium
June 6-8, 2018
DoubleTree Hotel, Bethesda, MD



AACR American Association
for Cancer Research
FOUNDATION FINDING CURES TOGETHER™



THE SONTAG FOUNDATION

Congressionally Directed Medical Research Programs

CDMRP



Melanoma
Research Alliance



THE MARY KAY
FOUNDATION



AMERICAN
CANCER
SOCIETY



DAMON
RUNYON
CANCER
RESEARCH
FOUNDATION

Sidi Chen Lab:

Matt Dong

Ryan Chow

John Park

Paul Renauer

Adan Codina

Chris Guzman

Guangchuan Wang

Lupeng Ye

Krista Chen

Johanna Shen

Zhigang Bai

QJ Yang

Li Zhang

Xiaoyun Dai

Youssef Errami

Hyunu Ray Kim

Xiaoya Zhang

Yaying Du

Jianjian Guo

Lyuyun Zhu, + Alumni



Yale University

The Evolutionary Systems Biology Lab @West Campus

Günter P. Wagner

Ecology and Evolutionary Biology

Obstetrics, Gynecology and Reproductive Sciences

Systems Biology Institute

Our Philosophy

- Nothing **Biology** makes sense except in the light of **Evolution**
 - T. Dobzhansky
- Nothing in **Evolution** makes sense except in the light of the rest of **Biology**.
 - James Griesemer
- Projects:
 - Evolutionary cell biology
 - Evolutionary biology of female reproduction
 - Evolutionary biology of cancer

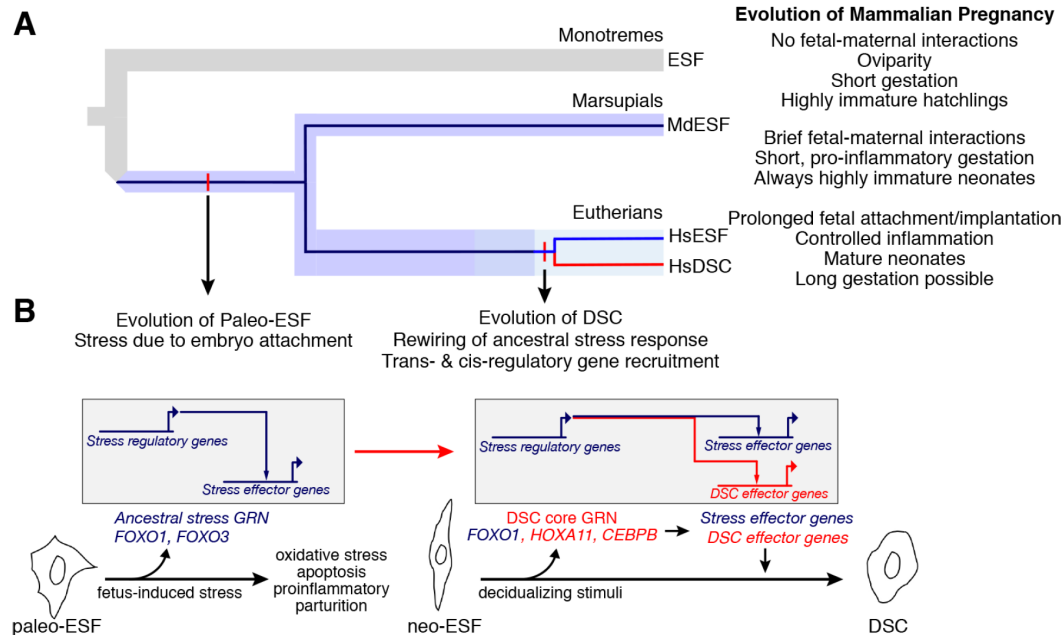
PLOS

BIOLOGY

2018

The mammalian decidual cell evolved from a cellular stress response

Eric M. Erkenbrack^{1,2}, Jamie D. Maziarz^{1,2}, Oliver W. Griffith^{1,2,3}, Cong Liang^{2,4}, Arun R. Chavan^{1,2}, Mauris C. Nnamani^{1,2}, Günter P. Wagner^{1,2,5,6*}



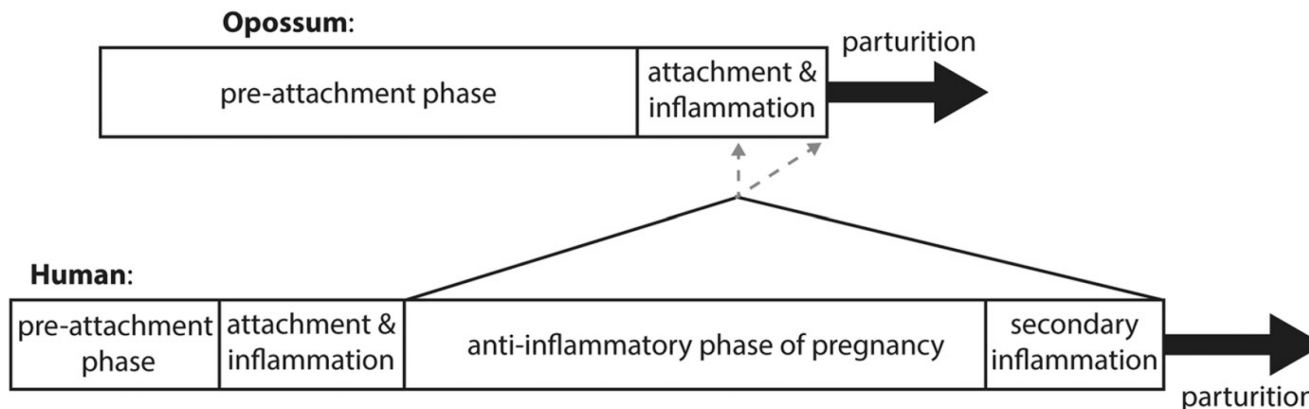
Eric Erkenbrack

Evolutionary biology of female reproduction

Embryo implantation evolved from an ancestral inflammatory attachment reaction

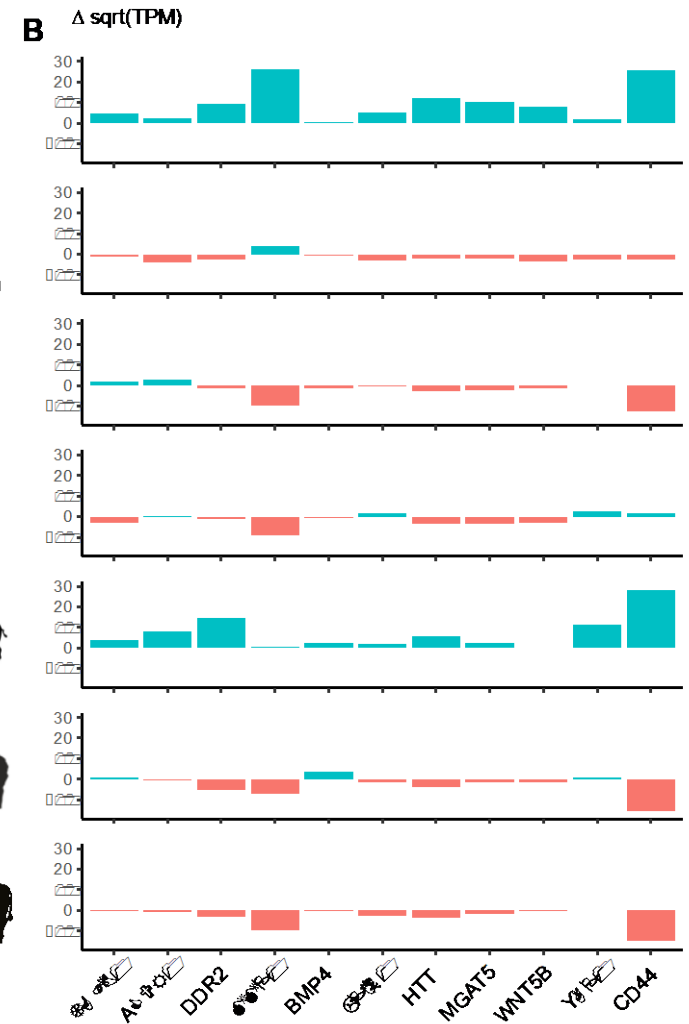
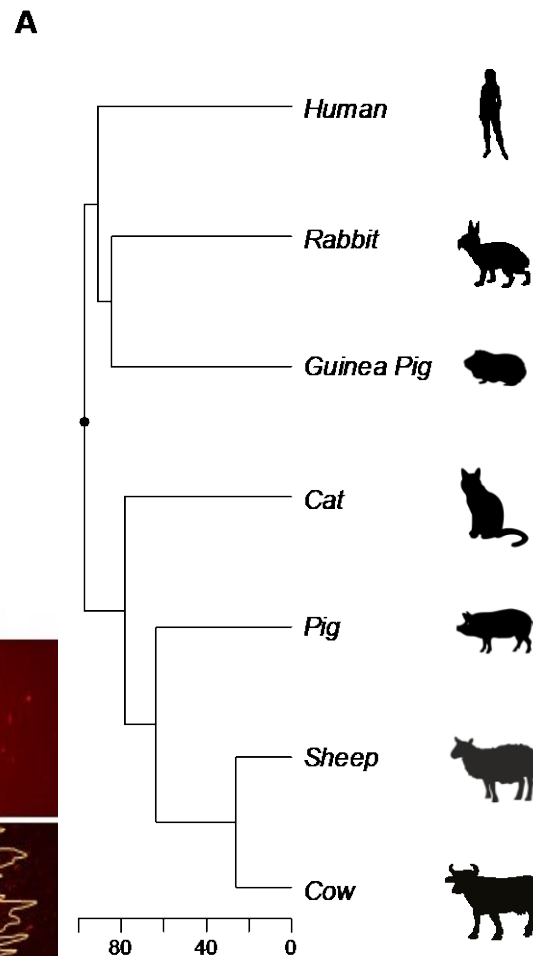
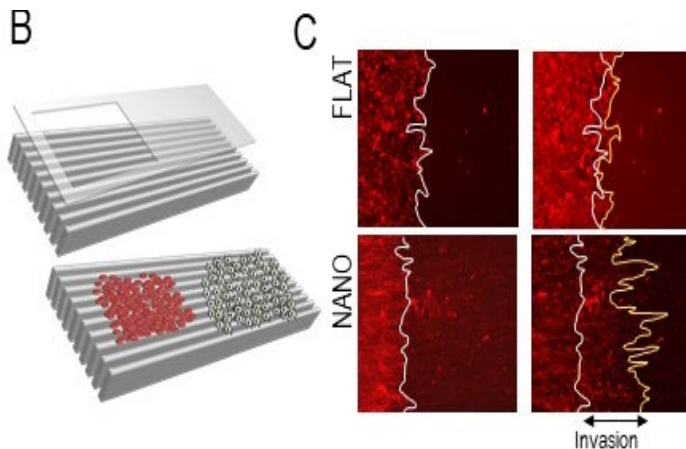
Oliver W. Griffith^{a,b,1}, Arun R. Chavan^{a,b}, Stella Protopapas^{a,b}, Jamie Maziarz^{a,b}, Roberto Romero^{c,d,e,f}, and Gunter P. Wagner^{a,b,g,h}

^aDepartment of Ecology and Evolutionary Biology, Yale University, New Haven, CT 06520; ^bYale Systems Biology Institute, Yale University, West Haven, CT 06516; ^cPerinatology Research Branch, Division of Obstetrics and Maternal-Fetal Medicine, Division of Intramural Research, Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Detroit, MI 48201; ^dDepartment of Obstetrics and Gynecology, University of Michigan, Ann Arbor, MI 48109; ^eDepartment of Epidemiology and Biostatistics, Michigan State University, East Lansing, MI 48824; ^fCenter for Molecular Medicine and Genetics, Wayne State University, Detroit, MI 48202; ^gDepartment of Obstetrics, Gynecology and Reproductive Sciences, Yale Medical School, New Haven, CT 06510; and ^hDepartment of Obstetrics and Gynecology, Wayne State University, Detroit, MI 48202



Oliver Griffith

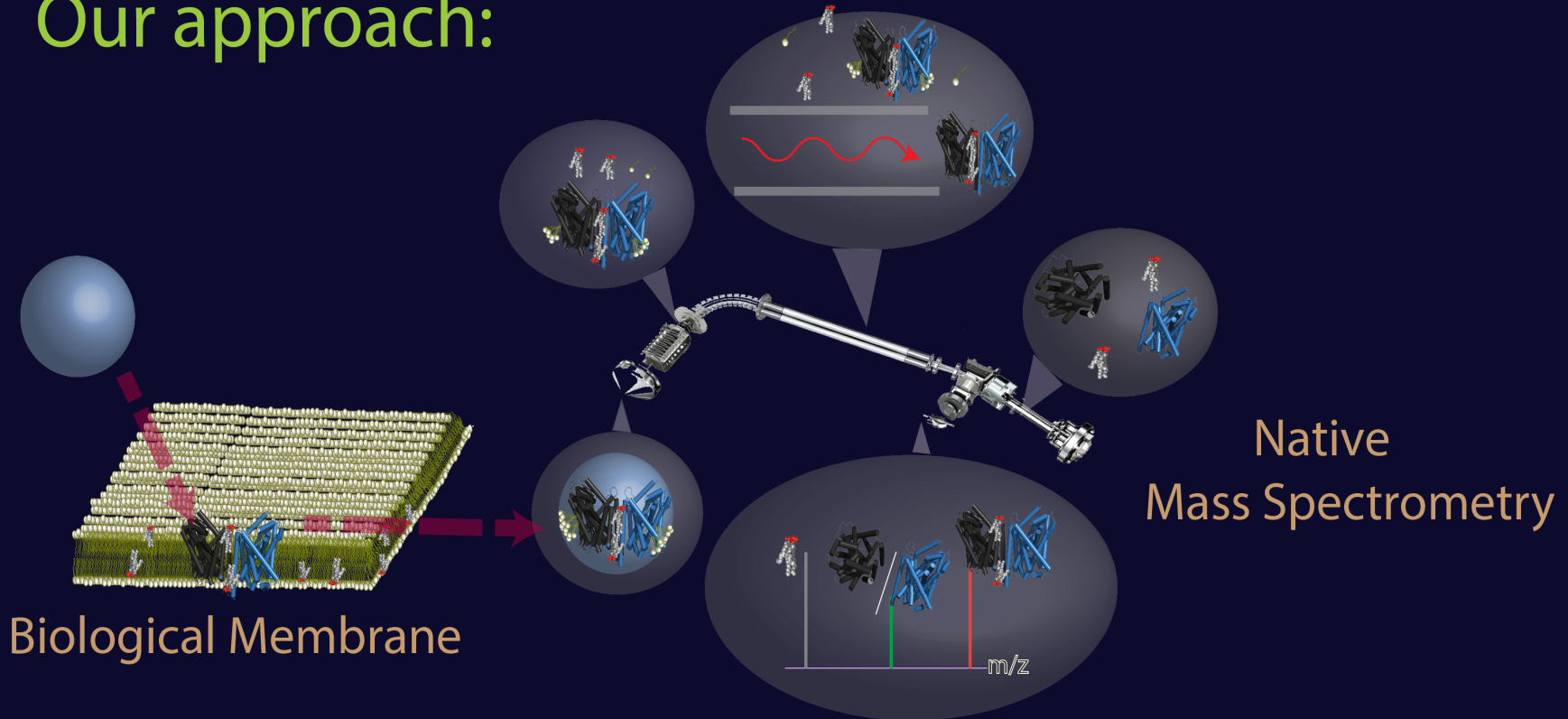
Evolutionary biology of cancer



Discovering membrane protein interactome through native mass spectrometry

Kallol Gupta (kallol.gupta@yale.edu)

Our approach:

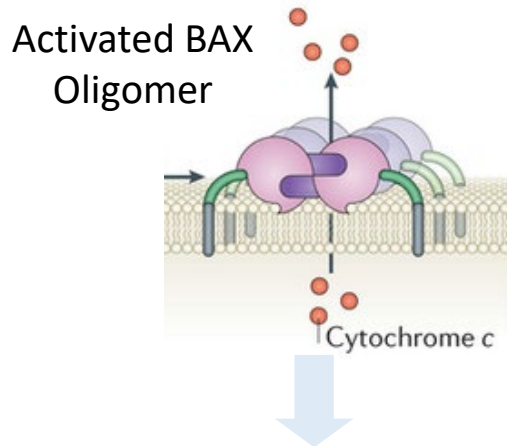


- (i) Direct identification of novel membrane complexes
- (ii) Spatio-temporal evolution of protein complexes through cell cycle
- (iii) Deconvoluting cellular signalling pathways - cellular pathogenesis
- (iv) Drug screening against native complexes

Why we care about membrane protein oligomerisation

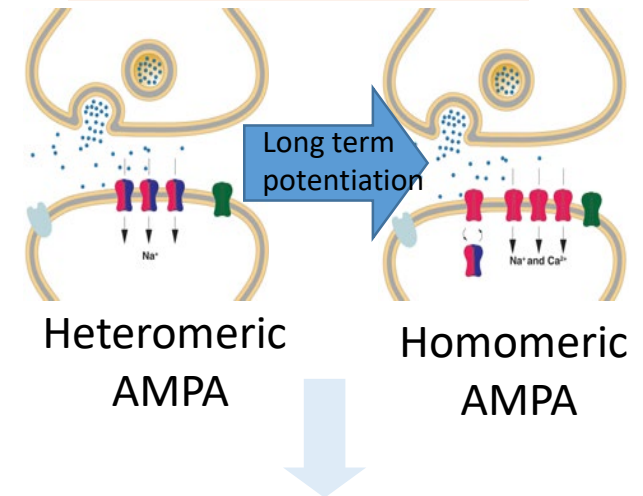
- A quarter of expressed genome codes for membrane proteins
- 60% marketed drugs target membrane proteins

- BAK/BAX: Cell death Apoptotic signaling



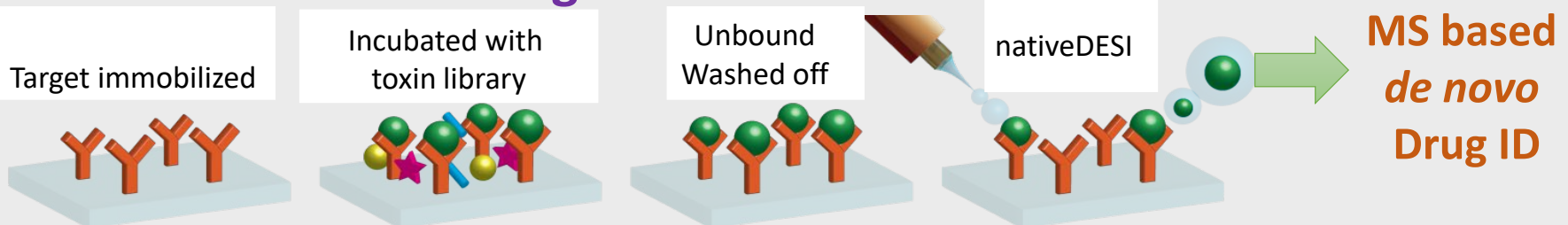
- Activators – Cancer
- Suppressors- Neurodegeneration

- Synaptic signaling



- Progression of memory
- Neurocognitive disorders

- nativeMS based drug screen



Acknowledgements

Anushka Halder

Fabian Giska

Joe Donlan

Stephen Ambrose

Jonathon Hopper, OMass Technologies

Kevin Giles (Waters Corp.)

Phill Stansfeld, Oxford

Gary Rudnik, Yale University

Moitrayee Bhattacharyya, UC Berkeley